

Dermatoscopic features of nodular melanoma with Breslow thickness up to 2mm: a multicentric collaborative study by the International Dermoscopy Society

D. Sgouros,¹ A. Lallas,² H. Kittler,³ A. Zarras,¹ A. Kyrgidis,⁴ C. Papageorgiou,² S. Puig,⁵ A. Scope,⁶ G. Argenziano,⁷ I. Zalaudek,⁸ MA Pizzichetta,^{8,9} A. Marghoob,¹⁰ K. Liopyris,¹⁰ J. Malvehy,⁵ C. Oikonomou,¹ Á. Flórez,¹¹ Braun,¹² H. Cabo,¹³ G. Nazzaro,¹⁴ S. Lanssens,¹⁵ S. Menzies,¹⁶ J. Paoli,¹⁷ G. Kaminska-Winciorek,¹⁸ C. Longo,^{19,20} A. Katoulis,²¹ Z. Apalla,²² D. Ioannides,² L. Thomas,²³ I. Tromme,²⁴ D. Ogata,²⁵ C. Desinoti,¹ A. Geller,²⁶ A. Stratigos¹

1. 1st Department of Dermatology-Venereology, Andreas Sygros Hospital, National and Kapodistrian University of Athens School of Medicine, Athens, Greece
2. 1st Department of Dermatology, Aristotle University, Thessaloniki, Greece
3. H. Kittler Department of Dermatology, Medical University of Vienna, Austria
4. Department of Clinical Pharmacology, Aristotle University, Thessaloniki, Greece
5. Melanoma Unit, Dermatology Department, Hospital Clinic Barcelona, Universitat de Barcelona IDIBAPS, Barcelona, Spain
6. Medical Screening Institute, Sheba Medical Center and Sackler School of Medicine, Tel Aviv University, Israel
7. Dermatology Unit, University of Campania “Luigi Vanvitelli”, Naples, Italy
8. Department of Dermatology, University of Trieste, Italy
9. Division of Medical Oncology - Preventive Oncology, National Cancer Institute, Aviano, Italy
10. Memorial Sloan Kettering Cancer Center, Hauppauge, New York
11. Department of Dermatology, Pontevedra University Hospital, Spain.
12. Department of Dermatology, University Hospital Zürich, Zürich, Switzerland
13. Dermatology Institute of Medical Research, University of Buenos Aires, Argentina
14. **G. Nazzaro (please fill in your affiliation)**
15. Private practice Dermatology Maldegem, Maldegem, Belgium
16. Discipline of Dermatology, Sydney Medical School, The University of Sydney and Sydney Melanoma Diagnostic Centre, Royal Prince Alfred Hospital, Camperdown, NSW Australia
17. Department of Dermatology, Institute of Clinical Sciences, Sahlgrenska Academy, University of Gothenburg, Gothenburg, Sweden
18. The Center for Cancer Prevention and Treatment, Katowice, Poland
19. Department of Dermatology, University of Modena and Reggio Emilia, Modena, Italy
20. Centro Oncologico ad Alta Tecnologia Diagnostica, Azienda Unità Sanitaria Locale - IRCCS di Reggio Emilia, Italy
21. 2nd Department of Dermatology-Venereology, Atticon Hospital, National and Kapodistrian University of Athens, Athens, Greece
22. State Clinic of Dermatology, Hospital for Skin and Venereal Diseases, Thessaloniki, Greece
23. Department of Dermatology, Lyon University, France
24. Department of Dermatology, King Albert II Institute, Cliniques Universitaires Saint Luc, Brussels, Belgium
25. **D. Ogata (please fill in your affiliation)**

26. Department of Social and Behavioral Sciences, Harvard TH School of Public Health, Boston, MA, United States.

DS and AL are equally contributing first authors

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Corresponding Author: Alexander J. Stratigos, MD, 1st Department of Dermatology – Venereology, Andreas Sygros Hospital, 5 I. Dragoumi Str, 16 121, Athens, Greece,
alstrat2@hol.gr

Question: Which are the dermoscopic characteristics of a thin nodular melanoma up to 2mm Breslow thickness?

Findings: In this multicenter cohort study of 254 nodular tumors, we observed that nodular melanomas with a Breslow thickness ≤ 2 mm are asymmetric lesions in terms of colors and dermoscopic structures. Light brown coloration is the most frequent color seen in these tumors. Irregular brown dots/globules, irregular blue structureless areas, ulceration, white shiny blotches/strands, white shiny streaks, irregular (eccentric) black blotches, dotted and serpentine vessels are typical of features of thin (≤ 2 mm) nodular melanoma.

Meaning: Based on these findings, dermoscopy can help detect a nodular melanoma at an early phase of its evolution, based on Breslow thickness.

Abstract

Importance: Thin nodular melanoma (NM) often lacks conspicuous melanoma-specific criteria, and escapes clinical detection until it progresses to an advanced stage.

