

CORRESPONDENCE



Impact of stem cell source in HLA-matched hematopoietic stem cell transplant for acute lymphoblastic leukemia in first complete remission: a study from the GRAALL and the SFGM-TC

© The Author(s), under exclusive licence to Springer Nature Limited 2026

Bone Marrow Transplantation; <https://doi.org/10.1038/s41409-026-02837-w>

TO THE EDITOR:

In myeloablative HLA-matched hematopoietic stem cell transplantation (HSCT) peripheral blood stem cells (PBSC) increase the risk of chronic graft-versus-host disease (cGVHD) compared to bone marrow (BM) [1, 2]. While cGVHD is associated with substantial morbidity, it may also augment graft-versus-leukemia activity. An advantage of PBSC over BM in reducing the risk of relapse has been reported in some trials [1, 3]; however, this superiority has not been consistently demonstrated across all studies [1, 2, 4]. This discrepancy likely stems from heterogeneous study populations in term of malignancies, prognosis factors and donor types, making it difficult to isolate the true impact of stem cell source on relapse.

Historically, HSCT was considered the standard of care for treatment of acute lymphoblastic leukemia (ALL), in first complete remission (CR1). Over the past two decades, pediatric-inspired chemotherapy regimens have significantly improved outcomes in adult ALL patients [5–7], leading to a redefinition of the place of HSCT. In the GRAALL-2003 and GRAALL-2005 trials, HSCT in CR1 was offered only to adult ALL patients with high-risk (HR) features. This analysis re-evaluates the impact of PBSC versus BM in HR adults transplanted in CR1, focusing on whether PBSC offers an advantage over BM in reducing relapse risk in both matched related and matched unrelated transplants.

The GRAALL-2003 and GRAALL-2005 trials, conducted from November 2003 to March 2014 in 70 centers, offered HSCT to patients ≤55 years with Ph-negative ALL and at least one HR ALL factor (Supplementary Data) [5, 6, 8]. Conditioning was myeloablative up to 50 years, and BM was the recommended stem cell source. For this analysis, only patients receiving a myeloablative matched sibling donor (MSD) or matched unrelated donor (MUD) transplant with BM or PBSC were included; The cohort comprised 275 patients (184 with B-cell and 91 with T-cell ALL), with follow-up updated to November 2023. Patient characteristics and HSCT modalities are summarized in Supplementary Tables 1, 2. Median age at transplant was 32 years (IQR, 23–43), and 60% were male. Most patients ($n = 259$, 94%) received a total body irradiation (TBI)-based conditioning regimen. Donors were MSD in 154 transplants (56%; 118 BM and 36 PBSC) and MUD in 121 transplants (44%; 67 BM and 54 PBSC). Anti-thymocyte globulin (ATG) was administered to 37 patients, mainly in the MUD group (34/121, 28%; 16 BM and 18 PBSC), while only 3 MSD recipients received ATG. Outcomes for the entire cohort are described in the Supplemental results. Briefly, the median follow-up was of 10.5 years [95% CI, 10.1–11.1], 10-year estimated cumulative incidence of relapse (CIR) was 19% [15–25], 10-

year disease-free survival (DFS) and overall survival (OS) were 63% [95% CI, 57–69] and 66% [60–71], respectively. In multivariate analysis, including stem cell source (PBSC versus BM), type of donor (MUD versus MSD), age (≥45 years versus <45 years), use of TBI, and use of ATG, PBSC was not associated with a lower relapse risk but with an increased incidence of any-grade and extensive cGVHD (Supplementary Table 3). Transplantation from a MUD was also associated with a higher incidence of cGVHD. For subsequent analyses, the impact of stem cell source was evaluated separately in MSD and MUD transplant populations.

We first assessed the risk of relapse according to stem cell source in patients undergoing MSD and MUD transplant. In MSD transplant, the 10-year CIR was 17% [95% CI, 8–34] with PBSC versus 24% [17–33] with BM (SHR 0.69 [0.29–1.66], $p = 0.41$; Fig. 1a). In MUD transplants, the 10-year CIR was 11% [5–23] with PBSC versus 19% [12–31] with BM (SHR 0.54 [0.21–1.43], $p = 0.25$; Fig. 1b). The absence of a PBSC benefit on relapse risk in both MSD and MUD transplants was confirmed in multivariate analysis (Supplementary Table 4).

