

Cardiac dysfunction in cancer patients: beyond direct cardiomyocyte damage of anticancer drugs: novel cardio-oncology insights from the joint 2019 meeting of the ESC Working Groups of Myocardial Function and Cellular Biology of the Heart

Carlo Gabriele Tocchetti ^{1,2,*†}, Pietro Ameri ^{3,4†}, Rudolf A. de Boer ⁵, Yuri D'Alessandra⁶, Michele Russo ^{1,7}, Daniela Sorriento ⁸, Michele Ciccarelli ⁹, Bernadett Kiss¹⁰, Luc Bertrand ¹¹, Dana Dawson ¹², Ines Falcao-Pires¹³, Mauro Giacca ^{14,15,16}, Nazha Hamdani ^{17,18}, Wolfgang A. Linke ¹⁹, Manuel Mayr ¹⁶, Jolanda van der Velden²⁰, Serena Zacchigna ^{14,15}, Alessandra Ghigo ⁷, Emilio Hirsch ⁷, Alexander R. Lyon ²¹, Anikó Görbe^{10,22}, Péter Ferdinandy^{10,22}, Rosalinda Madonna ^{23,24}, Stephane Heymans^{25,26}, and Thomas Thum ^{27*}

¹Department of Translational Medical Sciences, Federico II University, via Pansini 5, 80131 Naples, Italy; ²Interdepartmental Center of Clinical and Translational Sciences (CIRCET), Federico II University, Naples, Italy; ³Cardiovascular Disease Unit, IRCCS Ospedale Policlinico San Martino, Genova, Italy; ⁴Department of Internal Medicine, University of Genova, Genova, Italy; ⁵Department of Cardiology, University of Groningen, University Medical Center Groningen, AB31, PO Box 30.001, 9700 RB Groningen, The Netherlands; ⁶Immunology and Functional Genomics Unit, Centro Cardiologico Monzino IRCCS, Milan, Italy; ⁷Department of Molecular Biotechnology and Health Sciences, Molecular Biotechnology Center, University of Torino, Torino, Italy; ⁸Department of Advanced Biomedical Sciences, Federico II University, Naples, Italy; ⁹Department of Medicine Surgery and Odontology, University of Salerno, Salerno, Italy; ¹⁰Department of Pharmacology and Pharmacotherapy, Cardiometabolic Research Group and MTA-SE System Pharmacology Research Group, Semmelweis University, Budapest, Hungary; ¹¹IREC Institute, Pole of Cardiovascular Research, Université Catholique de Louvain, Brussels, Belgium; ¹²School of Medicine and Dentistry, University of Aberdeen, Aberdeen, UK; ¹³Unidade de Investigação e Desenvolvimento Cardiovascular, Departamento de Cirurgia e Fisiologia, Faculdade de Medicina, Universidade do Porto, Portugal; ¹⁴Department of Medicine, Surgery and Health Sciences and Cardiovascular Department, Centre for Translational Cardiology, Azienda Sanitaria Universitaria Integrata Trieste, Trieste, Italy; ¹⁵International Center for Genetic Engineering and Biotechnology (ICGEB), Trieste, Italy; ¹⁶King's British Heart Foundation Centre, King's College London, London, UK; ¹⁷Department of Molecular and Experimental Cardiology, Ruhr Universität Bochum, Bochum, Germany; ¹⁸Department of Cardiology, St. Joseph Hospital, Ruhr University Bochum, Witten, Germany; ¹⁹Institute of Physiology 2, University of Muenster, Germany; ²⁰Department of Physiology, Amsterdam UMC, Vrije Universiteit, Amsterdam Cardiovascular Sciences Institute, Amsterdam, The Netherlands; ²¹Cardio-Oncology Service, Royal Brompton Hospital, Imperial College London, London, UK; ²²Pharmahungary Group, Szeged, Hungary; ²³Institute of Cardiology, University of Pisa, Pisa, Italy; ²⁴Center for Cardiovascular Biology and Atherosclerosis Research, McGovern School of Medicine, University of Texas Health Science Center at Houston, Houston, TX, USA; ²⁵Department of Cardiology, Cardiovascular Research Institute Maastricht (CARIM), Maastricht University Medical Centre, Maastricht University, Maastricht, The Netherlands; ²⁶Department of Cardiovascular Sciences, University of Leuven, Leuven, Belgium; and ²⁷Institute for Molecular and Translational Therapeutic Strategies (IMTTS), Hannover Medical School, Hannover, Germany

