

# A modeling approach to determine the importance of dynamic regulation of plant hydraulic conductivities on the water uptake dynamics in the soil-plant-atmosphere system

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## Abstract

We present here a new model, PlaNet-Maize, with the purpose of investigating the effect of environmental and endogenous factors on the growth and water relations of the maize plant. This functional-structural plant model (FSPM) encompasses the entire soil-plant-atmosphere continuum with a sub-organ resolution. The model simulates the growth and development of an individual maize plant and the flux of water through the plant structure, from the rhizosphere to the leaf boundary layer. Leaf stomatal conductance and root radial and axial conductivities are considered as functions of local water potential. Finally, a simple carbon allocation rule is included in the model to allow the feedback effect of water deficit on plant growth. The model was successfully used to reproduce experimental plant hydraulic behavior in

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response to water deficit. The quantitative contribution of leaf conductance and root conductivities were assessed individually and in combination.

Our results highlight the importance of regulating hydraulic properties in FSPM as these can strongly modify the water uptake dynamics and lead to emerging water uptake behaviors. The modeling results also indicate that plant hydraulic properties can theoretically be tailored to improve plant water use in challenging environments.

*Keywords:*

maize, water uptake dynamics, functional-structural plant model

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## 1. Introduction

The flow of water in the soil-plant-atmosphere continuum is a passive process driven by water potential differences and enabled by the continuity of the liquid phase of water between the soil and the leaf mesophyll (Steudle, 2001). For a given driving force, the flow of water in the plant is set primarily, though not exclusively, by the radial conductance of the root system, the axial conductivity of the xylem and the leaf conductance.

The leaf conductance, which is largely determined by the density and aperture of the stomata, sets the maximum amount of water that can be transpired under a given environmental demand. Due to the obvious and major role of stomata, the mechanisms underlying their regulation have received much attention from the scientific community, leading to numerous

14 experimental and modeling approaches (reviewed by Damour et al. (2010)).

15     The root radial conductance is a complex property reflecting the compos-  
16 ite nature of water movement from the root surface to the xylem. It both  
17 depends on irreversible processes like the deposition of hydrophobic barriers  
18 in the endodermis and exodermis (Enstone et al., 2003), and on time-varying  
19 features like the activity of water channels (Maurel et al., 2008; Javot and  
20 Maurel, 2002). The crossing of root tissues is generally considered as an  
21 important limiting step in the water uptake process (Steudle and Peterson,  
22 1998).

23     The axial conductivity of the xylem vessels is also a complex property  
24 which combines the structural features of the xylem, the dynamics of cavi-  
25 tation events occurring when the tension in the xylem is too large (Sperry  
26 et al., 1998) and the extent of embolized vessels refilling that occurs when  
27 the tension vanishes but possibly also under tension (McCully et al., 1998;  
28 Zwieniecki and Holbrook, 2009; Holbrook and Zwieniecki, 1999).

29     While the leaf conductance sets the amount of water extracted by the  
30 plant, the distribution of root axial conductivities and radial conductances  
31 throughout the root system (the *hydraulic architecture*) determines the sites  
32 of water uptake in the soil. This role of root hydraulic properties has often  
33 been considered as minor in comparison with root length density, but this  
34 should be reconsidered in view of recent data (Draye et al., 2010). Therefore,  
35 considering leaf, root and environmental factors simultaneously may be an  
36 important contribution if we aim to tailor plants with improved resistance  
37 to water deficit.

38

39 For many years, scientists have developed computer models to simulate  
40 how biological processes, described individually or at an organ scale, integrate  
41 and scale up in the soil-plant-atmosphere system. On the root side, models  
42 evolved from architectural models reproducing the root system shape and  
43 growth (Diggle, 1988; Pagès et al., 1989; Lynch et al., 1997) to functional-  
44 structural models simulating physical, chemical or physiological processes  
45 such as nutrient acquisition (Ge et al., 2000), carbon allocation (Bidel et al.,  
46 2000) or water uptake (Somma et al., 1998; Javaux et al., 2008). A similar  
47 trend was observed for shoot models (Fournier and Andrieu, 1998). While  
48 some models address simultaneously the root and shoot systems (Drouet and  
49 Pagès, 2003; Janott et al., 2011), the vast majority focus on either part of the  
50 plant and use simplified descriptions of the other. As of today, we are not  
51 aware of any model that simulates the water dynamics of a complete plant  
52 with a detailed description of each organ.

53

54 This situation led us to develop the PlaNet-Maize model, which simulates  
55 at a sub-organ resolution the growth and architecture, the water flows and  
56 the main hydraulic regulations of a whole maize plant. The model uses sim-  
57 plified rules for the production and allocation of assimilates to the different  
58 organs.

59

## 60 **2. Model description**

### 61 *2.1. General principles*

62     PlaNet-Maize adheres to the principles of the metal-model PlaNet (Plant  
63 as a Network), developed by Loïc Pagès, that defines a building schema for  
64 the creation of various whole plant models. In PlaNet, the plant structure is  
65 viewed as a network of articles interconnected in a tree-like structure and lo-  
66 calized in space. Articles are typically organs, but can be defined as smaller  
67 or larger entities. They have three fundamental behaviors: (1) they can  
68 grow and create new articles (morpho-generator), (2) they have their own  
69 metabolic activity (bio-reactor) and (3) they can transport substances from  
70 and to neighboring articles and environment (carrier). As articles are local-  
71 ized in space and in the network topology, these behaviors can be dependent  
72 on local environment information as well as on network-derived information.

73  
74     Two types of articles are typically considered in PlaNet. The first are  
75 the segments, which make up the structure of the plant. The second are  
76 the meristems, which generate new segments and / or meristems and ensure  
77 growth and branching. These two types of articles can be sub-divided based  
78 on their botanical nature: stem, leaf or root. These six types of articles are  
79 sufficient to simulate the growth and development of a whole plant during  
80 the vegetative stage (fig. 1).

81  
82     The PlaNet-Maize model uses PlaNet rules to simulate the growth and  
83 structure of a maize plant and includes modules specific to water movement  
84 and hydraulic regulatory processes. PlaNet-Maize was developed in Java and

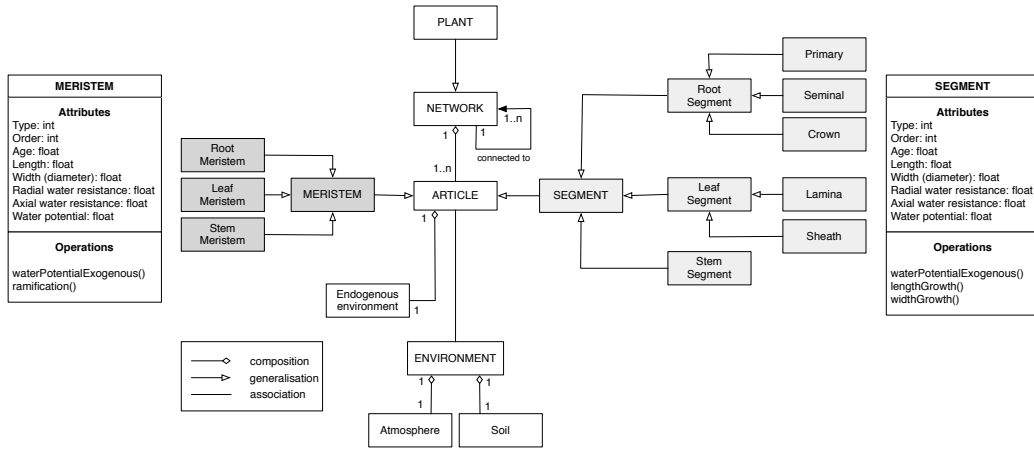


Figure 1: PlaNet-Maize UML scheme. Light grey boxes represent the segments classes. Dark grey boxes represent the meristem classes.

is integrated in the modeling platform CrossTalk (Draye and Pagès, 2006) that enables the coupling with environment models, the 3D visualization and the *in situ* interactive modification of the plant during a simulation (e.g. pruning). The CrossTalk platform allows users to easily modify a simulation parameters such as the initial environmental condition or the simulation time-step (in hours).

## 2.2. Architecture module

### 2.2.1. Root system

The root system of PlaNet-Maize comprises four types of roots: the primary root (first embryonic root), the seminal roots (embryonic roots initiated from the scutelar node), the crown roots (shoot-born roots) and the first and second order lateral roots (Hochholdinger et al., 2004). Its implementa-

tion was inspired from previous root architecture models (Pagès et al., 2004; Drouet and Pagès, 2003) with the distinction that root axes in PlaNet-Maize originate from different types of organs (fig. 2.B). The primary and seminal roots arise from the seed while crown root meristems are initiated by the stem meristem. It follows that there is no single connection between the root system and the shoot and that preferential water flow may occur between crown roots and the leaf initiated at the same node. Root elongation rate was computed as a function of the root meristem diameter (Drouet and Pagès, 2003; Mollier and Pellerin, 1999).

107

### 108 *2.2.2. Shoot system*

The shoot architectural module of PlaNet-Maize was largely inspired from the model GRAAL where leaf and stem growth and development are determined by morphogenetic processes and are functions of thermal time (Drouet and Pagès, 2003). Similarly as for the roots, leaves are made of successive segment articles. Every article is labeled as part of the sheath or the lamina. The leaf meristem is located at the junction between the sheath and the lamina (fig. 2.B).

### 116 *2.3. Carbon module*

A carbon module was implemented in PlaNet-Maize to allow various feedbacks between plant growth and water dynamics. For the sake of simplicity, the carbon module was not implemented at article but at plant level. The module was divided into three processes: production, demand and allocation of assimilates.