Objective: To investigate the dermatoscopic morphology of NM with a Breslow thickness up to 2 mm and to identify predictors for its differential diagnosis from other nodular tumors.

Design: Retrospective, morphological case-control study, conducted on behalf of the International Dermoscopy Society. Dermatoscopic images of NM and other nodular tumors were collected and analyzed by 5 independent investigators for the presence of pre-defined dermatoscopic variables. Predictors were calculated by multivariate logistic regression analysis.

Setting: Nineteen specialized skin cancer centers from 13 countries.

Participants: A total set of 254 patients with 254 tumors including 69 NM of Breslow thickness ≤ 2 mm, 96 NM thicker than 2mm and 89 non-melanoma ~~ocular~~ nodular lesions (BCC, dermal nevi, dermatofibromas, seborrheic keratosis, etc).

Main outcome and measures: Frequencies of dermatoscopic findings were calculated based on the categorization into 4 groups of dermatoscopic variables (overall architecture and colors, pigmented structures, non-pigmented structures excluding vessels, and vascular structures). Relative risks were calculated for the dichotomous variables. Adjusted odds ratio (OR) and corresponding 95% confidence intervals (CI) were calculated by conditional multivariate logistic regression.

Results: The most frequent colors seen in NMs with a Breslow thickness up to 2mm were light brown (50.7%), white (46.4%), blue (44.9%) and pink (43.5%).

Irregular brown dots/globules (42.0%), irregular blue structureless areas (33.3%), ulceration (29%), white shiny blotches/strands (27.5%), white shiny streaks (26.0%) and irregular (eccentric) black blotch (24.6%) were the most frequent dermatoscopic structures. The multivariate analysis revealed three dermatoscopic predictors of thin NM, as compared to non-melanoma nodular lesions: dotted vessels (3.4-fold), white shiny streaks (2.9-fold) and irregular blue structureless area (2.4-fold). In the comparative analysis between thinner and thicker NM, irregular brown dots/globules were found to be a predictor of NM up to 2mm of Breslow thickness (1.8-fold) and serpentine vessels a predictor of thicker NM (3.3-fold).

Conclusions and relevance: Our study sheds light into the dermatoscopic morphology of thin NM in comparison to thicker NM and provides useful clues for its differential diagnosis from other nodular tumors.

Keywords: nodular melanoma; thickness; pigmented tumors; non-pigmented tumors; non-melanoma tumors; dermatoscopy; dermoscopy

Introduction

Melanoma is a malignant skin tumor with an increasing incidence over the last decades. Melanoma is responsible for the majority of deaths from cutaneous malignancies, with more than 90000 new cases and almost 9000 deaths recorded in 2018 in the US.^{1,2} Despite primary and secondary prevention measures, melanoma represents a major public health issue in fair-skinned populations.

Among the four main histopathological subtypes of melanoma (superficial spreading, nodular, lentigo maligna and acral lentiginous), nodular melanoma (NM) comprises 12-30% of all melanomas, but at least 50% of all melanomas thicker than 2 mm, hence its worst prognosis and associated survival.^{3,4,5} A recent observational study including approximately 120000 patients showed that the nodular subtype was an independent risk factor of mortality, after adjustment for prognostic factors such as Breslow thickness, ulceration and stage.⁶ The detection of NMs while they are still thin could improve the prognosis, but is particularly difficult.

Dermatoscopy is a non-invasive diagnostic technique, widely applied for the clinical diagnosis of melanocytic and non-melanocytic tumors and other skin diseases. Several studies described the dermatoscopic characteristics of NM, suggesting that they substantially differ from the features known to typify superficial spreading melanoma.⁷⁻¹¹ Overall, the diagnostic accuracy of dermatoscopy for NM is considered to be lower when compared with other melanoma subtypes.¹¹

The primary aim of this international multicenter study was to investigate the dermatoscopic morphology of NM with a Breslow thickness up to 1mm and identify dermatoscopic predictors for its differential diagnosis from other nodular tumors.

Because only a few NM thinner than 1mm were collected, the threshold for analysis was re-defined to 2mm. Secondary aims were to compare the dermatoscopic features

seen in thin NM (as defined above) with those found in thicker NM, and to separately assess the dermatoscopic morphology of pigmented and non-pigmented NMs and non-melanoma tumors with nodular appearance.

Materials and methods

This was a retrospective, morphological case-control study, conducted on behalf of the International Dermoscopy Society (IDS) following an open call on the IDS website. The Ethics Committee approval was waived, since the study was retrospective, did not affect in any way patient's management and was conducted on anonymized datasets.

