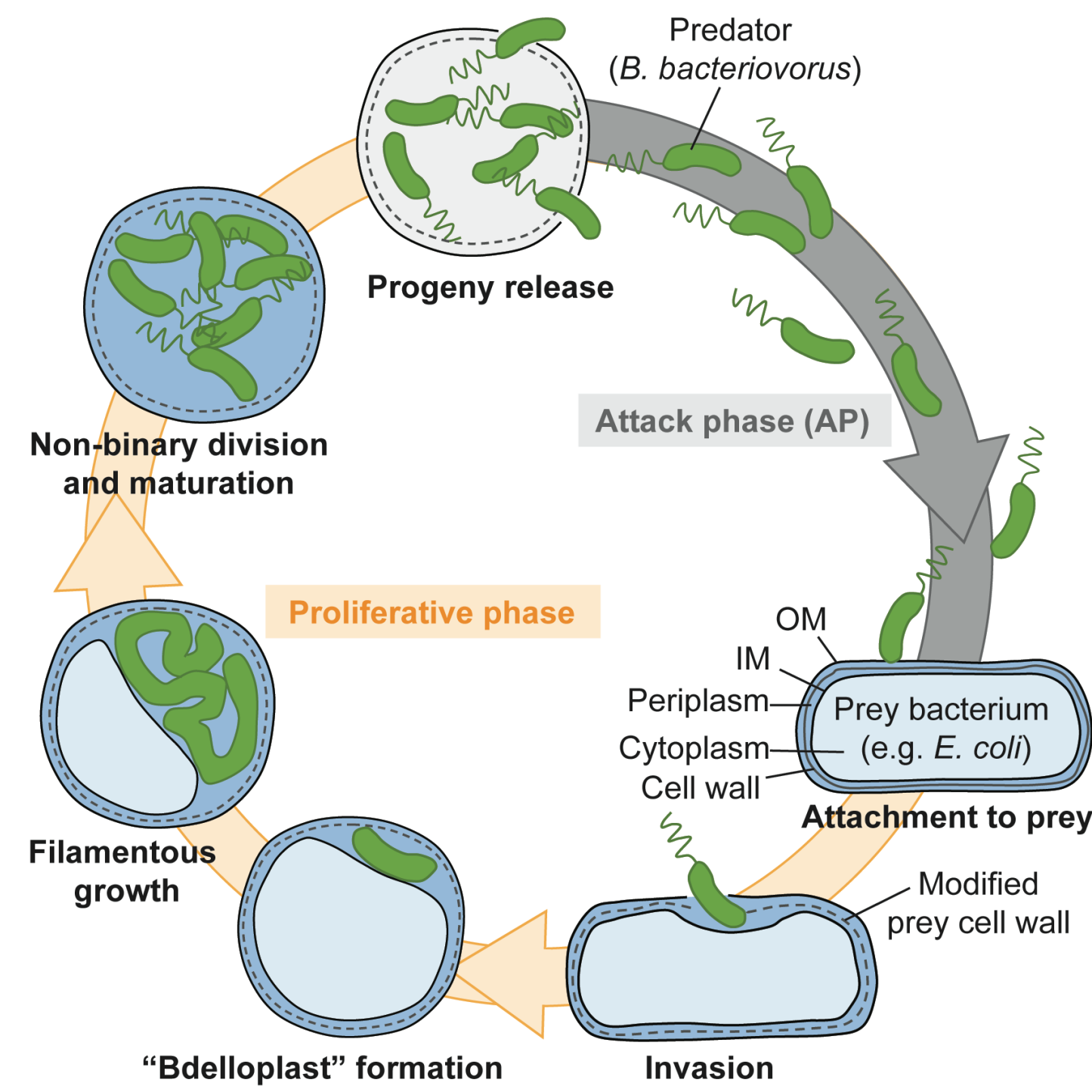


Uncovering the choreography of the chromosome during the non-binary cell cycle of the predatory bacterium *Bdellovibrio bacteriovorus*