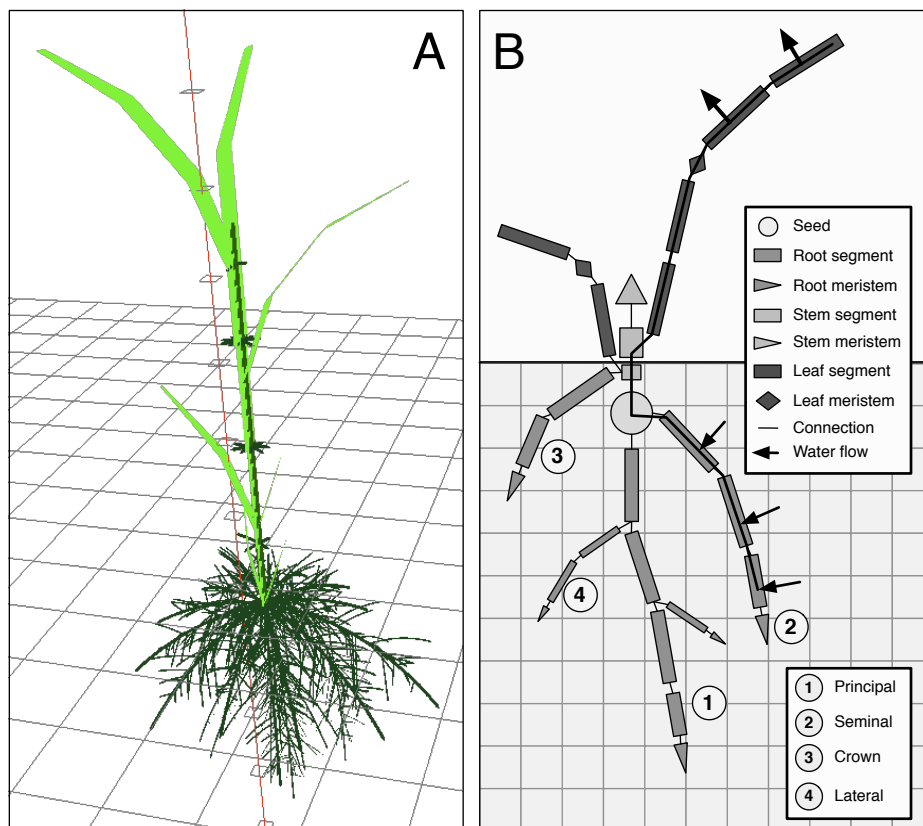


Figure 2: PlaNet-Maize representations. A. PlaNet-Maize visual output in CrossTalk. B. Schematic representation of the PlaNet-Maize structure.

### 122 2.3.1. Production

123 During the first steps of the simulation, the seed acts as a carbon source  
 124 and its supply is computed at each time step according to the seed dry weight  
 125 and a seed supply rate factor. When leaves start transpiring, the amount of  
 126 carbon produced at each time step ( $C_{av}$ ) is defined by the total amount of  
 127 water transpired by the plant as in the model presented by Somma et al.  
 128 (1998) and considering instantaneous water use efficiency values between 3.5  
 129 and 9.5 mg $_{CO_2}$ /g  $H_2O$  (Allen et al., 2011). Finally, at each time step, exceed-  
 130 ing carbon (see below) is stored and can be further released as a function of  
 131 the total amount of C available at the plant level and a reserve supply rate  
 132 (Drouet and Pagès, 2003).

### 133 2.3.2. Demand

134 Carbon demand is divided into maintenance and growth demands. The  
 135 maintenance demand is computed for every plant article such as:

$$C_m = mMQ10^{(T-T_{ref})/10}\Delta t \quad (1)$$

136 where  $C_m$  is the carbon maintenance demand [g $_{CO_2}$ ],  $M$  is the organ dry mass  
 137 [g],  $m$  the specific maintenance cost [g $_{CO_2}$ .g $^{-1}$ DM],  $Q10$  is a temperature co-  
 138 efficient [-],  $T$  is the environmental temperature [°C],  $T_{ref}$  is the temperature  
 139 above which maintenance cost is more than proportional to the organ dry  
 140 mass [°C] and  $\Delta t$  is the length of the time step [h] (Drouet and Pagès, 2003).

141

142 Similarly, the growth demand is defined for every article following the  
 143 equation:

$$C_g = \frac{G_p}{g} \Delta t \quad (2)$$

144 where  $C_g$  is the carbon growth demand [ $g_{CO_2}/h$ ],  $G_p$  is the potential growth  
 145 in dry mass [ $g_{CO_2}$ ],  $g$  is a growth conversion coefficient [-] and  $\Delta t$  is the length  
 146 of the time step [h] (Drouet and Pagès, 2003). The dry mass of every article  
 147 is a function of its volume and density, depending on the article age (Drouet  
 148 and Pagès, 2003, 2007).

### 149 2.3.3. Allocation

150 The allocation of the available carbon ( $C_{av}$ ) between all plant articles  
 151 follows precise priority rules.  $C_{av}$  is used in priority to meet the whole plant  
 152 maintenance demand, considered as an obligatory cost (Drouet and Pagès,  
 153 2003; Postma and Lynch, 2011). The remaining C:

$$C_{avg} = C_{av} - C_m, \quad (3)$$

154 is divided between the root and the shoot according to a root-to-shoot C  
 155 allocation ratio (0.6, Somma et al. (1998)). In the shoot, priority is given to  
 156 the leaf growth over the stem, while in the roots, priority is defined based  
 157 on the potential growth rate of each root (*sink term*). Once this allocation  
 158 is performed, a growth satisfaction coefficient is computed for every article  
 159 ( $G_p \cdot C_{avg}^{-1}$ , between 0 and 1). This coefficient will be used (1) to define the  
 160 actual growth of the different articles and (2) to define the initial diameter  
 161 of newly created root meristems. As in the GRAAL model, if this diameter  
 162 falls under a critical minimum value (0.14 cm, Drouet and Pagès (2003)), the  
 163 meristem is not initiated.

## 164 2.4. Water fluxes module

### 165 2.4.1. General principle

166 The network of articles intervening in the uptake and transport of water  
 167 in the plant can be conveniently analyzed using the analogy with an electric  
 168 circuit (Landsberg and Fowkes, 1978). In particular, radial water fluxes can  
 169 be described using the equation:

$$J_r(z) = L_r[\psi_e(z) - \psi_x(z)]S \quad (4)$$

170 where  $J_r(z)$  is the radial flux through root tissues or stomata [ $\text{m}^3.\text{s}^{-1}$ ],  $\psi_x(z)$   
 171 and  $\psi_e(z)$  are respectively the water potential of the article (viz. xylem) and  
 172 of the environment [MPa],  $L_r$  is the article radial conductivity [ $\text{m}.\text{s}^{-1}.\text{MPa}^{-1}$ ]  
 173 and  $S$  is the article surface [ $\text{m}^2$ ]. Similarly, the axial water flux can be  
 174 described as follow:

$$J_h(z) = -K_h \frac{\Delta\psi_x(z)}{\Delta z} \quad (5)$$

175 where  $J_h(z)$  is the axial flux between two articles [ $\text{m}^3.\text{s}^{-1}$ ],  $-K_h$  is the xylem  
 176 conductance [ $\text{m}^4.\text{s}^{-1}.\text{MPa}^{-1}$ ],  $\Delta\psi_x(z)$  is the water potential difference be-  
 177 tween the articles and  $\Delta z$  is the distance between these articles [m]. Finally,  
 178 the Kirchhoff law, which states that *in* fluxes must equal *out* fluxes, is ap-  
 179 plied to every article:

180

$$\sum^k I_{kin} = \sum^k I_{kout} \quad (6)$$

181 The combination of these three equations has been successfully used to  
182 simulate water transfers in the soil-plant system in different models (Doussan  
183 et al., 2006; Javaux et al., 2008). In these models, at each time step, the  
184 equations are specified for every root segment and are assembled as a system  
185 of linear equations that is solvable for all articles that comprise the root  
186 system. In PlaNet-Maize, we generalized this formulation in order to apply  
187 it to the whole plant level (roots, stem and leaves).

#### 188 2.4.2. *Soil water depletion*

189 The soil in PlaNet-Maize is implemented as a 3D grid of voxels with a  
190 mesh size of typically 1 cm<sup>3</sup>. Soil water depletion is simulated by removing  
191 the uptake of every root article from its enclosing voxel. The relationship  
192 between the soil water content and the soil water potential ( $\theta$ -h curve) is  
193 defined using a Mualen-van Genuchten equation (van Genuchten, 1980) and  
194 based on experimental measurement (data not shown).

195

196 It has to be noted that the current implementation of water fluxes in the  
197 soil 3D domain does not follow Richard's equation (unlike traditional soil  
198 models like HYDRUS). In PlaNet-Maize, horizontal redistribution of water  
199 in the bulk soil is obtained by the application of a mean smoothing operator  
200 on every soil element and its 4 horizontal neighbors. This operator may  
201 be modified to simulate different soil properties. Despite the fact that this  
202 method lacks physical basis, it has the advantage to mimic basic soil water  
203 movement at a minimum computational cost.

### 204 2.4.3. Hydraulic parameters

205 To the exception of the stomatal conductance, most of the conductances  
206 used in the water flux module remain poorly estimated experimentally. In  
207 addition, some of these conductances are expressed in different units. In  
208 particular, the root radial conductivity is expressed in  $[\text{m.s}^{-1}.\text{MPa}^{-1}]$  while  
209 the leaf radial conductivity ( $g_s$ ) is usually expressed in  $[\text{m}^3.\text{m}^{-2}.\text{s}^{-1}]$  ), which  
210 lead us to make some approximation in the model.

211 *Root.* The root hydraulic properties described by Doussan et al. (1998) were  
212 used in this study. These properties (axial and radial) are functions of the  
213 root segment age and type (axis or lateral). These values have been used  
214 successfully in previous models (Doussan et al., 2006; Javaux et al., 2008).

215 *Stem.* The stem radial conductivity was set to zero, assuming that the stem  
216 is not permeable to water. Its axial conductivity was calculated according to  
217 the Hagen-Poiseuille law,

$$K = \frac{\Pi r^4}{8\eta} \quad (7)$$

218 where  $K$  is the axial conductivity,  $r$  the radius of the xylem pipes and  $\eta$   
219 the water dynamic viscosity ( $= 1.002 \text{ mPa.s}$ ). Xylem radius was estimated  
220 to be 1.7% of the stem radius (Li et al., 2009). In our case, for the sake of  
221 simplicity, xylem vessels were considered as a single pipe.