Received 3 April 2020; revised 17 June 2020; editorial decision 13 July 2020; accepted 13 July 2020; online publish-ahead-of-print 19 July 2020

Abstract

In western countries, cardiovascular (CV) disease and cancer are the leading causes of death in the ageing population. Recent epidemiological data suggest that cancer is more frequent in patients with prevalent or incident CV disease, in particular, heart failure (HF). Indeed, there is a tight link in terms of shared risk factors and mechanisms between HF and cancer. HF induced by anticancer therapies has been extensively studied, primarily focusing on the toxic effects that anti-tumour treatments exert on cardiomyocytes. In this Cardio-Oncology update, members of the ESC Working Groups of Myocardial Function and Cellular Biology of the Heart discuss novel evidence

*Corresponding authors. Tel: +39 081 746 2242; fax: +39 081 746 2246, E-mail: carlogabriele.tocchetti@unina.it (C.G.T.); Tel: +49 511 532 9174; fax: +49 511 532 5274, E-mail: thum.thomas@mh-hannover.de (T.T.)

†The first two authors contributed equally to the study.

Published on behalf of the European Society of Cardiology. All rights reserved. © The Author(s) 2020. For permissions, please email: journals.permissions@oup.com.

interconnecting cardiac dysfunction and cancer via pathways in which cardiomyocytes may be involved but are not central. In particular, the multiple roles of cardiac stromal cells (endothelial cells and fibroblasts) and inflammatory cells are highlighted. Also, the gut microbiota is depicted as a new player at the crossroads between HF and cancer. Finally, the role of non-coding RNAs in Cardio-Oncology is also addressed. All these insights are expected to fuel additional research efforts in the field of Cardio-Oncology.

Keywords

Cardio-Oncology • Common pathways in heart failure and cancer • Multicellular and multiorgan mechanisms

1. Introduction

In the industrialized world, cardiovascular (CV) disease and cancer are the leading causes of death in the ageing population.¹ Left ventricular dysfunction (LVD) and heart failure (HF) are not rare across the broad population of cancer patients. In cancer patients, CV disease (CVD) is the most frequent non-cancer cause of death.² HF and cancer share the same risk factors (e.g. ageing, smoking, obesity, diabetes, dyslipidaemia, alcohol intake, inflammation).^{3,4} Furthermore, cancer and HF may have ancillary factors linking the two together.⁵ Registries have observed that HF patients have a higher cumulative incidence of cancer, with a worse prognosis when both co-exist,⁶ suggesting that cancer surveillance may be useful in the management of HF patients.⁷ Finally, an increased cumulative incidence of cancer among HF patients 30 days after myocardial infarction (MI) has been reported, compared to HF-free patients 30 days after MI.⁸

When considering these observations, it should be taken into account that there may be a surveillance bias, due to the fact that these study patients usually undergo an intense follow-up programme that may lead to anticipate cancer diagnosis, sometimes discovering malignancies that would have gone undiscovered. Moreover, some of the most common therapies used to treat HF patients may play a role in revealing tumours otherwise asymptomatic (e.g. a latent intestinal neoplasm can bleed due to anti-thrombotic therapy).³ Clinical presentations can also be difficult to distinguish between HF and cancer, since the two conditions can share some common symptoms (fatigue, dyspnoea, weight loss, muscle wasting, and oedema).^{1,3} This may delay the diagnosis of new-onset cancer in HF patients due to the overlap in clinical manifestation. Furthermore, CV function and predictors of exercise capacity are impaired in patients with cancer *per se*.⁹ Hence, symptoms due to a tumour may overlap with those of HF and be attributed to heart disease. This may even delay cancer diagnosis, as symptoms might be thought of as due to advancing disease rather than new cancer.³ Although the relationship between cancer and HF is not well-defined in clinical studies, there is increasing data to suggest mechanistic links between the two conditions that we discuss in our manuscript.