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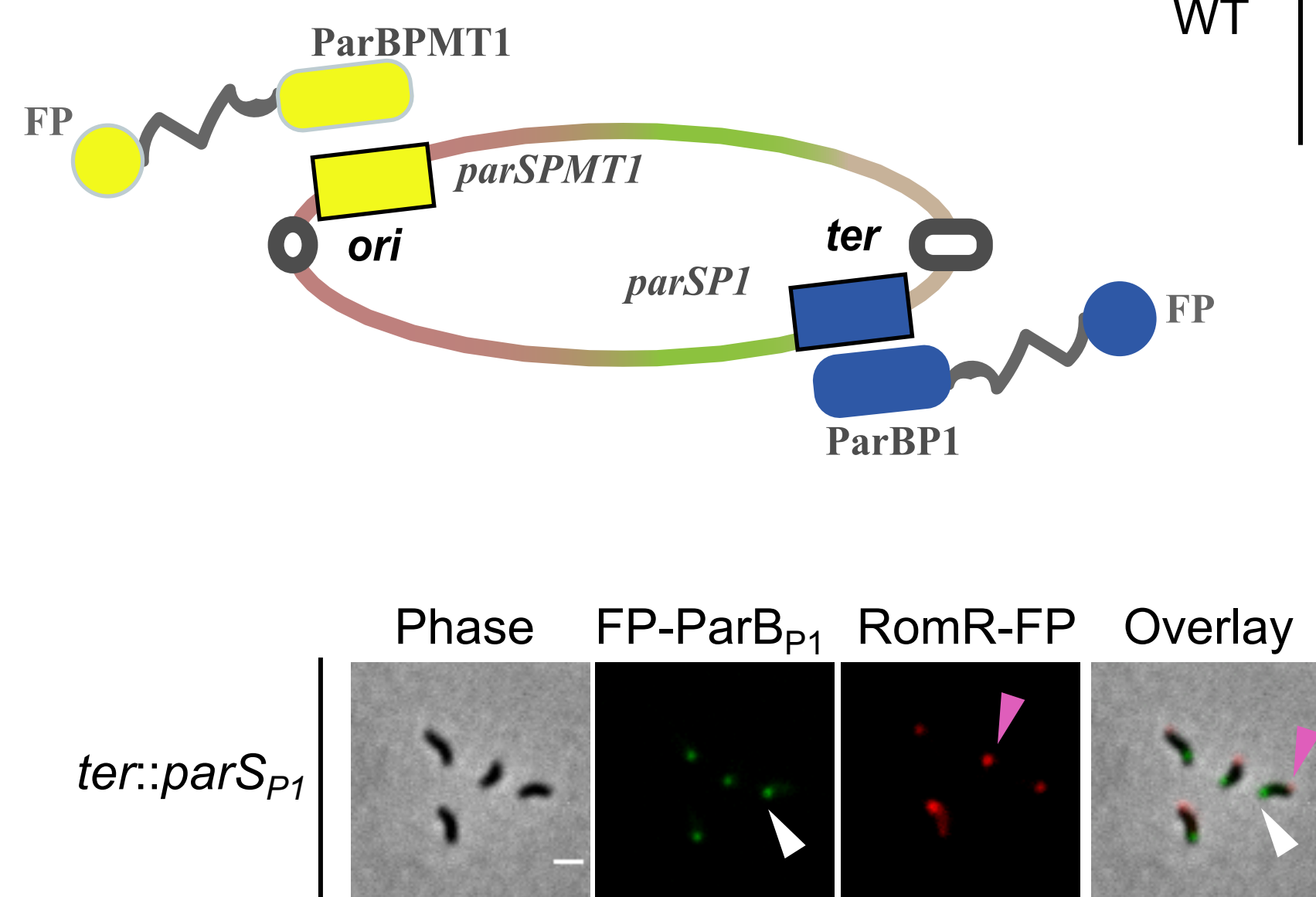
Bdellovibrio bacteriovorus is a predatory delta-proteobacterium that invades the periplasm of other Gram-negative bacteria. Once the *Bdellovibrio* cells are inside, the prey bacterium forms a structure termed “bdelloplast” whereas the predator grows as a filament before dividing synchronously into a variable number of monoploid cells by a non-binary division process. **Molecular understanding of how this micro-predator proliferates inside the envelope of other bacteria is still missing.**

Main questions:

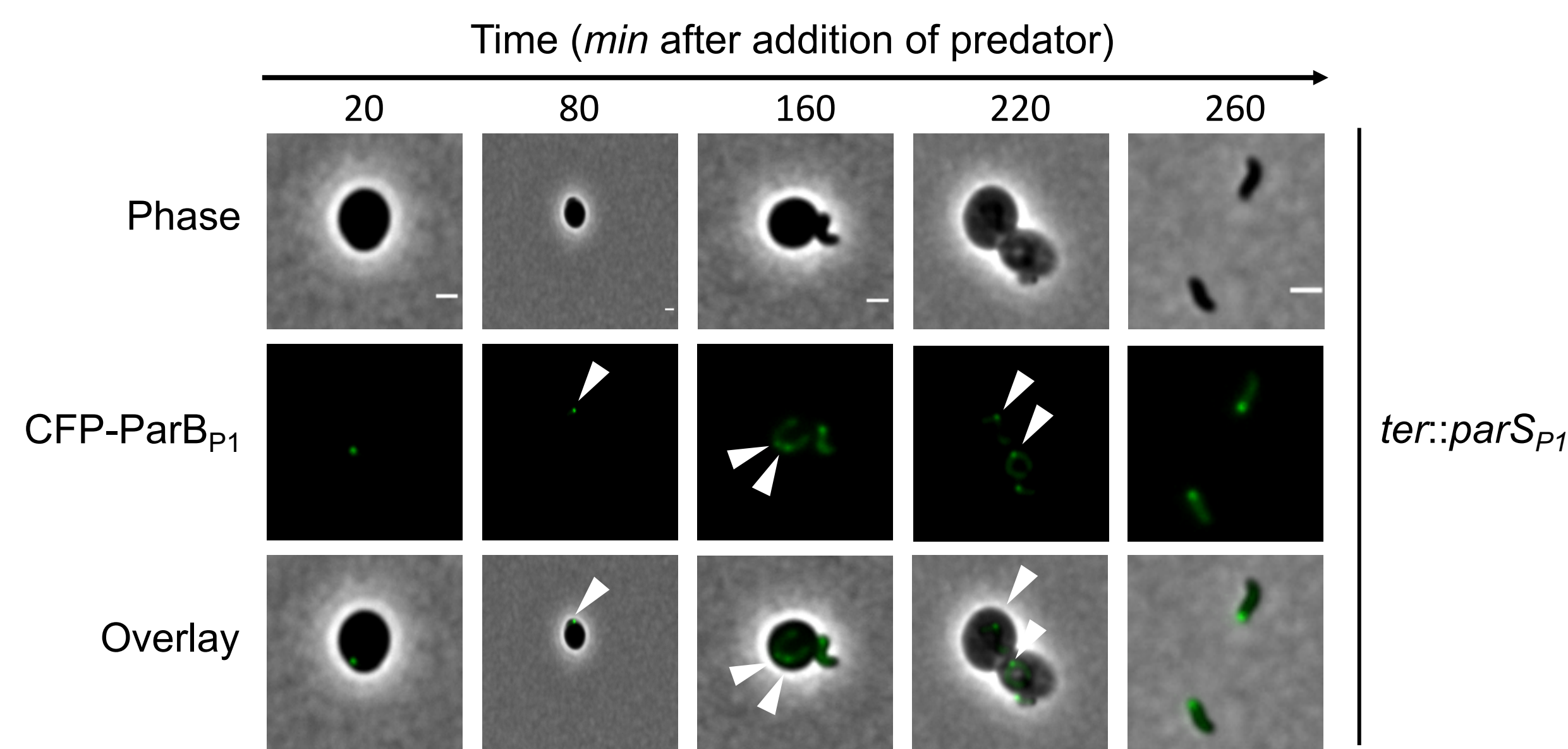
1. How are chromosomal regions positioned throughout the cell cycle of *Bdellovibrio*?
2. When and where are the chromosomal copies segregated during filamentous growth?