222 *Leaves.* As few quantitative data about the axial conductivity of the leaf were  
223 found in the literature, we set it to a tenth of the stem axial conductivity  
224 (Wei et al., 1999). As mentioned above, leaf radial conductivity values are  
225 generally based on the difference in relative humidity between the stomatal

cavity and the atmosphere and do not fit the water flux resolution in PlaNet-Maize that are based on on water potential differences. Therefore, we used the flux equation

$$L_r = \frac{\Delta\psi S}{J_{out}} \quad (8)$$

where  $\Delta\psi$  is the water potential difference between the atmosphere and the leaf [MPa],  $S$  is the leaf area [cm<sup>2</sup>] and  $J_{out}$  the radial water flux [m.s<sup>-1</sup>], with experimental data on water potential and flux (not shown), and could estimate the maximal leaf stomatal conductance to 3x10<sup>9</sup>m.s<sup>-1</sup>.MPa<sup>-1</sup>.

#### 2.4.4. Hydraulic regulation of the plant

In order to investigate the contribution of varying hydraulic properties to the regulation of the water status of the plant, we introduced modifier functions that reduce the radial conductance of roots, the axial conductivity of the xylem (roots, stem and leaf) and the leaf radial conductance. These modifiers were calculated locally for each article as a function of their (xylem) water potential.

*Leaf conductance ( $g_s$ ).* As stomata, which ranges from fully open to completely closed, ultimately define the transpiration demand, their regulation is therefore seen as the master control site in the transpiration flow. In PlaNet-Maize, stomatal closure is influenced by exogenous (light, temperature and vapor pressure deficit) and endogenous (leaf water potential) factors. Diurnal variations in the leaf stomatal conductivity due to exogenous factors were implemented following the Jarvis equation (Jarvis, 1976):

$$g_s = g_{smax} * F_1(PAR) * F_2(T_a) * F_3(VPD) \quad (9)$$

where  $g_s$  is the leaf stomatal conductance [ $\text{mm.s}^{-1}$ ] and  $g_{smax}$  is the maximum stomatal conductance of individual leaves under optimal conditions [ $\text{mm.s}^{-1}$ ].  $F_1(PAR)$ ,  $F_2(T_a)$  and  $F_3(VPD)$  are environmental stress functions, with  $0 \leq F_i(X) \leq 1$ . For the sake of simplicity, light interception was not computed as a function of leaf positioning and mutual shading.

252

The influence of the leaf water potential on the leaf radial conductivity (fig. 3.A) was implemented in PlaNet-Maize using a Weibull function (Janott et al., 2011; Bohrer et al., 2005) following data from Cochard (2002):

$$f(\psi) = 1 - e^{-(-\frac{\psi}{b})^c} \quad (10)$$

where  $b$  and  $c$  are curve parameters and  $\psi$  is the article water potential.

*Root radial conductivity ( $K_r$ ).* The dynamic regulation of the root radial conductivity is mainly performed through the regulation of aquaporin activity (Maurel et al., 2008; Javot and Maurel, 2002), although quantitative data linking aquaporin activity with the root radial conductivity are generally not available. Aquaporin activity is down regulated during water deficit, yet it is not clear to what extent aquaporin activity influences  $K_r$  (Bramley et al., 2009). Therefore, the regulation of root radial conductivity was implemented in PlaNet-Maize following a logistic function bounded between 1 and 0.4, considering that aquaporin regulation cannot reduce the root radial conductivity to zero (fig. 3.A).

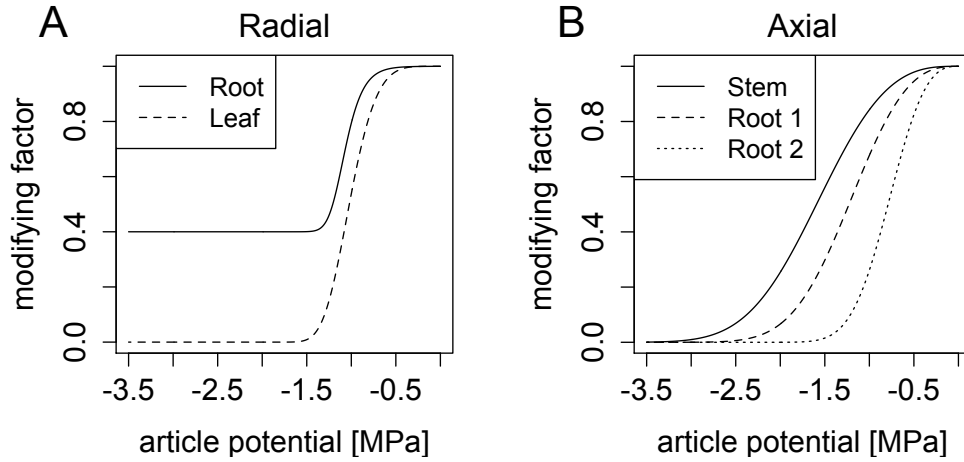


Figure 3: PlaNet-Maize conductivity modifiers. A. Modifiers for root/leaf radial conductivities. B. Modifiers for the axial conductance. PR = first order root. LR = lateral root.

267 *Axial conductance* ( $K_x$ ). Changes of axial conductance occur primarily un-  
 268 der the influence of xylem cavitation and embolism repair (Peirce, 1936).  
 269 The relationship between the tension in the xylem and the decrease in axial  
 270 conductance is commonly described using Weibull functions (Sperry et al.,  
 271 1988; Tyree and Sperry, 1989) (see eq. 10). In PlaNet-Maize, the Weibull  
 272 function was implemented in every article according to data from Li et al.  
 273 (2009) and considering different curves for the different plant organs (stem  
 274 > first order roots > second order roots, fig. 3.B, Hacke et al. (2000a)).

### 275 3. Method

#### 276 3.1. Conditions for simulations

277 The aim of the simulations presented here was to assess the significance  
 278 of the intimate processes by which the maize plant regulates its water uptake

behavior (hence its water status) by modifying architectural and/or hydraulic traits. Four sets of simulations were performed. The first set (I, tab. 1) highlights the effect of the root-to-shoot surface ratio (RSSR) by changing the number of seminal roots for given hydraulic parameters and transpiration demand. The second set of simulations (II, tab. 1) focuses on the effect of conductivity modifiers. The third set (IIIa-d, tab. 1) simulates the influence of management practices (well-watered, water-stressed, deficit irrigation and partial root-zone drying). Finally, the last set (IVa-f, tab. 1) was used to analyze the sensitivity of the water flow regulation mechanisms.

In all simulations, the soil and atmosphere initial water potential were respectively set to -0.01 and -95 MPa. Environmental conditions (temperature, light and VPD) were set according to a summer day in a greenhouse in Louvain-la-Neuve (Belgium, +50° 39' 59.91", +4° 37' 10.56"). A sandy soil with a low water retention capacity was considered in order to observe rapid water shortage for the plant. The simulation time step was 1h for the plant development and 10 minutes for the water fluxes. Data were exported every 12 hours, from the 50<sup>th</sup> to the 350<sup>th</sup> hour of the simulations.

### 3.2. *Effect of root system size and hydraulic regulations*

Water uptake by plants can be considered in economical terms, the shoot and the root defining respectively a potential demand and a supply capacity. In the plant-soil system, the evaporative demand is mainly defined by the leaf area, the stomatal conductance ( $g_s$ ) and the VPD. On the other hand, the supply capacity is set by the root system architecture, its hydraulic properties and the local soil water potential (Draye et al., 2010; Lobet et al., 2013). When the demand exceeds the supply capacity, the system is put under strain

Table 1: Conditions for simulations. PRD = partial root zone drying. RW = Rewatering.  
# of SR = number of seminal roots

| Simulation | # of SR | regulation      | carbon | water depletion |
|------------|---------|-----------------|--------|-----------------|
| I          | 0-6     | none            | no     | no              |
| II         | 0-6     | all             | no     | no              |
| IIIa       | 4       | all             | yes    | no              |
| IIIb       | 4       | all             | yes    | yes             |
| IIIc       | 4       | all             | yes    | PRD             |
| IIId       | 4       | all             | yes    | RW              |
| IVa        | 4       | enhanced $g_s$  | yes    | yes             |
| IVb        | 4       | inhibited $g_s$ | yes    | yes             |
| IVc        | 4       | enhanced $K_r$  | yes    | yes             |
| IVd        | 4       | inhibited $K_r$ | yes    | yes             |
| IVe        | 4       | enhanced $K_x$  | yes    | yes             |
| IVf        | 4       | inhibited $K_x$ | yes    | yes             |

304 until the demand decreases or the supply increases.

305

306 In order to test the effect of the root-to-shoot surface ratio (RSSR) on  
307 the hydraulic status of the plant, simulations with different initial numbers  
308 of seminal roots (between 0 and 6) and with the same root growth rate and  
309 shoot system development were performed. The carbon module was disabled  
310 to avoid feedbacks between transpiration, growth and C allocation (growth  
311 rates were set to their potential values).

312

313 In the first set of simulations, the hydraulic regulation and soil water de-  
314 pletion were disabled to isolate the effect of the root system size (tab. 1.I).  
315 These were enabled in the second set of simulations, through the use of mod-  
316 ifier functions (tab. 1.II).

317

### 318 *3.3. Effect of irrigation practices*

319 Four watering scenarios were simulated in order to compare the response  
320 of plants to different water management practices (tab. 1.III): well-watered  
321 (WW), water deficit (WD), partial root zone drying (PRD) and re-watering  
322 (RW). In the WW treatment, the soil water potential was maintained at a  
323 constant value (field capacity) during the whole simulation while in the WD  
324 treatment, the absorbed water was depleted from the corresponding soil vox-  
325 els and a basic horizontal redistribution of water in the soil was enabled. In  
326 the PRD treatment, the soil domain was split vertically in two compartments  
327 simulated respectively as WW and WD. Finally, in the RW simulation, the  
328 soil was treated as in the WD treatment, except that re-watering events were

329 simulated every 100 hours by resetting the initial soil water potential in all  
 330 voxels (fig. 7.A). All treatments were applied from the beginning of the sim-  
 331 ulations.