Another objective of the study was to assess the impact of stem cell source in MSD and MUD transplant on additional post-transplant outcomes, particularly cGVHD. PBSC use significantly increased the risk of any-grade cGVHD across both donor types. In MSD transplants, the 10-year incidence of cGVHD was 54% [95% CI, 39–71] after PBSC versus 29% [22–39] after BM (SHR 2.17 [1.25–3.76], $p = 0.006$). In MUD transplants, the 10-year incidence of cGVHD was 64% [51–77] after PBSC versus 40% [29–53] after BM (SHR 1.92 [1.14–3.22], $p = 0.01$; Fig. 1c, d). Compared to BM, PBSC was associated with an increased risk of extensive cGVHD in MUD transplants (32% [20–47] versus 8% [3–18]; SHR 4.92 [1.80–13.47], $p = 0.002$), but not in MSD transplants (SHR 1.87 [0.79–4.40], $p = 0.15$) (Fig. 1e, f). These incidences of cGVHD are lower than those reported in previous BM versus PBSC comparisons [1, 2], possibly due to the younger age of our cohort and to the use of ATG in some MUD transplants [2, 4, 9]. Multivariate analysis confirmed PBSC as an independent risk factor for any-grade cGVHD in both MSD (SHR 1.98 [1.18–3.33], $p = 0.01$) and MUD transplants (SHR 2.06 [1.18–3.60], $p = 0.01$), and for extensive cGVHD in MUD transplants (SHR 5.42 [1.97–14.93], $p = 0.001$). Moreover, in MUD transplants, PBSC use was associated with a higher risk of grade III–IV aGVHD (SHR 4.47 [1.62–12.34], $p = 0.004$), unlike in MSD transplants (Supplementary Table 4). The higher incidence of extensive cGVHD and grade III–IV aGVHD translated into higher NRM and poorer GRFS in the MUD PBSC setting compared with BM (SHR 2.88 [1.24–6.70], $p = 0.014$ for NRM and SHR 2.45 [1.46–4.09], $p < 0.001$ for GRFS) (Fig. 1g, h), whereas no significant differences in NRM or GRFS were observed in MSD transplants according to stem cell source (Fig. 1i, j). Stem cell source did not significantly impact OS or DFS (Fig. 1k, l).

Received: 11 December 2025 Revised: 26 January 2026 Accepted: 9 March 2026
Published online: 16 April 2026