1. Spatial arrangement of the *Bdellovibrio* chromosome

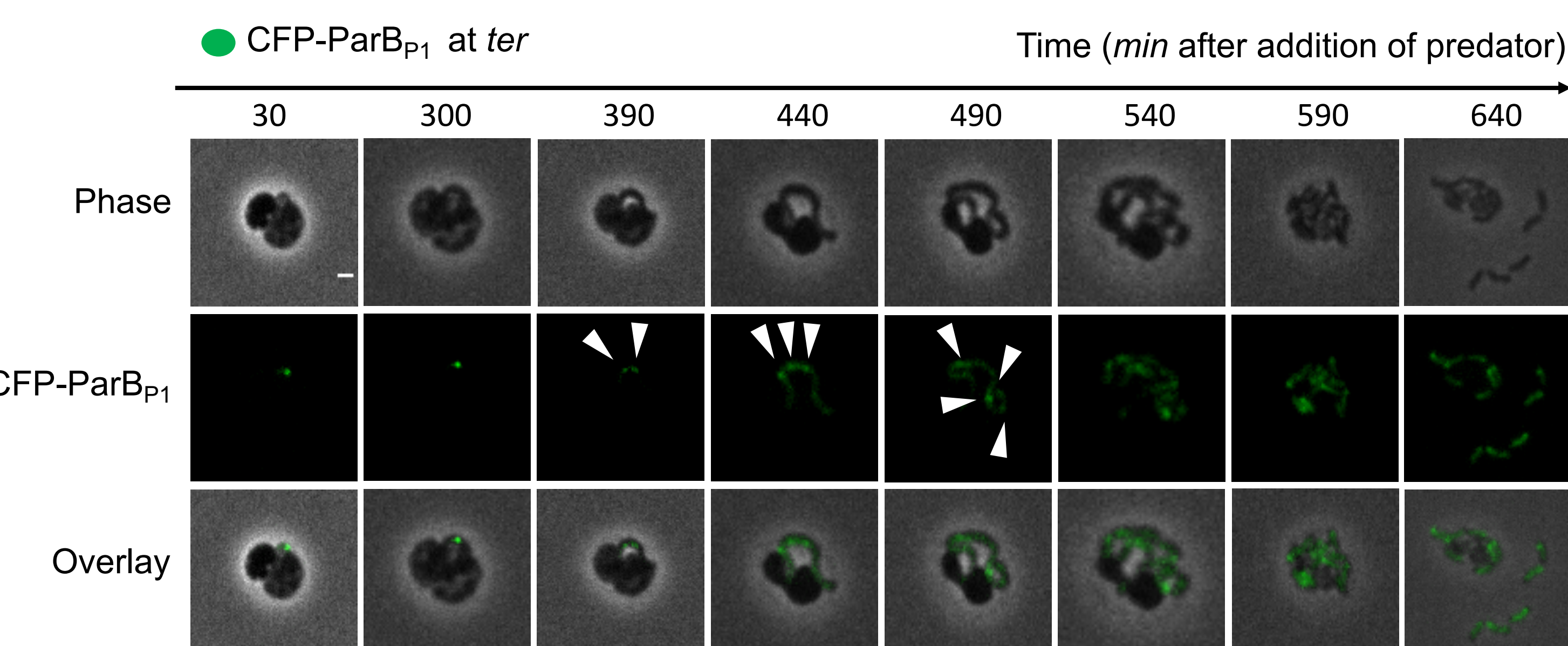
The spatio-temporal arrangement of the *Bdellovibrio* chromosome remains unknown. **We are investigating this feature using the orthogonal ParB-*parS* system.**



The *ter* locus is positioned at the flagellated pole in the attack phase of the cell cycle (arrowheads), in contrast with previously reported *ter* localization in other flagellated species. Colocalization with RomR-TdTomato (pink arrowheads) confirms that *ter* is at the non-invasive pole. Scale bar: 1 μ m. Representative cells are shown.