332

### 333 3.4. Effect of regulation sensibility

334 Finally, sensitivity analysis was performed to assess the effect of the reg-  
 335 ulation of  $g_s$ ,  $K_r$  and  $K_x$  implemented through the use of modifiers (tab.  
 336 1.IV). For each conductance, the modifier function was shifted towards lower  
 337 and higher water potential values, leading, respectively to delayed and faster  
 338 responses to changes in water potential (fig. 4). This analysis revealed  
 339 contrasting differences between the relative effects of the three regulation  
 340 processes.

341

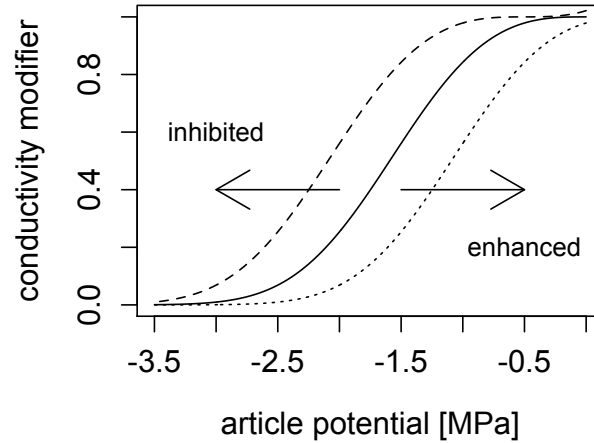


Figure 4: Regulation sensibility. Plain line represents the control regulation curve. Dashed line represent the delayed response. Dotted line represent the faster response.

## 342 4. Results and discussions

### 343 4.1. Model comparison with experimental data

344 The architectural and water flux modules were compared against exper-  
345 imental data independent of the one used to calibrate the model (Girardin,  
346 2000; Hu et al., 2011; Postma and Lynch, 2011; Mollier and Pellerin, 1999).  
347 PlaNet-Maize was able to reproduce realistic kinetics of root and shoot  
348 growth and water fluxes, integrated at the plant level (fig. 5). Since the  
349 aim of the model was not to reproduce a specific genotype X environment  
350 combination, but rather explore the dynamic of the system, we estimated  
351 such similarity between simulated and experimental data was sufficient to  
352 validate the model.

### 354 4.2. Root system size and hydraulic regulations

355 Figure 6.A summarizes the relationship between RSSR and the plant wa-  
356 ter potential (computed at the collar level). Considering a permanent wilting  
357 point at -1.5 MPa, the simulations suggest that a minimum RSSR value of  
358 2.0 is needed for a theoretical plant that would not regulate its root, leaf and  
359 axial conductances. Allowing the plant to regulate these hydraulic properties  
360 widens the range of viable RSSR values (fig. 6.B).

361  
362 These simulation results show that a shoot:root allometry is beneficial for  
363 the maintenance of water potential values above the permanent wilting point  
364 (-1.5 MPa). They also indicate that the regulation of hydraulic properties  
365 contributes to stabilize the plant water potential but does not completely

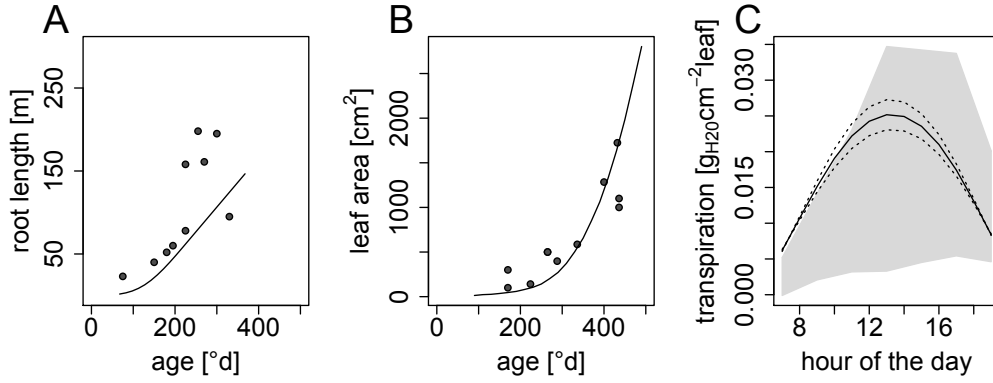


Figure 5: Comparison between experimental and simulated data. A. Comparison between experimental (circles, data from Hu et al. (2011); Mollier and Pellerin (1999)) and simulated (line) root length values. B. Comparison between experimental (circles, data from Girardin (2000); Postma and Lynch (2011)) and simulated (line) shoot area values. C. Comparison between experimental (grey area) obtained in greenhouse experiment (same environmental condition as for the simulations) and simulated (lines) transpiration values. Plain line represents the mean simulated transpiration value while the dashed lines represent the standard deviation range due to day to day variations .

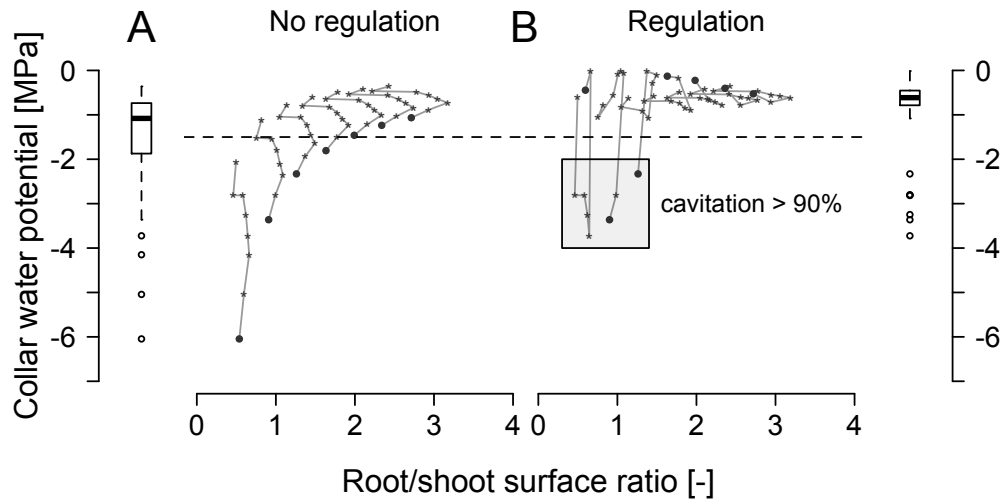


Figure 6: Relationship between the Root-to-shoot surface ratio and the collar water potential. A. Without hydraulic regulation. B. With hydraulic regulation. Dashed lines represent the permanent wilting point (-1.5 MPa). Each point represent the collar water potential at noon at different days, with plain circles representing the last day of simulation. Lines connect points from the same plant to highlight the dynamic of the system. In B, the grey box indicate the points were cavitation in the plant exceed 90% of the total system. Distribution of water potentials are highlighted by the boxplots on either side of the graph.

366 overcome the effect of structural allometry. As such, the simulations repro-  
367 duce many experimental results that showed that an undersized root system  
368 leads to greater tension in the xylem, leading to important cavitation events  
369 (fig. 6.B, grey box, Hacke et al. (2000b); Sperry et al. (1998). The simula-  
370 tions also suggest that the range of viable root-to-shoot ratios is rather large  
371 and influenced by the hydraulic regulation strategy used by plants.

372 *4.3. Hydraulic signals alone are not sufficient to reproduce the effects of dif-*  
373 *ferent irrigation practices*

374 Compared to WW, all treatments led to a clear reduction of the soil water  
375 potential. The PRD was able to maintain the soil water potential at a lower  
376 but stable value until 250 hours, thereby avoiding the abrupt transitions that  
377 occur in the RW treatment (fig. 7.A).

378  
379 Not surprisingly, reducing the water availability for the plant had a di-  
380 rect effect on the transpiration (fig. 7.B). Consistently with the soil water  
381 potential evolution, WD plants presented the lowest transpiration, while the  
382 PRD and RW scenario generated intermediate transpiration. The reduction  
383 of transpiration, in turn, led to a reduction of plant growth, although rela-  
384 tively small (fig. 7.C).

385  
386 Throughout the simulation, the leaf water potential displayed a regular  
387 decline in all treatments, which should be attributed to a decrease in the  
388 RSSR (fig. 7.D). Interestingly, until 250 hours, the leaf water potential dif-  
389 ference between the different treatments and WW were similar to the soil  
390 water potential difference.

391

392       Figure 8 describes the influence of the irrigation technique on the root  
393 water status. WD plants have smaller root systems, lower root water po-  
394 tential and conductivities and also a reduced uptake compared to the WW  
395 plant. It is unclear whether the latter results from the smaller uptake capacity  
396 (root surface) and/or the smaller demand (leaf size and leaf conductance).  
397 As for the previous results, the PRD and RW scenarios exhibit intermediate  
398 values.

399

400       One should notice that the water uptake profiles did not strictly cor-  
401 respond to the root surface profiles, particularly for the WD plant. The  
402 phenomenon, known as the compensation mechanism (Draye et al., 2010;  
403 Couvreur et al., 2012) arises when regions of the root system are surrounded  
404 by a soil layer that has a lower conductivity than the soil in the vicinity  
405 of the rest of the root system, or when the integrated conductance between  
406 roots and the collar differ among roots. When this occurs, the water uptake  
407 by roots with low soil/root conductances will be lowered and compensated  
408 by a larger water uptake by the other roots. This occurs typically in the  
409 WD, in the non-irrigated part of the PRD and in the RW treatments, where  
410 the lower root water potential increases water uptake with depth. In other  
411 words, uptake sites are not only defined by the root placement in the soil.  
412 They depend on the integration of the root system architecture, the root hy-  
413 draulic properties and the water availability in the surrounding soil. (Draye  
414 et al., 2010; Lobet et al., 2013).