Members of the IDS were invited to submit clinical and dermatoscopic images of: i) tumors surgically excised and histopathologically diagnosed as NMs, routinely defined by the presence of a dermal component with no or with a minimal junctional component with a maximum of 3 histopathologically involved rete ridges; or ii) surgically excised tumors with a nodular clinical morphology (with no or minimal flat component), for which melanoma was included in the clinical differential diagnosis, but the histopathologic diagnosis was other than melanoma. Tumors lacking a definite histopathologic diagnosis were not eligible. Members could submit images of tumors fitting to one or both of the above categories. All members submitting NM cases were asked to review the histopathologic slides to ensure that they meet the inclusion criteria. Basic epidemiologic data of the patients (age and sex) were also mandatory. The images were randomized in anonymized datasets.

The selection of the dermatoscopic variables was based on pre-existing literature and an expert consensus held by the IDS particularly for the present study. The variables included in the analysis were divided into 4 categories (overall

architecture and colors, pigmented structures, non-pigmented structures excluding vessels, vascular structures). The analytic list of criteria can be seen in e-figure 1 of the supplementary material.

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The clinical and dermoscopic images were evaluated by 5 independent investigators (DS, CP, HK, AS, SP) who were blinded for the histopathologic diagnosis. The evaluators were asked to assess the presence or absence of pre-defined variables selected as described above. The evaluation was performed using an on-line platform (Survey Monkey) and the images were randomly presented to the evaluators. The statistical analysis was performed on the basis of agreement by at least three of the five evaluators.

Statistical analysis

Absolute and relative frequencies were obtained for the dermoscopic characteristics. For the logistic regression analysis, outcome variables were set to thin NM or other diagnosis. Using the threshold of 1mm of Breslow thickness, the test group (thin NM) consisted of only 12 NM. Because of the large number of variables to be assessed, this sample size was deemed as inadequate for analysis. Therefore, the threshold was changed to 2mm. Relative risks were calculated for the dichotomous variables. Adjusted odds ratios (OR) and corresponding 95% confidence intervals (CI) were calculated by conditional multivariate logistic regression (backward elimination according to likelihood criteria). The alpha level was set at 0.05 while an alpha level of 0.10 was used as the cut-off point for removal of the variable in the automated model selection for multivariate logistic regression. Statistical analyses were performed using IBM SPSS vers. 2 (Armonk, NY, USA).

A subgroup analysis for dermoscopically pigmented versus non-pigmented tumors was performed for each study group. The tumors were classified as pigmented

or not based on the presence of at least one dermatoscopic color corresponding to melanin (black, light brown, dark brown, gray, blue), according to the dermatoscopic evaluation.

Results

Study sample and patients' demographics.

Our study finally included 254 tumors collected from 254 patients. Of them, 69 (27.2%) were NMs ≤ 2 mm Breslow thickness, 96 (37.8%) were thicker than 2mm NMs and 89 (35.0%) were non-melanoma nodular lesions. The latter group included 31 seborrheic keratoses, 28 basal cell carcinomas, 10 dermal nevi, 10 squamous cell carcinomas, 5 benign adnexal tumors, and 5 angiomas. The basic clinical and histopathological features of the 254 tumors and the demographic characteristics of the patients are shown in Table 1. As shown in the table, the sex distribution differed among study groups, with NMs ≤ 2 mm Breslow thickness being more common among females with a female/male ratio of 1.4, whereas thicker NM had a predilection for males with a male/female ratio of 1.63. Regarding anatomic location, NMs ≤ 2 mm Breslow thickness tended to develop on the extremities (60.3%) compared to the group of thicker NM (> 2 mm Breslow thickness) which mostly arose on the trunk (44.7%).

Dermatoscopic features

The analytic results of the dermatoscopic analysis are shown in e-tables 1-4 in the supplement. Out of the 69 NMs ≤ 2 mm Breslow thickness, 50 (72.5%) displayed a dermatoscopic asymmetry of colors and 44 (63.8%) an asymmetric distribution of

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structures. Thicker NMs exhibited color and structure asymmetry in 77.1% and 74.0%, respectively and non-melanomas in 39.3% and 53.9%, respectively. Light brown was the most commonly observed color in NMs up to 2mm of Breslow thickness (50.7%), followed by white (46.4%), blue (44.9%) and pink (43.5%). In thicker NMs, the most frequent colors were blue (64.6%), white (53.1%) and pink (52.1%) (e-table 1).

The most common dermatoscopic structures in NMs up to 2mm of Breslow thickness were irregular brown dots/globules (42.0%), irregular blue structureless area (33.3%), ulceration (29%), white shiny blotches/strands (27.5%), white shiny streaks (26.0%) and irregular (eccentric) black blotch (24.6%). In thicker NMs, the most frequent dermatoscopic structures were ulceration (68.7%), irregular blue structureless area (53.1%), white shiny blotches/strands (41.6%) and white shiny streaks (23.9%) (e-tables 2 and 3). In terms of vascular structures, the most commonly observed morphologic types of vessels in NMs up to 2mm of Breslow thickness were dotted (21.7%) and serpentine (18.8%), whereas in thicker NM, serpentine (43.7%), dotted (24.0%) and corkscrew vessels (17.8%) were the most common. (e-table 4).