Besides these reciprocal relations, cancer and HF carry an independent risk of mortality and also limit optimal treatment of the other condition when they co-exist, contributing to higher mortality. In addition, the cardiotoxicity risk related to treatment with anticancer drugs may unmask or deteriorate pre-existing HF.³ The mechanisms driving HF triggered by anticancer therapies have been extensively investigated over the last 20 years and important insights have been uncovered.^{10–12} Nonetheless, major questions are still open, and the answers to these questions may lay the foundations for new strategies to detect, monitor, and treat cancer-therapy induced cardiotoxicity. On the other hand, research into the common pathways linking cancer and HF regardless of anticancer drugs has just begun.¹³

The latest insights in translational Cardio-Oncology were discussed during the joint meeting of the Working Groups of Myocardial Function and the WG of Cellular Biology of the Heart of the European Society of Cardiology, held in Naples, Italy, in May 2019. In particular, given the systemic involvement of both HF and cancer, the Cardio-Oncology session focused on the contribution of organs, systems, and cells other than cardiomyocytes to the pathogenesis of cardiac dysfunction in cancer patients, and to the interconnection between cancer and HF, primarily via inflammation. Opportunities and the current limitations in the use of microRNAs (miRNA) in cardio-oncology were also discussed. These topics are reviewed here, to provide the reader with updated information and further stimulate research in the field.

2. Role of non-cardiomyocytes in cancer treatment-related cardiotoxicity

The heart is a multicellular organ composed by cardiomyocytes, fibroblasts, neurons, endothelial, and haematopoietic-derived cells. In fact, cardiomyocytes are not the most abundant cell type.¹⁴ The different cardiac cell populations have diverse functions and also interact through complex intercellular communications.¹⁵ Most studies performed so far have focused on the effects of anticancer drugs on cardiomyocytes, in both *in vitro* systems and *in vivo* models¹⁶ (see Table 1). Briefly, among the many forms of cardiotoxicity caused by several anticancer drugs (Table 2), cardiac dysfunction due to anthracyclines, such as doxorubicin (DOXO) has historically been the most relevant.¹⁷ From a pathophysiological point of view, anthracyclines induce cardiomyocyte death, mainly apoptosis and necrosis, via different molecular mechanisms, including but not limited to induction of oxidative stress, activation of DNA damage responses, and impairment of mitochondrial biogenesis and metabolism.^{18–20} Among other mechanisms involved in anthracycline-induced cardiotoxicity, abnormalities in myocardial energetics have also been studied.^{21,22} Also biological drugs, designed to target specific oncologic pathways may be cardiotoxic, since these pathways play a major role in the maintenance of cardiac homeostasis, especially during stressful conditions, such as hypertension or hypertrophy.²³ For instance, *human epidermal growth factor receptor 2 (HER/Erbb2)* and *angiogenesis inhibitors* profoundly affect cardiomyocytes metabolism and contractile proteins, as discussed in comprehensive reviews.^{16,24–26}

In addition, anti-tumour therapies likely also affect non-cardiomyocytes in the heart. For instance, DOXO has been shown to exert toxic effects on cultured cardiac endothelial cells²⁷ and fibroblasts.^{28–30} This direct activity on non-cardiomyocytes may partly account for the cardiotoxicity of the drug, e.g. endothelial cells lose their barrier function with increased permeability and myocardial injury.