415

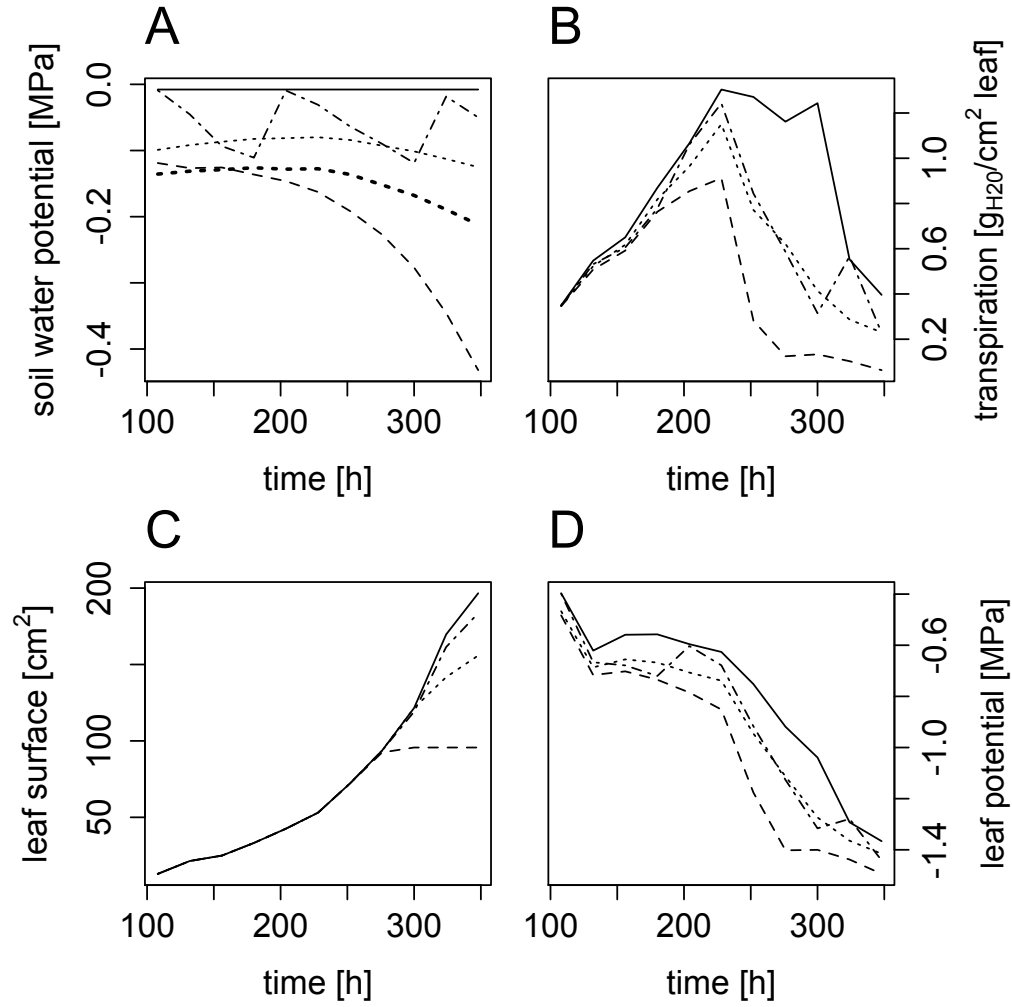


Figure 7: Effect of the irrigation technique on the plant water status. A. Average soil water content at the vicinity of the roots. B. Transpiration rate. C. Total leaf surface. D. Leaf water potential. Plain lines: WW. Dashed lines: WS. Dotted lines: PRD (all). Bold dotted line: PRD (dry compartment). Dashed-dotted lines: RW. Values were taken at 12:00 at every day of the simulation.

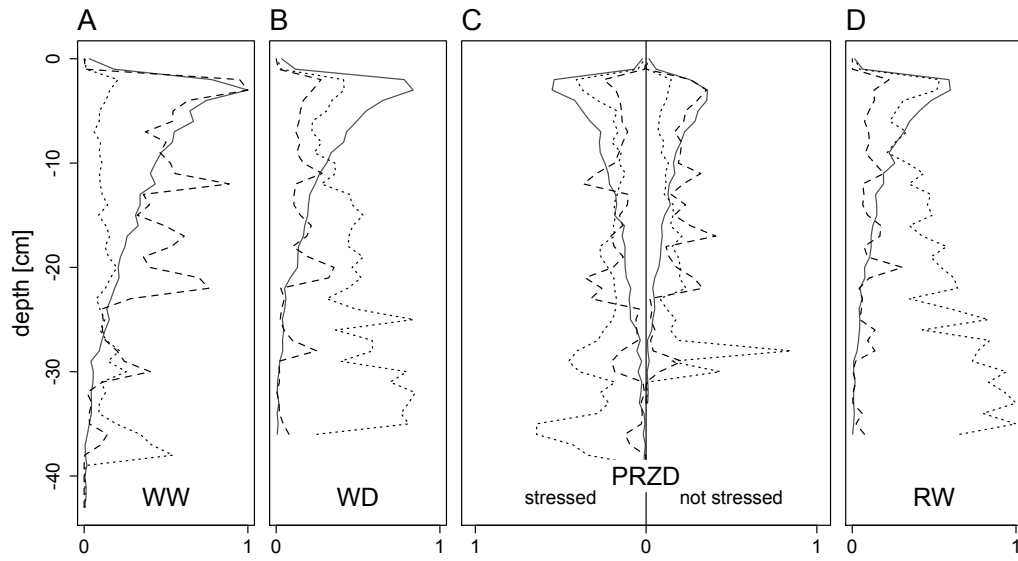


Figure 8: Effect of the irrigation technique on the roots: A. well watered, B. water deficit, C. partial root zone drying (left panel: water stress; right panel: well watered) and D. re-watering. Plain grey lines represent the root surface (sum per depth), dashed lines represent the water uptake (sum per depth) and dotted lines represent the root water potential (unsigned average per depth). All values are normalized to the maximal value of each variable across all scenarios.

416        PlaNet-Maize effectively reproduces the effect of different irrigation prac-  
417        tices that are generally observed experimentally. As a general rule, decreas-  
418        ing the amount of available water for the plant reduces its transpiration and,  
419        above a certain threshold, decreases plant growth as well (fig. 7).

420

421        However, PlaNet-Maize could not capture experimental difference classi-  
422        cally observed between PRD and RW treatment. Indeed, a growing body  
423        of evidence indicates that PRD has a beneficial influence on the plant water  
424        consumption and yield compared to RW (as an example, see the works of  
425        Dodd (2007) and Kang et al. (2000)). In these experiments, no major differ-  
426        ences in leaf water potential were observed (as in our simulations, fig. 7.C),  
427        but significant differences in final yield were recorded between the treat-  
428        ments. Evidently, the transpiration assimilation feedback that is used in  
429        PlaNet-Maize is not able to simulate this response. For this to work, the  
430        instantaneous WUE coefficient should be adjusted as a function of the soil  
431        water potential sensed by the plant.

432

433        The yield response to PRD is generally explained by differences in ab-  
434        scissic acid (ABA) production in the roots. It seems plausible from our  
435        simulations that a root signal informing leaves about the soil water potential  
436        might be sufficient to adjust stomatal conductance and restore a leaf wa-  
437        ter potential equivalent to the WW. This would reduce assimilation on the  
438        short term but would save water for future use. Including ABA signalling  
439        in PlaNet-Maize might thus help to reproduce the typical yield response  
440        to PRD treatment. This would lead ultimately to an apparent adjustment

441 of the instantaneous WUE coefficient as a function of the soil water potential.

442

443 Based on the response of the leaf water potential ( $\psi_{leaf}$ ) to decrease in  
444 soil water potential ( $\psi_{soil}$ ), plant can exhibit either an isohydric ( $\psi_{leaf}$  re-  
445 mains constant under decreasing  $\psi_{soil}$ ) or anisohydric ( $\psi_{leaf}$  decreases under  
446 decreasing  $\psi_{soil}$ ) behaviour. In our model,  $\psi_{leaf}$  tends to decrease with a  
447 decreasing  $\psi_{soil}$ , which suggest an anisohydric behaviour (fig. 9). However,  
448 experimental data have shown the opposite in the fields, hence labelling maize  
449 as an isohydric plant (Tardieu et al., 1992; Ben Haj Salah and Tardieu, 1997).  
450 Such isohydric behaviour has been characterised by an interplay between  $g_s$ ,  
451  $\psi_{leaf}$  and the concentration of ABA in the xylem sap (Tardieu and Simon-  
452 neau, 1998). Implementing ABA in PlaNet-Maize should help enabling an  
453 isohydric behaviour in our simulations.

#### 454 4.4. Regulation sensibility

455 The results of the analysis on  $g_s$  are given in figures 10.1A-1C and 11.1A-  
456 1C. Increasing the sensibility of  $g_s$  to the water potential leads to early stom-  
457 atal closure which causes the reduction of transpiration and the maintenance  
458 of a high water potential. This does not affect root conductances but has  
459 a dramatic impact on plant growth and leaf area. Fortunately, the saved  
460 water is available later and allows plants to transpire for a longer period (fig.  
461 10.1C, arrow). On the opposite, decreasing the sensibility of  $g_s$  to the water  
462 potential leads to stomata being more open, a higher transpiration and a  
463 rapid decrease of the plant water potential, which causes and expected de-  
464 crease of the root radial conductance and increase of xylem cavitation.

465

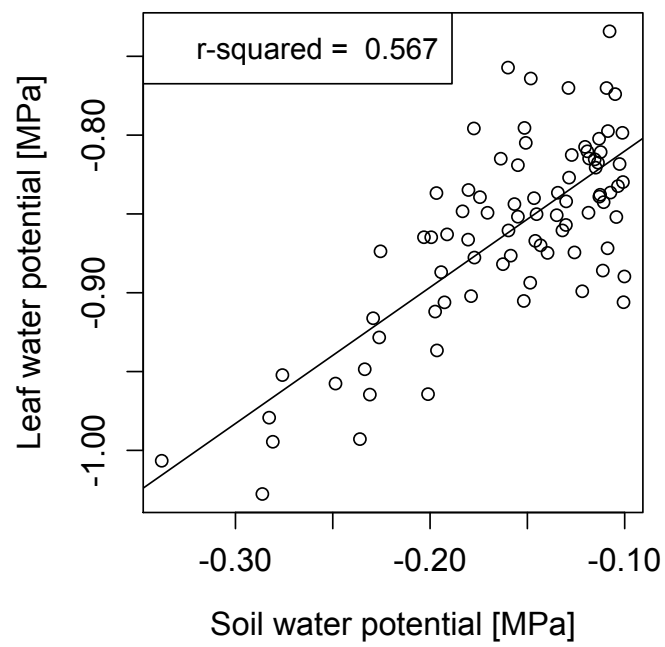


Figure 9: Response of leaf water potential to soil drying in PlaNet-Maize.