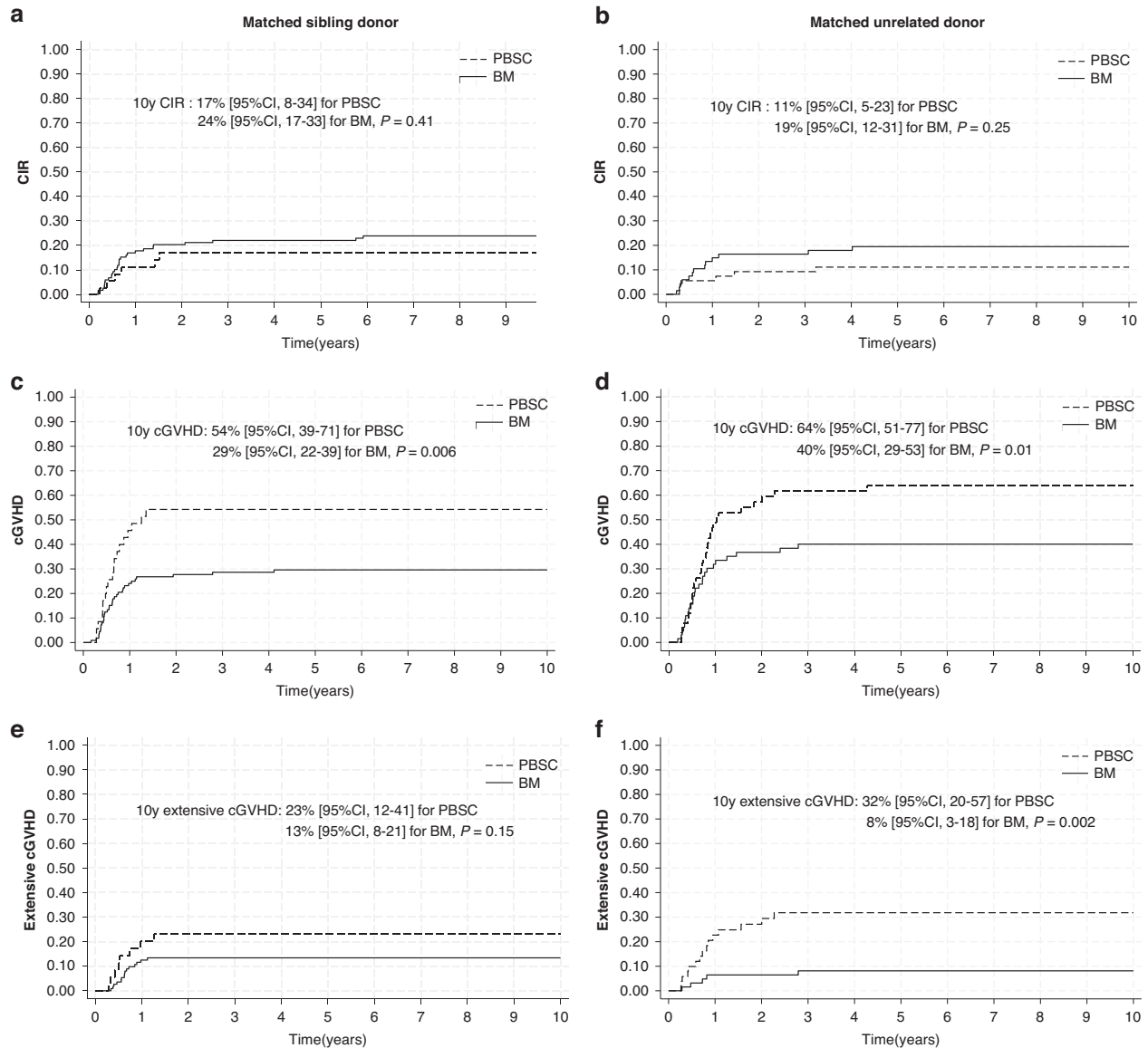


Fig. 1 Impact of stem cell source (PBSC versus BM) by donor type on outcomes. Panels **a, b** cumulative incidence of relapse (CIR); **c, d** cumulative incidence of any-grade chronic graft-versus-host disease (cGVHD); **e, f** cumulative incidence of extensive cGVHD; **g, h** non-relapse mortality (NRM); **i, j** grade III–IV aGVHD and/or extensive cGVHD-free, relapse-free survival (GRFS); **k, l** disease-free survival (DFS). Matched sibling donor (MSD) transplants are shown on the left, 10/10 HLA-matched unrelated donor (MUD) transplants on the right. Dashed lines represent peripheral blood stem cells (PBSC); solid lines represent bone marrow (BM). HR hazard ratio, IC confidence interval.

To our knowledge, this study is the first to specifically evaluate the impact of stem cell source in HR ALL adults transplanted in CR1. In this population, no benefit of PBSC over BM was observed in terms of relapse risk, whereas previous studies suggesting a relapse-protective effect of PBSC included only a minority of patients with ALL transplanted in CR1 [1, 3]. In the MSD setting, both BM and PBSC appear acceptable, as no significant differences were observed in extensive cGVHD, NRM, or relapse. However, in the MUD setting, BM may be preferred due to the higher incidence of extensive cGVHD and increased NRM associated with PBSC, without any corresponding benefit in relapse reduction. Our population is particular in that the majority of patients received BM stem cells in accordance with protocol recommendations, and only a minority received ATG. This practice is no longer standard, as current recommendations support the use of ATG in the MUD setting and with PBSC [10, 11]. While ATG effectively reduces the incidence of GVHD, this beneficial effect may be counterbalanced by an increased risk of infections and a potential reduction in the graft-versus-leukemia effect [12].

Although BM is no longer the stem cell source currently prioritized, partly for logistical reasons, our study allows a re-evaluation of its place in MUD transplantation for patients with ALL in CR1. This study also highlights the importance of reassessing stem cell source in light of individual disease characteristics and relapse risk. These findings warrant confirmation in other cohorts, particularly those incorporating ATG and the now widely used post-transplant cyclophosphamide for GVHD prophylaxis.