The multivariate analysis revealed three dermatoscopic predictors of NMs ≤ 2 mm Breslow thickness as compared to non-melanomas: dotted vessels, white shiny streaks and irregular blue structureless area, which were associated with a 3.4-fold, 2.9-fold and 2.4-fold higher probability for melanoma, respectively (Table 2). In the comparative analysis between thinner and thicker NM, irregular brown dots/globules were found to be a predictor of NM up to 2mm of Breslow thickness (1.8-fold) and serpentine vessels a predictor of thicker NM (3.3-fold).

Subgroup analysis

In the subgroup analysis for pigmented and non-pigmented tumors, the respective numbers of tumors in each group were: 57/69 (82.6%) pigmented and 12/69 (17.4%) non-pigmented NMs $\leq 2\text{mm}$, 75/96 (78.1%) pigmented and 21/96 (21.9%) non-pigmented NMs thicker than 2mm, and 46/89 (51.7%) pigmented and 43/89 (48.3%) non-pigmented non-melanomas. Obviously, dermatoscopic pigmented structures were detected only in the subgroups of pigmented tumors. Of the remaining dermatoscopic criteria, those with the most important differences in frequency between pigmented and non-pigmented NMs are shown in tables 5 and 6 of the supplement.

As shown in the tables, non-pigmented NMs were generally more symmetric, especially in terms of colors. Vascular structures were more frequently detected in non-pigmented NMs as compared to pigmented ones in both groups (thinner and thicker NMs). In the group of NMs $\leq 2\text{mm}$ Breslow thickness, the dotted type was the most frequent morphologic type of vessels in non-pigmented tumors and the serpentine type in pigmented NMs. In the group of thicker NMs, serpentine vessels prevailed in both pigmented and non-pigmented subgroups. A noteworthy finding of the subgroup analysis was the greater frequency of dermatoscopically observed ulceration in non-pigmented NMs $\leq 2\text{mm}$ Breslow thickness (41.7%) as compared to pigmented ones (26.3%). This difference was reversed in the group of thicker NMs, with respective frequencies of 51.3% for non-pigmented tumors and 73.4% for pigmented NMs.

The multivariate analysis was repeated within the subgroups of pigmented and non-pigmented tumors in order to identify predictors of NM $\leq 2\text{mm}$ Breslow thickness as compared to non-melanomas (Table 2). No predictor was found for non-pigmented tumors, whereas irregular blue structureless area represented a melanoma predictor among pigmented tumors (2-fold increased probability).

Discussion

Our study investigated the dermatoscopic morphology of NM at a relatively early phase of its progression based on histopathologic thickness, to identify dermatoscopic predictors for its differential diagnosis from other tumors and to provide insights into the morphologic evolution of NM as tumor thickness increases.

In this descriptive study, demographic data and the clinical characteristics of the tumors revealed findings of interest. The mean age of patients with NM ≤ 2 mm was 8 years younger (52 years) compared to those with thicker tumors (60 years). In addition, patients with thinner NMs were more likely to be female compared with the male-dominated thicker NMs (Table 1). While this difference might be partially attributed to sex-related biologic characteristics, it is more likely to be explained by the increased awareness of the presence of melanoma warning signs for females as compared to males, which has been repeatedly reported in the literature.¹²⁻¹⁶

Additionally, 35% of NMs ≤ 2 mm Breslow thickness and 23% of thicker NMs were clinically non-pigmented. This finding is in line with prior literature suggesting that NM is much more frequently hypo- or amelanotic, as compared to other melanoma subtypes.¹⁷⁻¹⁹ However, the dermatoscopic analysis showed that the percentage of NM without pigmentation was 17.4% for NMs ≤ 2 mm Breslow thickness and 21.9% for thicker NMs, highlighting that dermatoscopy often reveals clinically invisible pigmentation. The latter finding is particularly relevant for clinical practice, since it is known that dermatoscopic pigmented structures usually provide significant diagnostic information.²⁰⁻²²

The descriptive dermatoscopic analysis revealed that NM ≤ 2 mm Breslow thickness is usually a polychromatic tumor, combining 2 or more of light brown, blue,

white and pink color (Figure 1). However, in line with previous publications, we found that an important proportion of NMs (up to 30%) remain symmetric in terms of dermatoscopic colors and structures.^{2, 8-10, 23, 24}

Irregular brown dots/globules, irregular blue structureless area, ulceration, white shiny blotches/strands and white shiny streaks are the most frequently seen dermatoscopic structures in the group of thin NMs (Figure 1). In contrast, atypical network, regression structures and irregular streaks were rarely found in our overall sample of NM. The latter finding is consistent with pre-existing literature^{8, 10, 25} and is explained by the fact that these features correspond to histopathologic alterations at the level of the dermo-epidermal junction, which are absent or minimal in NM.^{11, 26, 27} In terms of vascular structures, dotted vessels and serpentine vessels were the most frequent morphologic types found in NMs up to 2mm of Breslow thickness.