466 The changes in the sensibility of  $K_r$  (figs. 10.2A-2C and 11.2A-2C) and  
467  $K_x$  (figs. 10.3A-3C and 11.3A-3C) have smaller effect on the plant water  
468 status compared to  $g_s$ . The largest effects are seen under faster  $K_r$  response,  
469 where a rapid reduction of the root radial conductivity leads to a sensible  
470 reduction of the plant water potential, stomatal opening and transpiration.  
471 Interestingly, this effect was obtained without any consequences on growth  
472 and leaf area and appears thus as plausible way to achieve conservative wa-  
473 ter use. Neither delayed  $K_r$  nor altered  $K_x$  responses have important conse-  
474 quences on the plant water dynamics.

475

476 Contrasting transpiration dynamics were thus observed between plants  
477 with modified  $g_s$  and  $K_r$  responses. On the one side, the delayed  $g_s$  response  
478 causes the plant to follow the evaporative demand at the cost of a quick  
479 drying of the soil, and is therefore valuable in conditions where water supply  
480 is non limiting. On the other side, the faster  $g_s$  and  $K_r$  responses lead to a  
481 conservative use of water that allows transpiration maintenance later in the  
482 season. Recent experimental data suggest that such water saving strategy  
483 during the vegetative growth is correlated with terminal drought resistance  
484 (Zaman-Allah et al., 2011a,b; Kholová et al., 2010; Tardieu, 2011). Some of  
485 our simulations differed from these results by the strong reduction in plant  
486 size which was not observed in field conditions. It has to be noted, however,  
487 that the drought susceptible and resistant genotypes used in these experi-  
488 ments did not differ necessarily by those mechanisms that are considered in  
489 our simulations.

490

491 Beyond their effect on the plant water dynamics, the modifier functions  
492 also change the spatial patterns of water extraction by roots in slightly dif-  
493 ferent ways. These modifications rely mostly on the changes of radial vs  
494 axial root conductivities, as a large  $K_r/K_x$  ratio tends to concentrate water  
495 uptake in the upper layers, while a lower ratio tends to distribute the uptake  
496 vertically (Draye et al., 2010). This is most noticeable in the  $K_r$  sensitivity  
497 analysis where the  $K_r/K_x$  ratio increases under a delayed response of  $K_r$  and  
498 decreases under a faster response (fig. 11). As a consequence, the uptake  
499 pattern is decreased by a constant value under the faster  $K_r$  response while  
500 it is increased in upper layers under the delayed  $K_r$  response.

501

502 These sensitivity analyses emphasize the interplay between the regulation  
503 of  $g_s$ ,  $K_r$  and  $K_x$ . Obviously, making these regulations all dependent on the  
504 local water potential is largely responsible for this interplay, but this only  
505 reflects the structural integration of these regulations which all contribute  
506 to the same water flow. It follows that the classical experimental methodol-  
507 ogy consisting in the manipulation of a single regulating mechanism should  
508 be complemented with at least some characterisation of how this manipula-  
509 tion could affect the other mechanisms. PlaNet-Maize provides the required  
510 framework to integrate many different factors influencing the water dynam-  
511 ics.

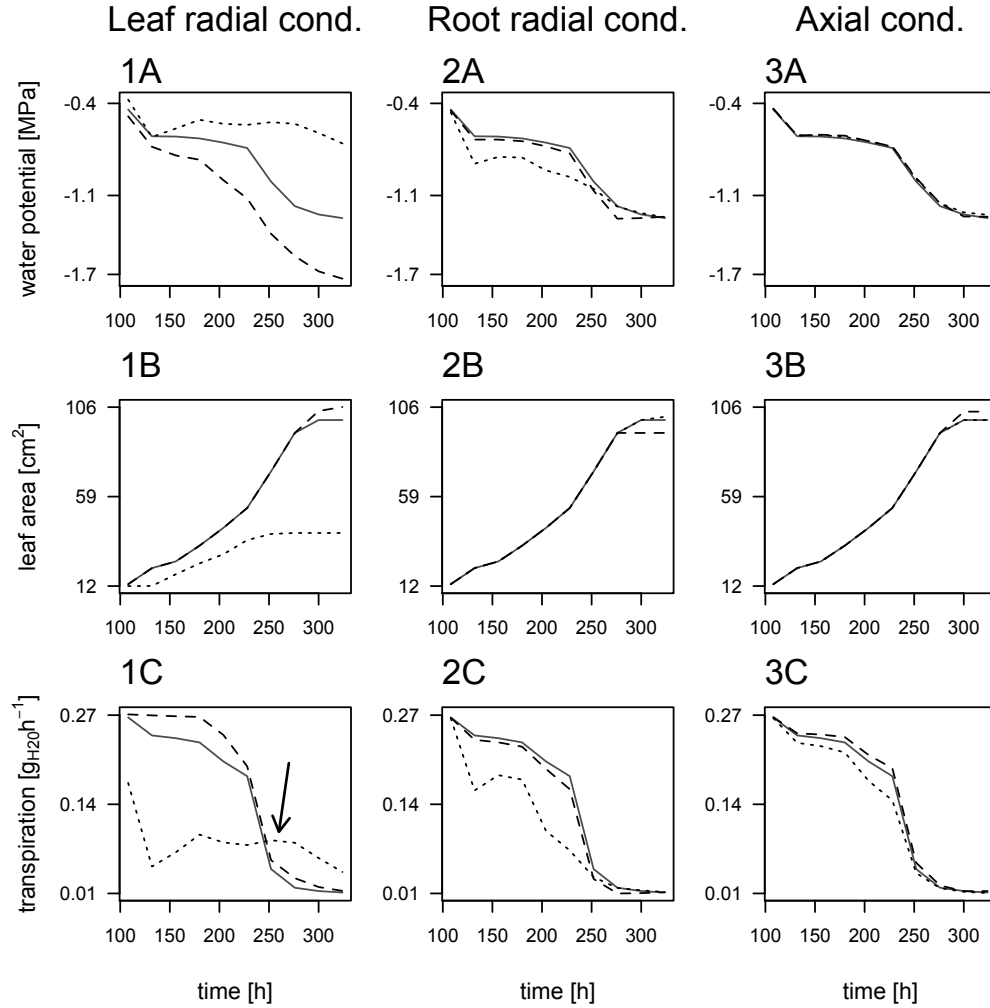


Figure 10: Effect of the regulation processes on the plant water status. 1-. changed regulation of leaf radial conductivity; 2-. changed regulation of root radial conductivity; 3-. changed regulation of axial conductance; -A. evolution of the collar water potential; -B. evolution of shoot surface; -C. evolution of the relative transpiration (to the leaf area). Plain lines represent the controls, with no modification of the regulation processes, dashed lines represents the inhibited regulation processes and dotted lines represent the enhanced regulation processes.

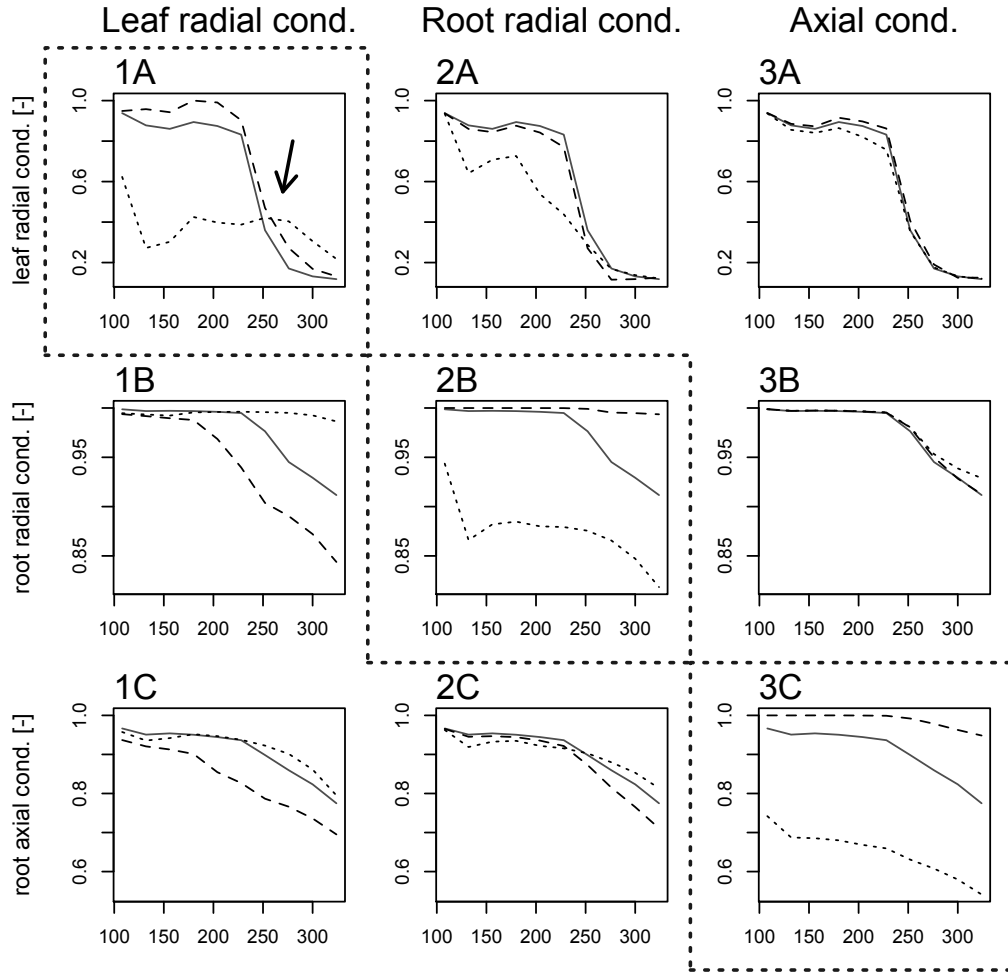


Figure 11: Effect of the regulation processes on the plant water status. 1-. changed regulation of leaf radial conductivity; 2-. changed regulation of root radial conductivity; 3-. changed regulation of axial conductance; -A. evolution of the leaf radial conductivity; -B. evolution of the root radial conductivity; -C. evolution of the root axial conductance. Plain lines represent the controls, with no modification of the regulation processes, dashed lines represents the inhibited regulation processes and dotted lines represent the enhanced regulation processes. Framed plots highlight the differences in regulation sensibility for each scenario.