Matthieu Jestin¹, Sébastien Maury², Anne Huynh³, Edouard Forcade⁴, Micha Srouf⁵, Cristina Castilla-Llorente⁶, Jean-Baptiste Mear⁷, Sylvain Chantepie⁸, Dominik Schneidawind^{9,10}, Jakob Passweg^{10,11}, Mustafa Alani¹², Claude-Eric Bulabois¹³, Jérôme Cornillon¹⁴, Michael Loschi¹⁵, Jean-Valère Malfuson¹⁶, Eolia Brissot¹⁷, Pascal Turlure¹⁸, Xavier Poiré¹⁹, Carlos Graux²⁰, Nicole Raus²¹, Véronique Lhéritier²², Alban Villate²³, Etienne Daguindau²⁴,

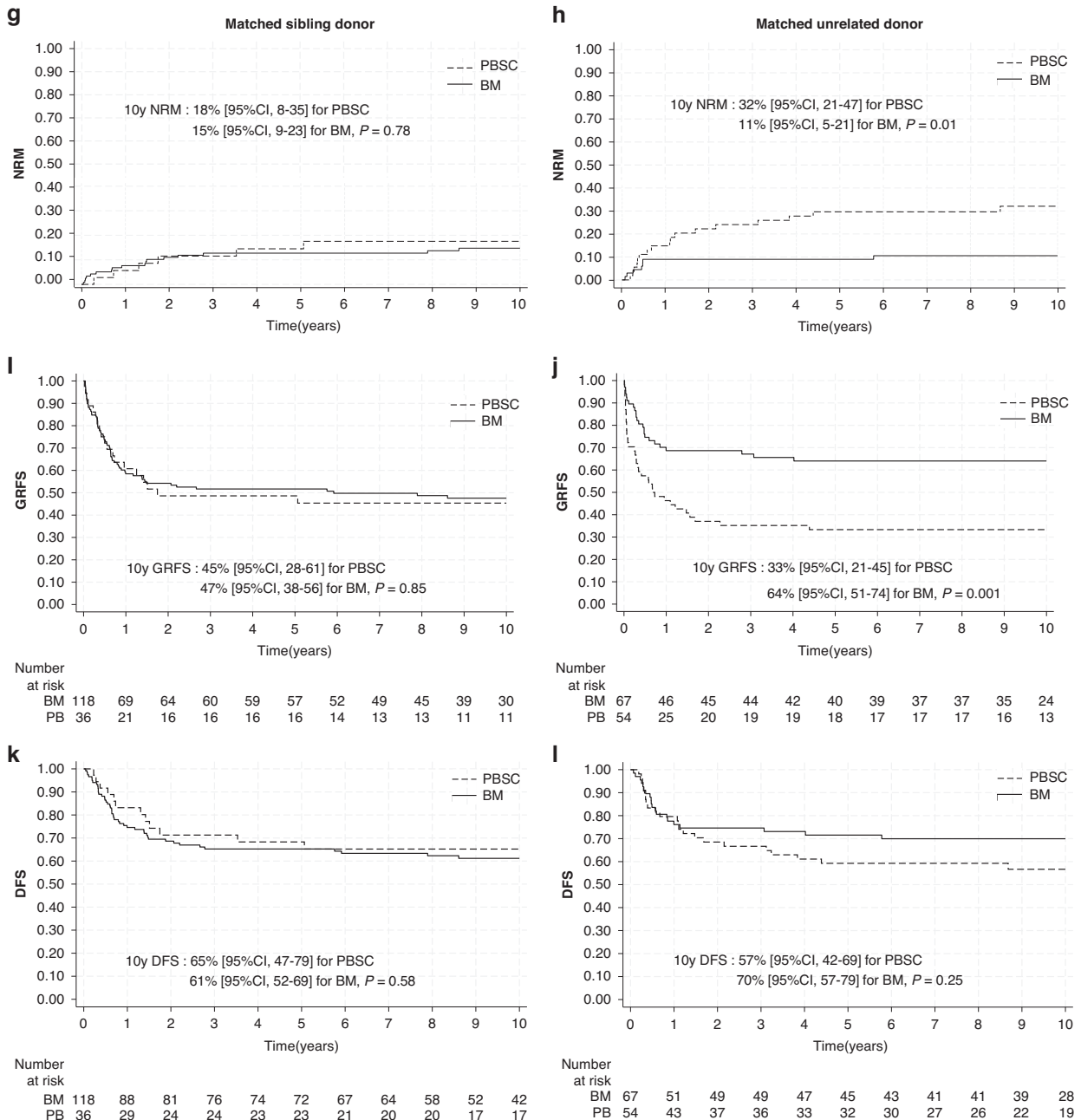


Fig. 1 Continue

Yves Chalandon^{10,25}, Karin Bilger²⁶, Gaelle Guillermin²⁷, Jacques-Olivier Bay²⁸, Clémence Loiseau²⁹, Natacha Maillard³⁰, Raynier Devillier³¹, Marie-Thérèse Rubio³², Franciane Paul³³, Sylvie François³⁴, Patrice Chevallier³⁵, Hélène Labussière-Wallet³⁶, Hervé Dombret³⁷, Stéphanie Nguyen³⁸, Nicolas Boissel^{1,39,40} and Nathalie Dhedin^{1,40}✉

¹Adolescent and Young Adult Hematology Department, St-Louis Hospital, APHP, Paris, France. ²Service d'Hématologie Clinique, Hôpitaux universitaires Henri-Mondor, AP-HP, Créteil, France. ³Service d'Hématologie, CHU Toulouse, IUCT Oncopole, Toulouse, France.

⁴Service d'Hématologie Clinique et Thérapie Cellulaire, CHU de Bordeaux, Bordeaux, France. ⁵CHU de Lille, University of Lille, INSERM U1286, Infnite, Lille, France. ⁶Institut de Cancérologie Gustave Roussy, Villejuif, France. ⁷Department of Hematology, L'Hôpital Pontchaillou, CHU de Rennes, Rennes, France. ⁸Institut d'Hématologie

de Basse Normandie, CHU de Caen, Paris, France. ⁹University Hospital Zurich, Zurich, Switzerland. ¹⁰Swiss Group for Clinical Cancer Research (SAKK), Bern, Switzerland. ¹¹Division of Hematology, University Hospital of Basel, Basel, Switzerland. ¹²Hematology Department, University Hospital of Rouen, Rouen, France. ¹³CHU Grenoble Alpes - Université Grenoble Alpes, Grenoble, France. ¹⁴Département