The multivariate analysis revealed 3 important predictors of NM ≤ 2 mm Breslow thickness as compared to non-melanomas: dotted vessels, white shiny streaks/lines and irregular blue structureless area (Figure 1). A comparison of these results with previously reported findings is problematic because of the scarce published evidence. The largest previous case-control study that included 83 cases of NM reported numerous univariate predictors of NM, but a multivariate logistic regression analysis was not performed.⁸

Dotted vessels can be found in several benign and malignant skin tumors and inflammatory skin diseases, including common nevi, Spitz nevi, dermatofibromas, lichenoid keratoses, Bowen's disease, superficial basal cell carcinomas when located on the legs, psoriasis, eczema and many other entities.²⁸⁻³³ They have also been reported in flat pigmented and amelanotic melanoma.^{10, 34} However, almost all of the aforementioned conditions manifest as clinically flat lesions, whereas dotted vessels

in nodular tumors have not been frequently described. Clear cell acanthoma represents an exception, but it is dermatoscopically characterized by a peculiar reticular arrangement of dotted vessels.³⁵ Considering that dotted vessels represented the strongest predictor of $NM \leq 2\text{mm}$ thickness in our analysis, our data combined with pre-existing evidence strongly suggest that a nodule displaying dotted vessels should be considered as highly suspicious for NM (Figure 1).^{8, 36, 37}

White shiny streaks/lines have been previously suggested to represent a melanoma-specific structure and our findings support this also for $NM \leq 2\text{mm}$ Breslow thickness.³⁸ White shiny blotches/strands were also frequently detected, but their diagnostic significance was lost in the multivariate analysis. This is because white shiny blotches/strands can also be found in basal cell carcinoma, which represented a significant part of our control group (Figure 1).³⁹

Irregular blue structureless area was an independent multivariate predictor of $NM \leq 2\text{mm}$ thickness in the overall set, as well as in the subgroup analysis for pigmented tumors only (Figure 1). The presence of blue pigmentation in NM had been reported in previous studies.^{7, 8, 40-41} In one of them, it was suggested that the combination of blue and black color within a nodular lesion is strongly suggestive of NM (“blue-black rule”).¹¹ The results of the present study confirm that a positive “blue-black” rule is highly specific for NM, as compared to non-melanomas. However, only a small minority of NMs $\leq 2\text{ mm}$ Breslow thickness displayed both blue and black color (13%).

Our comparative analysis between $NM \leq 2\text{mm}$ Breslow thickness and thicker NMs may provide insights into the evolution of NM from a dermatoscopic point of view. A reasonable finding of the latter analysis was that the proportion of NMs that retain a dermatoscopic symmetry decreases as the thickness of NM increases. In terms

of dermatoscopic colors, the most important difference was the predominance of light brown color in thinner NMs and blue color in thick NMs. The most striking difference between thinner and thicker NMs in terms of dermatoscopic structures was the predominance of irregular brown dots/globules in NMs ≤ 2 mm Breslow thickness (Figure 1), as compared to their much lower frequency in thicker NMs. Concerning vascular morphologic types, dotted vessels prevailed in NMs ≤ 2 mm Breslow thickness (Figure 1) and serpentine vessels in the group of thicker tumors (Figure 2). Corkscrew vessels were also much more frequent in thicker NMs, possibly corresponding to the increased tumor neo-angiogenesis (Figure 2). These findings are consistent with pre-existing literature on the vascular pattern of melanoma.^{8, 28, 36, 42-45} In summary, the main dermatoscopic changes in NM as thickness increases are that brown color becomes less evident and blue more prominent, dermatoscopically observed ulceration increases and dotted vessels are replaced by serpentine and highly tortuous linear vessels (Figure 2).

The subgroup analysis of non-pigmented and pigmented tumors revealed some interesting findings. First, non-pigmented NMs were much more frequently symmetric as compared to pigmented ones, especially in terms of color. This lack of asymmetry is consistent with previous evidence and reflects a common problem in everyday practice leading to delayed diagnosis or inappropriate treatment.⁴⁶ In contrast, most of the pigmented NMs displayed a dermatoscopic asymmetry of colors and structures. The latter is consistent with 2 previous case-control studies on pigmented NM that reported a high frequency of asymmetric pigmentation.^{9, 26}

Our study represents the largest collection of images from NM of different tumor thicknesses, including thinner tumors of Breslow less than 2 mm. Even though it includes a highly selected case series with potential selection bias and, in the

absence of a central pathology review, carries the potential of diagnostic misclassification, a problem often seen with the diagnosis of nodular melanoma, our study identifies important morphologic criteria that can help distinguish this particular subtype from non-melanoma nodular lesions and even help to identify thinner NM.