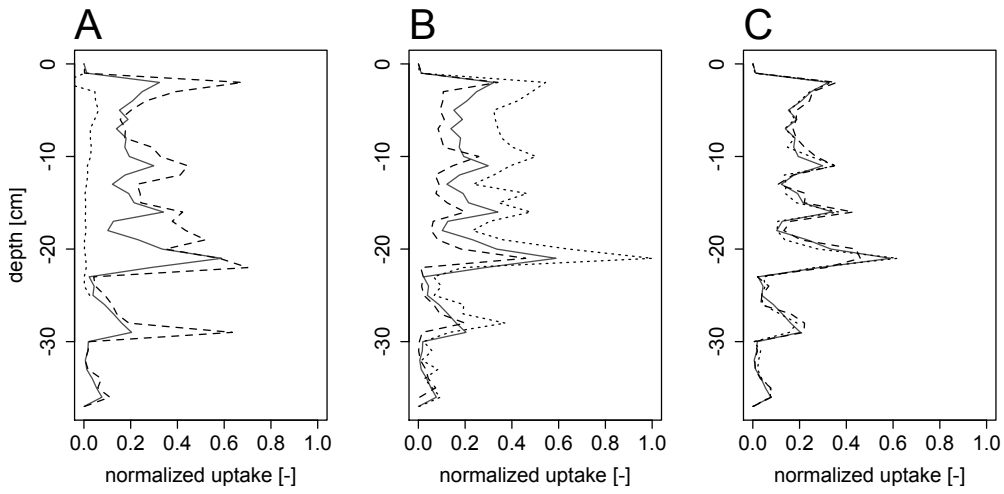


Figure 12: Effect of the regulation processes on the root water uptake profiles. A. modified leaf radial conductivity; B. modified root radial conductivity; C. modified axial conductance. Plain grey lines represent the controls, with no modification of the regulation processes, dashed lines represents the inhibited regulation processes and dotted lines represent the enhanced regulation processes. All values are normalized.

## 512 5. Conclusions and perspectives

### 513 5.1. *Functional-structural modeling as a useful tool to decrypt plant water* 514 *relations*

515 It is now commonly accepted that functional-structural models are essen-  
516 tial tools to access the complexity of plant systems (Tardieu, 2010; Godin  
517 and Sinoquet, 2005). They allow scientists to investigate the dynamics of  
518 complex systems and unravel the effect of multiple regulation mechanisms  
519 (Hill et al., 2013).

520  
521 PlaNet-Maize was successfully used to simulate water flows in the entire  
522 plant continuum with a resolution down to the plant element level. This first  
523 version of the model was able to reproduce the effect of root architecture and  
524 hydraulic properties on the water relations in the plant as previously shown  
525 in former models (Draye et al., 2010; Janott et al., 2011). In addition, by  
526 implementing simple regulatory processes acting on the local values of  $K_r$   
527 and  $K_x$ , we found that the virtual plant was able to tightly control its water  
528 status.

529  
530 Divergences were observed between experimental and modeling results  
531 for contrasted water management practices. PlaNet-Maize was not able to  
532 reproduce the beneficial effect of PRD over RW treatment. This difference  
533 appeared supposably because a finer signaling of local soil conditions (via the  
534 production and translocation of ABA) was not implemented in the model.

535  
536 Finally, PlaNet-Maize was used to show the effect of modified sensibilities

537 for the different regulation sites. Enhanced sensibilities enable the plant to  
538 react more quickly to environmental constrain and therefore to have conser-  
539 vative water use behaviors. Results show that changing the sensibilities at  
540 the root system level (either  $K_x$  and/or  $K_r$ ) can have a non-negligible effect  
541 on the plant water status by increasing its overall water use efficiency.

## 542 5.2. *Envisaged model development*

543 A first envisaged development path for PlaNet-Maize would be the im-  
544 provement of its water-related modules. Firstly, in order to model more  
545 realistic condition and test the influence of different soil environment, inte-  
546 gration of a more realistic soil module, such as Hydrus or R-SWMS (Javaux  
547 et al., 2008), is required. Moreover, as discussed previously, the implementa-  
548 tion of a long distance signal between the root and the shoot, such as ABA,  
549 should be implemented to enable a finer sensing of the local variations in the  
550 soil water content.

551

552 A second direction for the development of the model would be to link it  
553 with lower scale models, down to the cell level (Band et al., 2012). Indeed,  
554 the modular nature of the PlaNet based models allows an easy and straight-  
555 forward integration with other models.

556

557 Finally, a third direction would be to extend the model from vegetative  
558 to reproductive stages, in order to assess the effect of terminal drought stress  
559 on grain formation and, ultimately, yield. Again, the modular structure  
560 of PlaNet-Maize easily allows the creation of new article types needed to  
561 simulate flowering and grain filling.

### 562 5.3. *Future experimental needs*

563 Functional-structural plant models rely on quantitative data to be ac-  
564 curate and representative of the reality. Unfortunately, in some area, no-  
565 tably regarding plant hydraulics, this quantitative knowledge is missing. As  
566 functional-structural models are now recognized as useful tools for the plant  
567 science community (Tardieu, 2010; Hill et al., 2013), more and more modeling  
568 projects arise and therefore there is an increasing need for more quantitative  
569 biological data. In particular, more data are needed to characterise hydraulic  
570 properties of the different plant organs as function of endogenous (e.g. age,  
571 type and order of root segments) and exogenous factors (e.g. response curves  
572 to drying soil).

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## 580 **References**

581 Allen, Jr., L.H., Kakani, V.G., Vu, J.C.V., Boote, K.J., 2011. Elevated CO<sub>2</sub>  
582 increases water use efficiency by sustaining photosynthesis of water-limited  
583 maize and sorghum. *Journal of Plant Physiology* 168, 1909–1918.

- 584 Band, L.R., Fozard, J.A., Godin, C., Jensen, O.E., Pridmore, T., Bennett,  
585 M.J., King, J.R., 2012. Multiscale Systems Analysis of Root Growth and  
586 Development: Modeling Beyond the Network and Cellular Scales. *The*  
587 *Plant Cell* 24, 3892–3906.
- 588 Ben Haj Salah, H., Tardieu, F., 1997. Control of Leaf Expansion Rate of  
589 Droughted Maize Plants under Fluctuating Evaporative Demand. *Plant*  
590 *Physiology* , 893–900.
- 591 Bidel, L., Pagès, L., Riviere, L., Pelloux, G., Lorendeau, J., 2000. Mass-  
592 FlowDyn I: A Carbon Transport and Partitioning Model for Root System  
593 Architecture. *Annals of Botany* 85, 869–886.
- 594 Bohrer, G., Mourad, H., Laursen, T.A., Drewry, D., Avissar, R., Poggi,  
595 D., R, O., Katul, G.G., 2005. Finite element tree crown hydrodynamics  
596 model (FETCH) using porous media flow within branching elements: a  
597 new representation of tree hydrodynamics. *Water Resources Research* 41,  
598 1–17.
- 599 Bramley, H., Turner, N., Turner, D., Tyerman, S.D., 2009. Roles of Mor-  
600 phology, Anatomy, and Aquaporins in Determining Contrasting Hydraulic  
601 Behavior of Roots. *Plant Physiology* 150, 348–364.
- 602 Cochard, H., 2002. Xylem embolism and drought-induced stomatal closure  
603 in maize. *Planta* 215, 466–471.
- 604 Couvreur, V., Vandenberg, J., Javaux, M., 2012. A simple three-dimensional  
605 macroscopic root water uptake. *Hydrology and Earth System Sciences* 16,  
606 2957–2971.

- 607 Damour, G., Simonneau, T., Cochard, H., Urban, L., 2010. An overview of  
608 models of stomatal conductance at the leaf level. *Plant, Cell and Environ-*  
609 *ment* 33, 1419–1438.
- 610 Diggle, A.J., 1988. ROOTMAP—a model in three-dimensional coordinates  
611 of the growth and structure of fibrous root systems. *Plant and Soil* 105,  
612 169–178.
- 613 Dodd, I.C., 2007. Soil moisture heterogeneity during deficit irrigation alters  
614 root-to-shoot signalling of abscisic acid. *Functional Plant Biology* 34, 439–  
615 448.
- 616 Doussan, C., Pierret, A., Garrigues, E., Pagès, L., 2006. Water Uptake by  
617 Plant Roots: II – Modelling of Water Transfer in the Soil Root-system  
618 with Explicit Account of Flow within the Root System – Comparison with  
619 Experiments. *Plant and Soil* 283, 99–117.
- 620 Doussan, C., Vercambre, G., Pagès, L., 1998. Modelling of the hydraulic  
621 architecture of root systems: and integrated approach to water absorption  
622 - Distribution of axial and radial conductances in maize. *Annals of Botany*  
623 81, 225–232.
- 624 Draye, X., Kim, Y., Lobet, G., Javaux, M., 2010. Model-assisted integration  
625 of physiological and environmental constraints affecting the dynamic and  
626 spatial patterns of root water uptake from soils. *Journal of Experimental*  
627 *Botany* 61, 2145–2155.
- 628 Draye, X., Pagès, L., 2006. CrossTalk: a simulation platform for the linking

629 of existing soil, plant and atmosphere models. IEEE-Conference Proceeding  
630 Service PM06, 95–100.