Table 1 Main direct toxic effects of cancer therapies on cardiomyocytes

Cellular toxicity	Treatment(s) most commonly involved
Type II topoisomerase poisoning ^a	Anthracyclines
Mitochondrial dysfunction	Anthracyclines, VEGFR/multitargeted RTK inhibitors
Oxidative stress	Anthracyclines
Impaired autophagy	Anthracyclines, proteasome inhibitors
Altered protein handling	Proteasome inhibitors
Induction of HIF pathways	VEGFR/multitargeted RTK inhibitors

^aThis toxicity is peculiar of anthracyclines.
HIF, hypoxia-inducible factor; RTK, receptor tyrosine kinase; VEGFR, vascular endothelial growth factor receptor.

Table 2 Cardiovascular toxicities of cancer therapies

Type of toxicity	Treatment(s) most commonly involved
LVD, HF	Anthracyclines, HER2-targeting drugs, VEGFR/multitargeted RTK inhibitors, proteasome inhibitors, radiation therapy (HFpEF)
Myocardial ischaemia	Fluoropyrimidines, VEGFR inhibitors, radiation therapy
Myocarditis	ICIs, cyclophosphamide (rarely)
Atrial fibrillation	Ibrutinib
QT prolongation	Arsenic trioxide, vandetanib, androgen deprivation therapy (enzalutamide)
Valvular heart disease	Radiation therapy
Pericarditis	ICIs, cyclophosphamide
Hypertension	VEGFR inhibitors
Peripheral artery disease	Nilotinib, ponatinib
Vascular thrombosis ^a	Cisplatin, nilotinib, ponatinib, thalidomide and lenalidomide, VEGFR inhibitors, proteasome inhibitors, aromatase inhibitors
Pulmonary arterial hypertension	Dasatinib, cyclophosphamide

HF, heart failure; HFpEF, heart failure with preserved ejection fraction; ICIs, immune checkpoint inhibitors; LVD, left ventricular dysfunction; RTK, receptor tyrosine kinase; VEGFR, vascular endothelial growth factor receptor.
^aAcute myocardial ischaemia will ensue if thrombosis occurs at coronary artery atherosclerotic plaques.

The impact of the toxicity of DOXO and any other anti-tumour treatment on non-cardiomyocytes can be better understood when it is placed into the context of the intercellular cross-talks in the heart. This concept is exemplified by the current knowledge about the cardiotoxicity of anti-HER2 drugs.³¹ Besides being expressed in breast cancer cells, HER2/ErbB2 is also physiologically present in cardiomyocytes together with another receptor tyrosine kinases (RTK) of the same family, HER4/ErbB4.³² Upon binding of HER4/ErbB4 by neuregulin-1 (NRG) and other ligands secreted by cardiac microvascular endothelial cells, HER2/ErbB2, and HER4/ErbB4 form heterodimers and initiate protective signalling cascades. Therefore, drugs targeting HER/ErbB2 are postulated to disrupt the NRG1-HER2/ErbB2-mediated endothelial cell-cardiomyocyte crosstalk and make cardiomyocytes more vulnerable to other stressors (Figure 1). It is notable that trastuzumab, used in the treatment of human epidermal growth factor receptor (HER)-2+ breast cancer, also directly damages cardiomyocytes and endothelial cells.^{33,34}

The inhibitors of the RTK for vascular endothelial growth factor (VEGF) and platelet-derived growth factor (PDGF) cause cardiac microvascular dysfunction secondary to depletion of coronary microvascular pericytes.³⁵ The resulting myocardial hypoxia leads to sustained expression of hypoxia-inducible factor alpha (HIF-α), which was demonstrated to be sufficient to cause cardiomyopathy.^{36,37} Indeed, enhanced vascular permeability and reversible microvascular vasoconstriction have been

reported in patients receiving therapies targeting VEGF and PDGF receptor (VEGFR and PDGFR, respectively).³⁸ Moreover, this mechanism of toxicity well explains the clinical observation that cardiomyopathy associated with anti-VEGFR/PDGFR agents is reversible.³⁹