In conclusion, our study confirms the insidious nature of a NM at its early stages, suggests irregular brown dots/globules, irregular blue structureless area, white shiny streaks/lines and dotted vessels as useful dermoscopic clues for its clinical diagnosis and sheds light into the morphologic evolution of the tumor. Considering the scant available evidence on the topic, our findings need to be further elucidated, but may assist in the early recognition of this aggressive type of melanoma.

References

1. Siegel RL, Miller KD, Jemal A. Cancer statistics. *CA Cancer J Clin* 2019;69(1):7-34. doi: 10.3322/caac.21551. Epub 2019 Jan 8.
2. Klebanov N, Gunasekera N, Lin W et al. Clinical spectrum of cutaneous melanoma morphology. *J Am Acad Dermatol* 2019; 80(1):178-188.e3. doi: 10.1016/j.jaad.2018.08.028.
3. Weir H.K.,Thompson TD, Soman A, Møller B, Leadbetter S. The past, present, and future of cancer incidence in the United States: 1975 through 2020. *Cancer* 2015;121(11):1827-37. doi: 10.1002/cncr.29258. Epub 2015 Feb 3.
4. Chamberlain A, Giles G, Dowling J, Kelly J. Nodular Type and Older Age as the Most Significant Associations of Thick Melanoma in Victoria, Australia. *Arch Dermatol* 2002;138(5):609-14
5. Demierre MF, Chung C, Miller D, Geller A. Early Detection of Thick Melanomas in the United States. *Arch Dermatol* 2005;141(6):745-506. Lattanzi M, Lee Y, Simpson D, et al. Primary Melanoma Histologic Subtype: Impact on Survival and Response to Therapy. *J Natl Cancer Inst* 2019;111(2):180. doi: 10.1093/jnci/djy086
6. Lattanzi M, Lee Y, Simpson D, et al. Primary Melanoma Histologic Subtype: Impact on Survival and Response to Therapy. *J Natl Cancer Inst* 2019;111(2):180. doi: 10.1093/jnci/djy086
7. Rosendahl C, Hishon M, Cameron A, Barksdale S, Weedon D, Kittler H. Nodular melanoma: five consecutive cases in a general practice with polarized and non-polarized dermatoscopy and dermatopathology. *Dermatol Pract Concept* 2014; 4(2):69-75. doi: 10.5826/dpc.0402a15. eCollection 2014 Apr

8. Menzies SW, Moloney FJ, Bythe K et al. Dermoscopic evaluation of nodular melanoma. *JAMA Dermatol* 2013; 149(6):699-709. doi: 10.1001/jamadermatol.2013.2466.
9. Kalkhoran S, Milne O, Zalaudek I et al. Historical, clinical, and dermoscopic characteristics of thin nodular melanoma. *Arch Dermatol* 2010; 146(3):311-8. doi: 10.1001/archdermatol.2009.369.
10. Pizzichetta MA, Kittler H, Stanganelli I et al. Pigmented nodular melanoma: The predictive value of dermoscopic features using a multivariate analysis. *Br J Dermatol* 2015; 173(1):106-14. doi: 10.1111/bjd.13861. Epub 2015 Jun 2
11. Argenziano G, Longo C, Cameron A et al. Blue-black rule: a simple dermoscopic clue to recognize pigmented nodular melanoma. *Br J Dermatol* 2011;165: 1251-1255. doi: 10.1111/j.1365-2133.2011.10621.x.
12. Schwartz JL, Wang TS, Hamilton TA, Lowe L, Sondak VK, Johnson TM. Thin primary cutaneous melanomas: associated detection patterns, lesion characteristics, and patient characteristics. *Cancer* 2002;95(7):1562-1568. Doi: 10.1002/cncr.10880
13. Swetter S, Layton C, Johnson T, Brooks K, Miller D, Geller A. Gender differences in melanoma awareness and detection practices between middle-aged and older men and their female spouses. *Arch Dermatol* 2009;145(4):488-490. Doi: 10.1001/archdermatol.2009.42
14. Lee KC, Higgins HW 2nd, Lindon O, Cruz AP. Gender differences in tumor and patient characteristics in those undergoing Mohs surgery. *Dermatol Surg* 2014;40(6):68 6-690. Doi: 10.1111/dsu.0000000000000020
15. Al-Dujaili Z, Henry M, Dorizas AS, Sadick NS. Skin cancer concerns particular to women. *Int J Womens Dermatol* 2017;3(1 Suppl): S49-S51. Doi: 10.1016/j.ijwd.2017.02.009