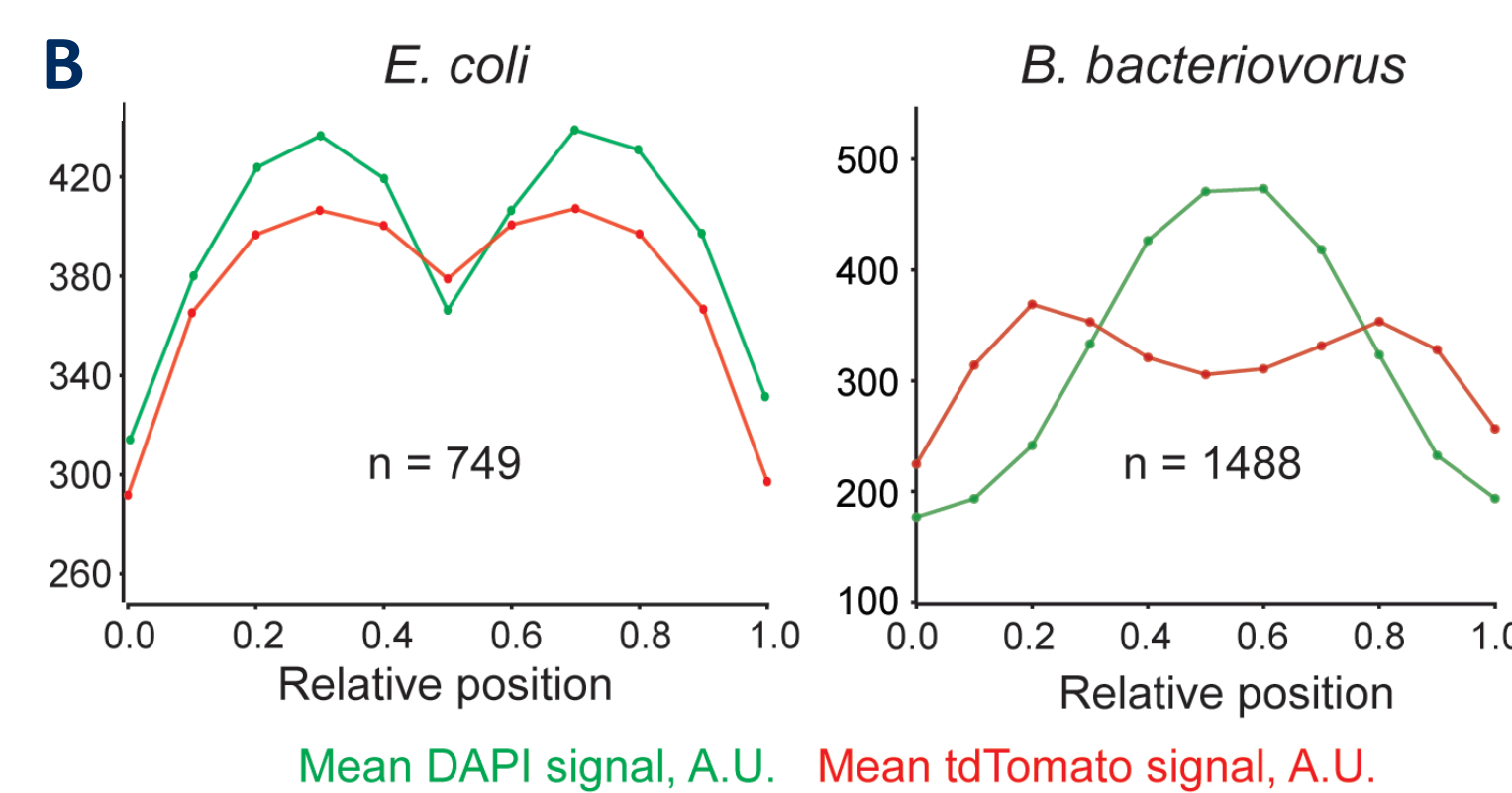
Synchronous predation of *E. coli* by *B. bacteriovorus* HD100 *ter::parSP1* with constitutive expression of CFP-ParBP1, used as a proxy for the visualization of terminus of replication (arrowheads).



The *ter* locus is polar at early stages after invasion, then *ter* moves towards the midcell point, where multiplication of *ter* takes place (arrowheads). Scale bar: 1 μ m. Representative cells are shown.