However, evidence obtained over the last years suggests that blockade of VEGF signalling also interrupt endothelial cell-cardiomyocyte communication (Figure 1). VEGF binds VEGFR on endothelial cells to stimulate angiogenesis and also to induce the release of angiocrines (including ErbB4 and ErbB1 ligands) that modulate the function and homeostasis of adjacent cardiomyocytes.⁴⁰ Thus, drugs that inhibit VEGFR may alter cardiac function by interfering with the VEGF-VEGFR signalling axis, and by promoting endothelial cell dysfunction and death.^{41,42} High-throughput screening of RTK inhibitors pinpointed those targeting VEGFR2 and PDGFR as the most toxic in human-induced pluripotent stem cell (hiPSC)-derived endothelial cells.⁴³

Experimental models and analyses of human biopsies indicate that some features of HF with preserved ejection fraction (HFpEF) are at least in part driven by cardiac endothelial cell dysfunction. This latter elicits inflammatory infiltration of the myocardium, fibroblasts activation to deposit collagen excessively and increased stiffness triggered by a reduction of nitric oxide-dependent signalling.^{44–46} Consistently with the epidemiological finding that the risk of HFpEF is correlated with prior radiotherapy for breast cancer,⁴⁷ similar features were demonstrated in

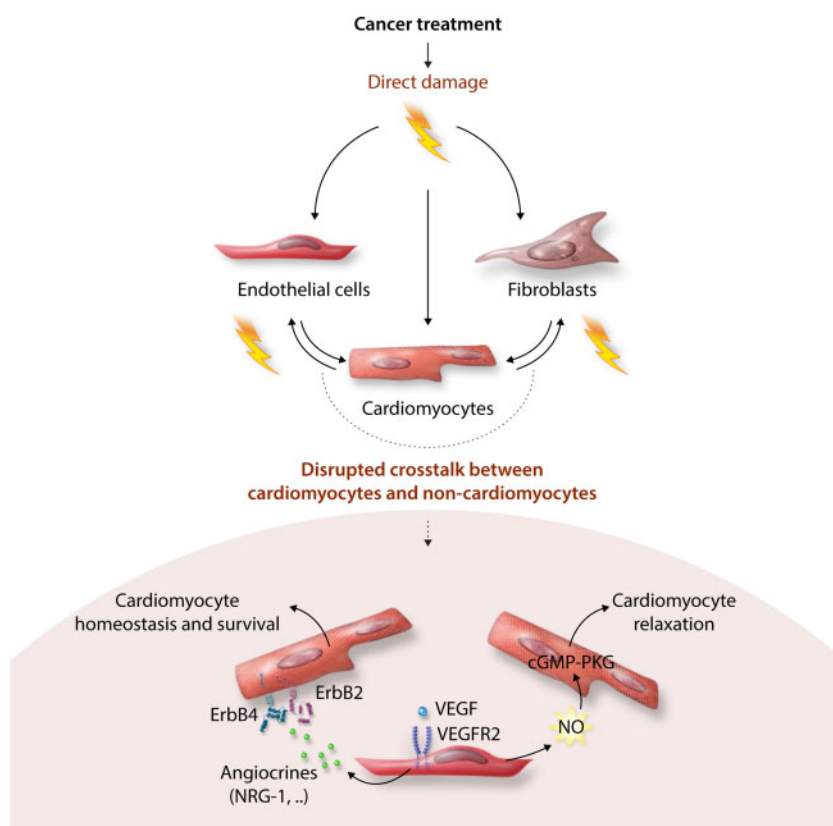


Figure 1 Besides directly affecting cardiomyocytes and the other cardiac cell populations, cancer treatments may disrupt the intercellular communications between cardiomyocytes and non-cardiomyocytes. The inset in the lower part of the figure shows key endothelial cell-cardiomyocyte paracrine signalling axes that may be impaired by anti-tumour therapies. cGMP, cyclic guanosine monophosphate; NO, nitric oxide; NRG-1, neuregulin-1; PKG, protein kinase G; VEGF, vascular endothelial growth factor; VEGFR2, vascular endothelial growth factor receptor type 2.