16. Buchanan Lunsford N, Berkthold J, Holman D, Stein K, Prempeh A, Yerkes A. Skin cancer knowledge, awareness, beliefs and preventive behaviors among black and hispanic men and women. *Prev Med Rep* 2018;12: 203-209 doi: 10.1016/j.pmedr.2018.09.017
17. Corneli P, Zalaudek I, Magaton-Rizzi G, Di Meo N. Improving the early diagnosis of early nodular melanoma: can we do better? *Expert Rev Anticancer Ther* 2018;18: 1007-1012. doi: 10.1080/14737140.2018.1507822. Epub 2018 Aug 13.
18. Wee E, Wolfe R, Mclean C, Kelly JW, Pan Y. Clinically amelanotic or hypomelanotic melanoma: anatomic distribution, risk factors, and survival. *J Am Acad Dermatol* 2018;79(4):645-651.e4 doi: 10.1016/j.jaad.2018.04.045
19. Gong HZ, Zheng HY, Li J. Amelanotic melanoma. *Melanoma Res* 2019;29(3):221-230. Doi: 10.1097/CMR.0000000000000571
20. Kittler H, Pehamberger H, Wolff K, Binder M. Diagnostic accuracy of dermoscopy. *Lancet Oncol* 2002;3(3):159-165
21. Rosendhal C, Tschandl P, Cameron A, Kittler H. Diagnostic accuracy of dermoscopy for melanocytic and non-melanocytic pigmented lesions. *J Am Acad Dermatol* 2011;64(6):1068-1073. Doi: 10.1016/j.jaad.2010.03.039
22. Lallas A, Argenziano G, Kyrgidis A, et al. Dermoscopy uncovers clinically undetectable pigmentation in basal cell carcinoma. *Br J Dermatol* 2014;170(1):192-195 doi: 10.1111/bjd.12634
23. Warycha MA, Christos PJ, Mazumdar M, et al. Changes in the presentation of nodular and superficial spreading melanomas over 35 years. *Cancer* 2008;113(12):3341-3348 doi: 10.1002/cncr.23955
24. Dessinioti C, Dimou N, Geller A, et al. Distinct clinicopathological and prognostic features of thin nodular primary melanomas: an international study from

17 centers. *J Natl Cancer Inst* 2019 pii: djz034. doi: 10.1093/jnci/djz034. [Epub ahead of print]

25. Fargnoli MC, Sera F, Suppa M, et al. Dermoscopic features of cutaneous melanoma are associated with clinical characteristics of patients and tumours and with MC1R genotype. *J Eur Acad Dermatol Venereol* 2014;28(12):1768-1775 doi: 10.1111/jdv.12411

26. Segura S, Pellacani G, Puig S et al. In vivo microscopic features of nodular melanomas: dermoscopy, confocal microscopy, and histopathologic correlates. *Arch Dermatol* 2008;144: 1311-1320. doi: 10.1001/archderm.144.10.1311. Erratum in: *Arch Dermatol*. 2009 May;145(5):556.

27. Russo T, Piccolo V, Ferrara G, et al. Dermoscopy, pathology correlation in melanoma. *J Dermatol* 2017;44(5):507-514 doi: 10.1111/1346-8138.13629

28. Zalaudek I, Kreusch J, Giacomel J Ferrara G, Catricalà C, Argenziano G. How to diagnose nonpigmented skin tumors: A review of vascular structures seen with dermoscopy: part II. Nonmelanocytic skin tumors *J Am Acad Dermatol* 2010;63(3):377-86; quiz 387-8. doi: 10.1016/j.jaad.2009.11.697.

29. Lallas A, Apalla Z, Ioannides D, et al. Update on dermoscopy of Spitz/Reed naevi and management guidelines by the International Dermoscopy Society. *Br J Dermatol* 2017;177(3):645-655 doi: 10.1111/bjd.15339

30. Ferrari A, Argenziano G, Buccini P, et al. Typical and atypical dermoscopic presentations of dermatofibroma. *J Eur Acad Dermatol Venereol* 2013;27(11):1375-1380 doi: 10.1111/jdv.12019

31. Liopyris K, Navarette-Dechent C, Dusza S, et al. Clinical and dermoscopic features associated with lichen planus-like keratoses that undergo skin biopsy: a

single center observational biopsy. *Australas J Dermatol* 2019;60(2):e119-e126 doi: 10.1111/ajd.2955

32. Papageorgiou C, Apalla Z, Variaah G, et al. Accuracy of dermoscopic criteria or the differentiation between superficial basal cell carcinoma and Bowen's disease. *J Eur Acad Dermatol Venereol* 2018;32(11):1914-1919 doi: 10.1111/jdv.14995

33. Sgouros D, Apalla Z, Ioannides D, et al. Dermoscopy of common inflammatory disorders. *Dermatol Clin* 2018;36(4):359-368 doi: 10.1016/j.det.2018.05.003

34. Jaimes N, Braun RP, Thomas L, Marghoob AA. Clinical and dermoscopic characteristics of amelanotic melanomas that are not of the nodular subtype. *J Eur Acad Dermatol Venereol* 2012;26(5):591-596 doi: 10.1111/j.1468-3083.2011.04122

35. Todorovic-Zivkovic D, Lallas A, Longo C, Moscarella E, Zalaudek I, Argenziano G. Dermoscopy of clear cell acanthoma. *J Am Acad Dermatol* 2015;72(1 Suppl):S47-49 doi: 10.1016/j.jaad.2014.06.039

36. Menzies S, Kreusch J, Byth K, et al. Dermoscopic evaluation of amelanotic and hypomelanotic melanoma. *Arch Dermatol* 2008;144(9):1120-1127 doi: 10.1001/archderm.144.9.1120

37. Corneli P, Zalaudek I, Magaton-Rizzi G, Di Meo N. Improving the early diagnosis of early nodular melanoma: can we do better? *Expert Rev Anticancer Ther* 2018;18: 1007-1012. doi: 10.1080/14737140.2018.1507822.