631 Drouet, J.L., Pagès, L., 2003. GRAAL: a model of GRowth, Architecture  
632 and carbon ALlocation during the vegetative phase of the whole maize  
633 plant. Model description and parametrisation. Ecological Modelling 165,  
634 143–173.

635 Drouet, J.L., Pagès, L., 2007. GRAAL-CN: A model of GRowth, Archi-  
636 tecture and ALlocation for Carbon and Nitrogen dynamics within whole  
637 plants formalised at the organ level. Ecological Modelling 206, 231–249.

638 Enstone, D.E., Peterson, C.A., Ma, F., 2003. Root Endodermis and Exoder-  
639 mis: Structure, Function, and Responses to the Environment. Journal of  
640 Plant Growth Regulation 21, 335–351.

641 Fournier, C., Andrieu, B., 1998. A 3D Architectural and Process-based Model  
642 of Maize Development. Annals of Botany 81, 233–250.

643 Ge, Z., Rubio, G., Lynch, J.P., 2000. The importance of root gravitropism  
644 for inter-root competition and phosphorus acquisition efficiency: results  
645 from a geometric simulation model. Plant and Soil 218, 159–171.

646 van Genuchten, M., 1980. A closed-form equation for predicting the hydraulic  
647 conductivity of unsaturated soils. Soil Science Society of America Journal  
648 44, 891–898.

649 Girardin, P., 2000. Ecophysiologie du maïs. Association Generale des Pro-  
650 ducteurs de Maïs, Montardon.

651 Godin, C., Sinoquet, H., 2005. Functional-structural plant modelling. *New*  
652 *Phytologist* 166, 705–708.

653 Hacke, U.G., Sperry, J.S., Ewers, B.E., Ellsworth, D.S., Schäfer, K.V.R., RR,  
654 O., 2000a. Influence of soil porosity on water use in *Pinus taeda*. *Oecologia*  
655 124, 495–505.

656 Hacke, U.G., Sperry, J.S., Pittermann, J.J., 2000b. Drought experience and  
657 cavitation resistance in six shrubs from the Great Basin, Utah. *Basic and*  
658 *Applied Ecology* 1.

659 Hill, K., Porco, S., Lobet, G., Zappala, S., Mooney, S., Draye, X., Ben-  
660 nett, M.J., 2013. Root Systems Biology: bridging regulatory networks to  
661 rhizosphere-scale processes. *Plant Physiology* .

662 Hochholdinger, F., Park, W.J., Sauer, M., Woll, K., 2004. From weeds to  
663 crops: genetic analysis of root development in cereals. *TRENDS in Plant*  
664 *Science* 9, 42–48.

665 Holbrook, N.M., Zwieniecki, M., 1999. Embolism Repair and Xylem Tension:  
666 Do We Need a Miracle? *Plant Physiology* 120, 7–10.

667 Hu, T., Kang, S., Li, F., Zhang, J., 2011. Effects of partial root-zone irri-  
668 gation on hydraulic conductivity in the soil-root system of maize plants.  
669 *Journal of Experimental Botany* 62, 4163–4172.

670 Janott, M., Gayler, S., Gessler, A., Javaux, M., Klier, C., Priesack, E., 2011.  
671 A one-dimensional model of water flow in soil-plant systems based on plant  
672 architecture. *Plant and Soil* 341, 233–256.

- 673 Jarvis, P.G., 1976. The Interpretation of the Variations in Leaf Water Potent-  
674 tial and Stomatal Conductance Found in Canopies in the Field. Philosoph-  
675 ical Transactions of the Royal Society B: Biological Sciences 273, 593–610.
- 676 Javaux, M., Schroeder, T., Vanderborght, J., Vereecken, H., 2008. Use of  
677 a three-dimensional detailed modeling approach for predicting root water  
678 uptake. . Vadose Zone Journal 7, 1079–1088.
- 679 Javot, H., Maurel, C., 2002. The role of aquaporins in root water uptake.  
680 Annals of Botany 90, 301–313.
- 681 Kang, S., Liang, Z., Pan, Y., Shi, P., 2000. Alternate furrow irrigation for  
682 maize production in an arid area. Agricultural water management 45,  
683 267–274.
- 684 Kholová, J., Hash, C.T., Kakkerá, A., Kocová, M., Vadez, V., 2010. Con-  
685 stitutive water-conserving mechanisms are correlated with the terminal  
686 drought tolerance of pearl millet [*Pennisetum glaucum* (L.) R. Br.]. Jour-  
687 nal of Experimental Botany 61, 369–377.
- 688 Landsberg, J., Fowkes, N., 1978. Water movement through plant roots.  
689 Annals of Botany 42, 493–508.
- 690 Li, Y., Sperry, J.S., Shao, M., 2009. Hydraulic conductance and vulnerability  
691 to cavitation in corn (*Zea mays* L.) hybrids of differing drought resistance.  
692 Environmental and Experimental Botany 66, 341–346.
- 693 Lobet, G., Hachez, C., Chaumont, F., Javaux, M., Draye, X., 2013. Root wa-  
694 ter uptake and water flow in the soil-root domain, in: Eshel, A., Beeckman,  
695 T. (Eds.), Plant Roots: The Hidden Half. CRC Press, New york.

696 Lynch, J.P., Nielsen, K.L., Davis, R.D., Jablokow, A.G., 1997. SimRoot:  
697 Modelling and visualization of root systems. *Plant and Soil* 188, 139–151.

698 Maurel, C., Verdoucq, L., Luu, D., Santoni, V., 2008. Plant aquaporins:  
699 membrane channels with multiple integrated functions. *Annual Review of*  
700 *Plant Biology* 59, 595–624.

701 McCully, M.E., Huang, C.X., Ling, L.E.C., 1998. Daily embolism and refilling  
702 of xylem vessels in the roots of field-grown maize. *New Phytologist* 138,  
703 327–342.

704 Mollier, A., Pellerin, S., 1999. Maize root system growth and development  
705 as influenced by phosphorus deficiency. *Journal of Experimental Botany*  
706 50, 487–497.

707 Pagès, L., Jordan, M.O., Picard, D., 1989. A simulation model of the three-  
708 dimensional architecture of the maize root system. *Plant and Soil* 119,  
709 147–154.

710 Pagès, L., Vercambre, G., Drouet, J.L., Lecompte, F., Collet, C., LeBot, J.,  
711 2004. RootTyp: a generic model to depict and analyse the root system  
712 architecture. *Plant and Soil* 258, 103–119.

713 Peirce, G.J., 1936. The State of Water in Ducts and Tracheids. *Plant Phys-*  
714 *iology* 11, 623–628.

715 Postma, J.A., Lynch, J.P., 2011. Theoretical evidence for the functional ben-  
716 efit of root cortical aerenchyma in soils with low phosphorus availability.  
717 *Annals of Botany* 107, 829–841.

- 718 Somma, F., Hopmans, J., Clausnitzer, V., 1998. Transient three-dimensional  
719 modeling of soil water and solute transport with simultaneous growth, root  
720 water and nutrient uptake. *Plant and Soil* 202, 281–293.
- 721 Sperry, J.S., Adler, F.R., Campbell, G.S., Comstock, J.P., 1998. Limitation  
722 of plant water use by rhizosphere and xylem conductance: results from a  
723 model. *Plant, Cell and Environment* 21, 347–359.
- 724 Sperry, J.S., Donnelly, J.R., Tyree, M.T., 1988. Seasonal Occurrence of  
725 Xylem Embolism in Sugar Maple (*Acer saccharum*). *American Journal of*  
726 *Botany* 75, 1212–1218.
- 727 Steudle, E., 2001. The cohesion-tension mechanism and the acquisition of  
728 water by plant roots. *Annual Review of Plant Physiology and Plant Molec-*  
729 *ular Biology* 53, 847–875.
- 730 Steudle, E., Peterson, C.A., 1998. How does water get through roots? *Journal*  
731 *of Experimental Botany* 49, 775–788.
- 732 Tardieu, F., 2010. Why work and discuss the basic principles of plant mod-  
733 elling 50 years after the first plant models? *Journal of Experimental*  
734 *Botany* 61, 2039–2041.
- 735 Tardieu, F., 2011. Any trait or trait-related allele can confer drought tol-  
736 erance: just design the right drought scenario. *Journal of Experimental*  
737 *Botany* 63, 25–31.
- 738 Tardieu, F., Simonneau, T., 1998. Variability among species of stomatal con-  
739 trol under fluctuating soil water status and evaporative demand: modelling

740 isohydric and anisohydric behaviours. *Journal of Experimental Botany* 49,  
741 419–432.

742 Tardieu, F., Zhang, J., Katerji, N., Bethenod, O., Palmer, S., Davies, W.J.,  
743 1992. Xylem ABA controls the stomatal conductance of field-grown maize  
744 subjected to soil compaction or soil drying. *Plant, Cell and Environment*  
745 15, 193–197.

746 Tyree, M.T., Sperry, J.S., 1989. Vulnerability of Xylem to Cavitation and  
747 Embolism. *Annual Review of Plant Physiology and Plant Molecular Biol-*  
748 *ogy* 40, 19–36.

749 Wei, C., Tyree, M.T., Steudle, E., 1999. Direct Measurement of Xylem Pres-  
750 sure in Leaves of Intact Maize Plants. A Test of the Cohesion-Tension  
751 Theory Taking Hydraulic Architecture into Consideration. *Plant Physiol-*  
752 *ogy* 121, 1191–1205.

753 Zaman-Allah, M., Jenkinson, D.M., Vadez, V., 2011a. A conservative pattern  
754 of water use, rather than deep or profuse rooting, is critical for the terminal  
755 drought tolerance of chickpea. *Journal of Experimental Botany* 62, 4239–  
756 4252.

757 Zaman-Allah, M., Jenkinson, D.M., Vadez, V., 2011b. Chickpea genotypes  
758 contrasting for seed yield under terminal drought stress in the field differ  
759 for traits related to the control of water use. *Functional Plant Biology* 38,  
760 270.

761 Zwieniecki, M.A., Holbrook, N.M., 2009. Confronting Maxwell’s demon:

762 biophysics of xylem embolism repair. *TRENDS in Plant Science* 14, 530–  
763 534.