2. Dynamics of the *Bdellovibrio* chromosome

A) Our observations also reveal intriguing dynamics of the level of nucleoid compaction in *Bdellovibrio*. After a certain period without the prey, polar localization of FP-ParBP1 was lost (72h, inset). We hypothesize that this is due to DNA condensation preventing ParB binding to *parS* at *ter*. When mixed with prey, signal was again visible (bottom panel, arrowheads)

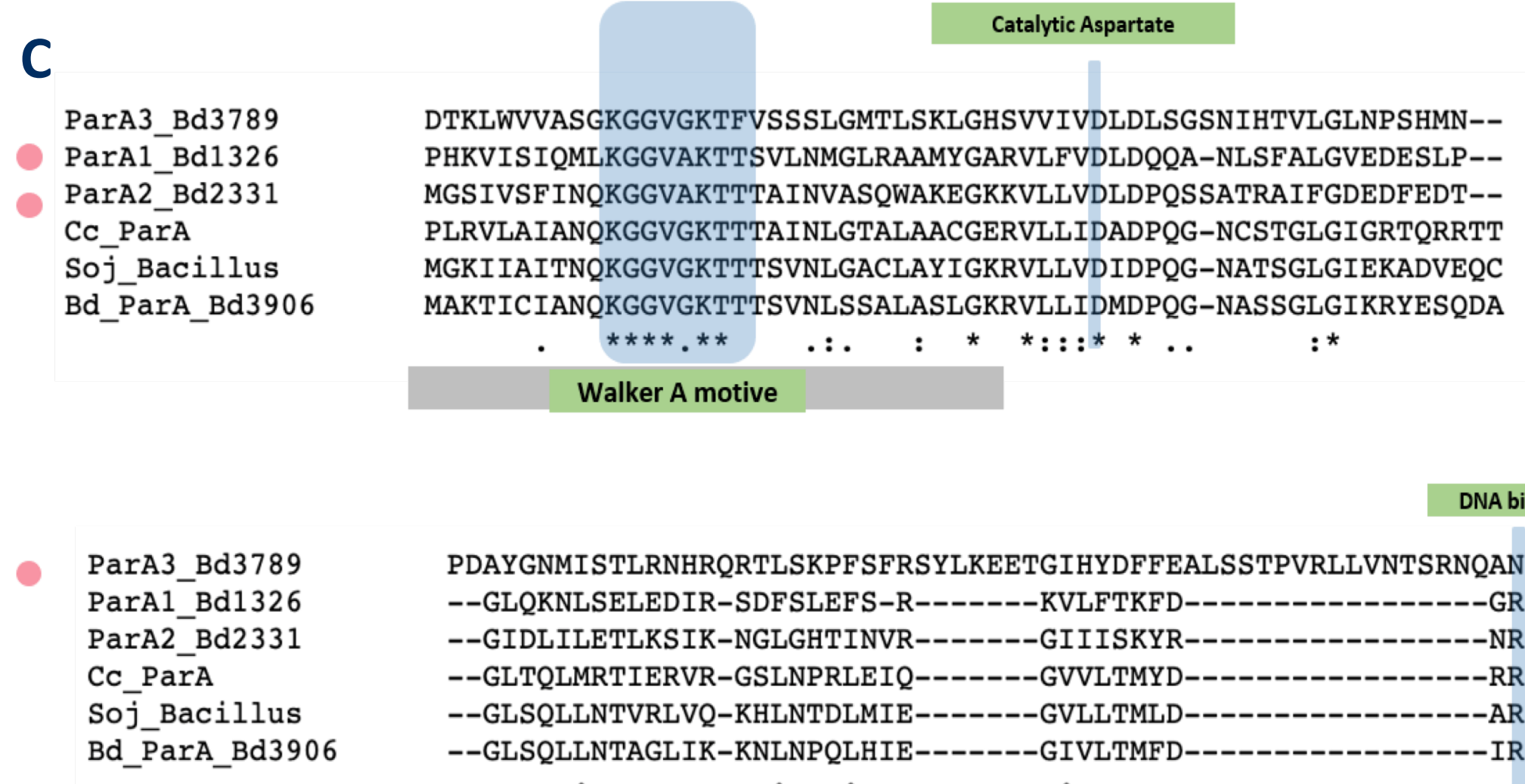
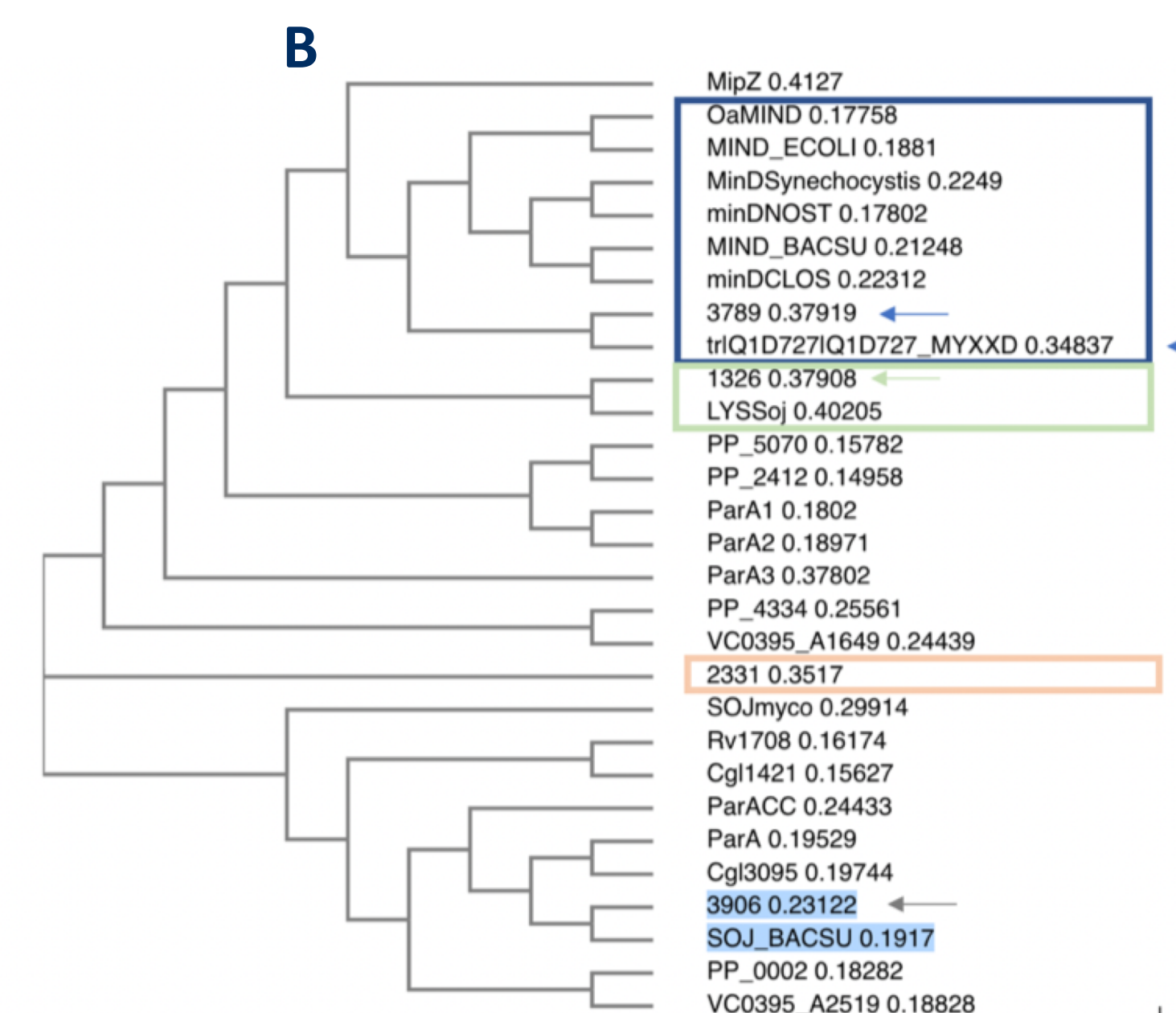
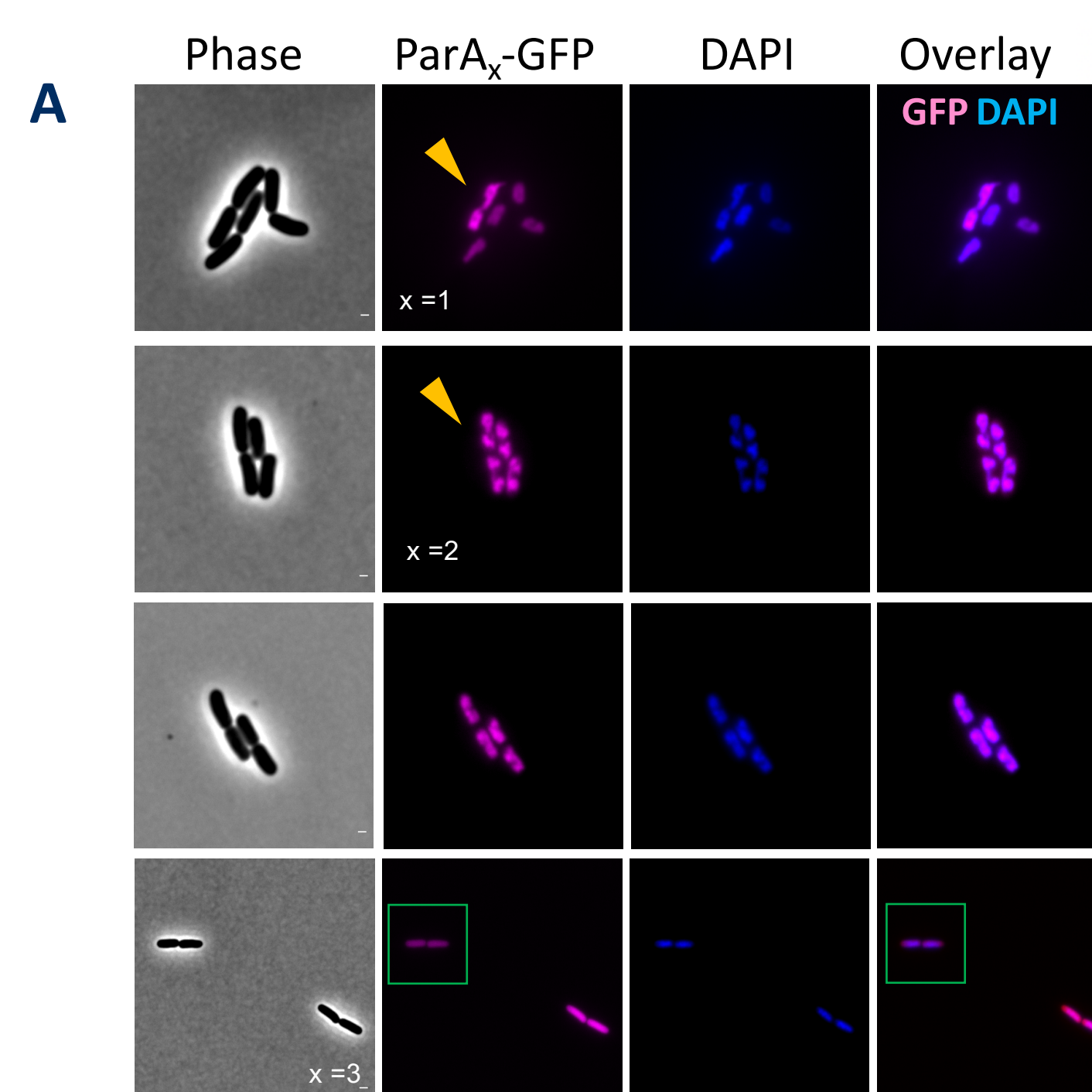


B) Average pole-to-pole profiles from *Bdellovibrio* attacking phase cells, showing nucleoid occlusion of tdTomato in *B. bacteriovorus*.

These intriguing data suggest that *Bdellovibrio* cells can “sense” the prey via an unknown mechanism that modifies nucleoid compaction. Scale bar: 1 μ m. Representative cells are shown.

3. Segregation of the *Bdellovibrio* chromosome

In most species, chromosome segregation is driven by the ParABs system. Interestingly, in addition to the canonical ParA, we found **three additional ParA-like proteins** encoded in the *Bdellovibrio* genome.



Analyses of *Bdellovibrio* ParA homologs (Figure B). Alignment of ParAs sequences (Figure C). Key motifs are highlighted in blue. Corresponding names are in green.

Walker A motifs in ParA1 and ParA2 are different from known ParA's (Figure C). ParA3 does not harbor the typical DNA binding residue (Figure A and C).

ParA1 and ParA2 colocalize and affect chromosome segregation/compaction when expressed in *E. coli* (yellow arrowheads). ParA3 has a diffused pattern when produced in *E. coli* (green inset), consistent with our prediction from sequence analysis.