38. Corneli P, Zalaudek I, Magaton-Rizzi G, Di Meo N. Improving the early diagnosis of early nodular melanoma: can we do better? *Expert Rev Anticancer Ther* 2018;18: 1007-1012. doi: 10.1080/14737140.2018.1507822.

39. Navarette-Dechent C, Bajaj S, Marchetti MA, Rabinovitz H, Dusza SW, Marghoob AA. Association of shiny white blotches and strands with nonpigmented

basal cell carcinoma: Evaluation of an additional dermoscopic diagnostic criterion.

JAMA Dermatology 2016;152(5): 546-552. doi: 10.1001/jamadermatol.2015.573144.

40. Longo C, Farnetani F, Moscarella E et al. Can noninvasive imaging tools

potentially predict the risk of ulceration in invasive melanomas showing blue and

black colors? *Melanoma Res* 2013;23(2): 125-131. doi:

10.1097/CMR.0b013e32835d90b8

41. Longo C, Scope A, Lallas A, et al. Blue lesions. *Dermatol Clin* 2013;31(4):637-

647 doi:10.1016/j.det.2013.07.001

42. Rosendahl C, Cameron A, Tschandl P, Bulinska A, Zalaudek I, Kittler H.

Prediction without pigment: a decision algorithm for non-pigmented skin malignancy.

Dermatol Pract Concept 2014;4(1):59-66 doi: 10.5826/dpc.0401a09

43. De Carvalho N, Welzel J, Schuh S, et al. The vascular morphology of melanoma

is related to Breslow index: an in vivo study with dynamic optical coherence

tomography. *Exp Dermatol* 2018;27(11):1280-1286 doi: 10.1111/exd.13783

44. Zalaudek I, Kreusch J, Giacomel J Ferrara G, Catricalà C, Argenziano G. How to

diagnose nonpigmented skin tumors: A review of vascular structures seen with

dermoscopy: part I. Melanocytic skin tumors *J Am Acad Dermatol* 2010;63(3):361-

374;quiz 375-376 doi: 10.1016/j.jaad.2009.05.035

45. Pizzichetta M, Kittler H, Stanganelli I, et al. Dermoscopic diagnosis of

melanotic/hypomelanotic melanoma. *Br J Dermatol* 2017;177(2):538-540

doi:10.1111/bjd.15093

46. Moloney F, Menzies S. Key points in the dermoscopic diagnosis of

hypomelanotic melanoma and nodular melanoma. *J Dermatol* 2011;38(1):10-15 doi:

10.1111/j.1346-8138.2010.01140.x

Figure Legends

e-figure 1: Analytic list of the dermatoscopic criteria for each study group

Figure 1: Nodular Melanoma (NM) with a Breslow thickness ≤ 2 mm. **A**, Irregular brown dots and globules (white arrows) at the periphery of a 1.7mm Breslow thickness melanoma along with white shiny strands and blue structureless area (white circle) in the center of the lesion. **B**, A thin NM (1.5 mm Breslow thickness) with an eccentric black blotch (white arrow), irregular light brown coloration (white star) and white shiny structures (black arrow). **C**, Dotted vessels (white circles) prevail in this 0.5 mm Breslow thickness NM along with an irregular blue structureless area (white arrow). **D**, Serpentine vessels (white arrows) are prominent in this hypopigmented thin NM (1.1mm Breslow thickness) with scarce light brown coloration (white circle) at the periphery.

Figure 2: Nodular Melanoma (NM) with a Breslow thickness >2 mm. **A**, Irregular blue-white structureless areas (white circle) prevail at the upper part of this 3mm Breslow thickness NM, while pink-whitish coloration with a combination of serpentine vessels (black circle) are prominent at the lower part of the lesion. **B**, Areas of ulceration (white circle) along with serpentine vessels prevail in this amelanotic NM (4.3 mm Breslow thickness). **C**, At the same context with B ulceration (white arrow) is also striking in this advanced NM (7mm Breslow thickness). On the contrary this tumor is heavily pigmented with blue-black coloration (white circle). **D**, Corkscrew vessels (white circle) can be seen in the amelanotic part of the lesion along with serpentine vessels (black arrow). At the right part an area of blue-black pigmentation can be observed with a combination of a whitish hue (white arrow) in this 2.1 mm Breslow thickness NM.