d'Hématologie Clinique et de Thérapie Cellulaire, CHU de St-Etienne, Caen, France. ¹⁵Department of Hematology, Hôpital L'Archet, CHU de Nice, Nice, France. ¹⁶Department of Hematology, Hôpital d'Instruction des Armées, Percy, France. ¹⁷EBMT Unit, Sorbonne University, Saint-Antoine Hospital, AP-HP, INSERM UMRs 938, Paris, France. ¹⁸Department of Hematology, CHU de Limoges, Université de Limoges, Limoges, France. ¹⁹Cliniques Universitaires St. Luc, Brussels, Belgium. ²⁰CHU UCL Namur (Godinne), Yvoir, Belgium. ²¹Hôpital Lyon Sud, SFGM-TC, Lyon, France. ²²Hematology Department, Coordination Group for research on Adult Acute

Lymphoblastic Leukemia (GRAALL), HCL, Hôpital Lyon Sud, Pierre-Bénite, France.²³Hematology unit, Tours university hospital, Tours, France.²⁴Hopital Jean Minjot, Besancon, France.²⁵University Hospital Geneva, Hematology Service and Faculty of Medicine, University of Geneva, Geneva, Switzerland.²⁶CHRU de Strasbourg, Strasbourg, France.²⁷Service d'Hématologie et d'Hémostase Clinique, CHU de Brest, Brest, France.²⁸CHU Estaing, Clermont-Ferrand, France.²⁹Haematology Department, Necker-Enfants Malades Hospital, AP-HP, Université Paris Cité, Paris, France.³⁰Department of Hematology, CHU de Poitiers, Poitiers, France.³¹Institut Paoli-Calmettes, Marseille, France.³²CHRU de Nancy-Brabois, Nancy, France.³³CHU de Montpellier, Montpellier, France.³⁴Maladies du Sang, CHU Angers, Angers, France.³⁵CHU Nantes, Nantes, France.³⁶Hopital Lyon Sud, Hospices Civils de Lyon, Pierre Bénite, Paris, France.³⁷Department of Hematology, Saint Louis Hospital, AP-HP, Paris, France.³⁸Hôpital de la Pitié Salpêtrière, Paris, France.³⁹UMR 1342, Saint-Louis Research Institute, Université Paris Cité, Inserm, Paris, France.⁴⁰These authors contributed equally: Nicolas Boissel, Nathalie Dhédin. ✉email: nathalie.dhedin@aphp.fr

REFERENCES

1. Stem Cell Trialists' Collaborative Group. Allogeneic peripheral blood stem-cell compared with bone marrow transplantation in the management of hematologic malignancies: an individual patient data meta-analysis of nine randomized trials. *J Clin Oncol*. 2005;23:5074–87.
2. Anasetti C, Logan BR, Lee SJ, Waller EK, Weisdorf DJ, Wingard JR, et al. Peripheral blood stem cells versus bone marrow from unrelated donors. *N Engl J Med*. 2012;367:1487–96.
3. Bensinger WJ, Martin PJ, Storer B, Clift R, Forman SJ, Negrin R, et al. Transplantation of bone marrow as compared with peripheral-blood cells from HLA-identical relatives in patients with hematologic cancers. *N Engl J Med*. 2001;344:175–81.
4. Schmitz N, Beksac M, Hasenclever D, Bacigalupo A, Ruutu T, Nagler A, et al. Transplantation of mobilized peripheral blood cells to HLA-identical siblings with standard-risk leukemia. *Blood*. 2002;100:761–7.
5. Huguet F, Leguay T, Raffoux E, Thomas X, Beldjord K, Delabesse E, et al. Pediatric-inspired therapy in adults with philadelphia chromosome-negative acute lymphoblastic leukemia: the GRAALL-2003 study. *J Clin Oncol*. 2009;27:911–8.
6. Huguet F, Chevret S, Leguay T, Thomas X, Boissel N, Escoffre-Barbe M, et al. Intensified therapy of acute lymphoblastic leukemia in adults: report of the randomized GRAALL-2005 clinical trial. *J Clin Oncol*. 2018;36:2514–23.
7. DeAngelo DJ, Stevenson KE, Dahlberg SE, Silverman LB, Couban S, Supko JG, et al. Long-term outcome of a pediatric-inspired regimen used for adults aged 18–50 years with newly diagnosed acute lymphoblastic leukemia. *Leukemia*. 2015;29:526–34.
8. Dhédin N, Huynh A, Maury S, Tabrizi R, Beldjord K, Asnafi V, et al. Role of allogeneic stem cell transplantation in adult patients with Ph-negative acute lymphoblastic leukemia. *Blood*. 2015;125:2486–96.
9. Morton J, Hutchins C, Durrant S. Granulocyte-colony-stimulating factor (G-CSF)-primed allogeneic bone marrow: significantly less graft-versus-host disease and comparable engraftment to G-CSF-mobilized peripheral blood stem cells. *Blood*. 2001;98:3186–91.
10. Soiffer RJ, Kim HT, McGuirk J, Horwitz ME, Johnston L, Patnaik MM, et al. Prospective, randomized, double-blind, phase III clinical trial of anti-T-lymphocyte globulin to assess impact on chronic graft-versus-host disease-free survival in patients undergoing HLA-matched unrelated myeloablative hematopoietic cell transplantation. *J Clin Oncol Off J Am Soc Clin Oncol*. 2017;35:4003–11.
11. Kröger N, Solano C, Wolschke C, Bandini G, Patriarca F, Pini M, et al. Antilymphocyte globulin for prevention of chronic graft-versus-host disease. *N Engl J Med*. 2016;374:43–53.
12. Storek J, Mohty M, Boelens JJ. Rabbit anti-T cell globulin in allogeneic hematopoietic cell transplantation. *Biol Blood Marrow Transplant*. 2015;21:959–70.

AUTHOR CONTRIBUTIONS

MJ analyzed data, draw the figures and wrote the paper. MR and VL collected clinical and biological annotations on patients. NB performed statistical analysis. NB and ND conceived the study and critically reviewed the article. SM, AH, EF, MS, CCL, JBM, SC, DS, JP, MA, CEB, JC, ML, JVM, EB, PT, XP, AV, ED, YC, KB, GG, JOB, CL, NM, RD, MTR, FP, SF, PC, HLW, SN and ND included patients. All authors revised and approved the manuscript.

COMPETING INTERESTS

The authors declare no competing interests.

ADDITIONAL INFORMATION

Supplementary information The online version contains supplementary material available at <https://doi.org/10.1038/s41409-026-02837-w>.

Correspondence and requests for materials should be addressed to Nathalie Dhédin.

Reprints and permission information is available at <http://www.nature.com/reprints>

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.