rats receiving cardiac radiation.⁴⁸ Since non-proliferating cardiomyocytes are considered resistant to ionizing radiation, other cell types, and in particular, endothelial cells, are predicted to be the main target of radiation therapy leading leads to HF.⁴⁹

Fibroblasts also regulate cardiomyocytes and inflammatory cells through their secretome.^{50,51} In a recent study, DOXO caused both apoptosis of cardiac fibroblasts and secretion of Fas ligand, which in turn promoted cardiomyocyte death in a paracrine manner.⁵² Conditional deletion of ataxia telangiectasia mutated kinase (ATM) in cardiac fibroblasts attenuated cardiac cell apoptosis, LVD, and mortality in response to DOXO, suggesting that fibroblast are central in the pathogenesis of DOXO cardiotoxicity through ATM. The interactions between fibroblasts and other cardiac cell types, and the mechanisms in the cardiotoxicity of anticancer therapies, are an important area for future research.⁵² Senescence of fibroblasts and possibly other cardiac stromal cells is especially worth being investigated since it has been proposed that it plays a major role in the pathogenesis of heart disease.⁵³

In conclusion, oncological drugs and radiotherapy induce abnormalities in non-cardiomyocytes, which secondarily derange the networks with cardiomyocytes and may lead to LVD and HF. Additional studies are needed,^{54,55} also considering that cardiotoxicity may be evident in an already damaged myocardium, but may remain latent or hidden in the healthy heart.⁵⁶ Since cardiac diseases and their comorbidities significantly change the global cardiac transcriptome, proteome, and

metabolome, it is not surprising that several drugs may act differently on the diseased vs. healthy hearts.^{57,58} Novel cardiac safety testing platforms involving combined experimental models of cardiac diseases in the presence and absence of major CV comorbidities and/or co-treatments are needed.⁵⁹ In this regard, cardiac organoids may allow modelling the complexity of the interactions between the different cardiac cell populations and, thereby, comprehensively evaluate the effects of anticancer therapies.⁵⁵

3. Interconnections between cancer and HF

Recently, attention has been drawn to the fact that cancer and heart disease have a reciprocal relationship: while the presence of cancer may cause LVD, the presence of HF associates with excess incident cancer.^{3-5,60} The communication between these two threatening diseases is complex, intriguing, and involves many components.

First, during life and ageing, several risk factors accumulate, which lead to chronic inflammation, oxidative stress, and protein and DNA instability. Classical CV risk factors, including obesity, diabetes, dyslipidaemia, and inflammation, are also associated with the development of cancer. Many of these risk factors lead to accumulation of fat mass, which is an active endocrine organ, secreting inflammatory factors, and adipokines,

which in turn have been associated with new-onset CVD and new-onset cancer.^{3–5,60}

Second, genetic mutations that accumulate throughout life, such as clonal haematopoiesis of indeterminate potential (CHIP), defined as the presence of clonal leucocytes with impaired immune proprieties derived by acquired mutation in haematopoietic stem cells, have been associated with both cancer and CVD, including HF.^{61–65} These mutations usually occur in a few genes, including DNMT3A, TET2, ASXL1, PPM1D, JAK2, TP53, SF3B1, and SRSF2.⁶⁶ The risk of developing CHIP increases with ageing and, although it rarely results in development of haematologic malignancies, it seems to be tightly linked to increased CV events and worse HF prognosis.^{61–65,67}

Also, genetic mutations in sarcomeric proteins predispose to HF in patients undergoing chemotherapy. Unrecognized rare variants in cardiomyopathy-associated genes, particularly *Titin* truncating variants, have been shown to increase the risk for systolic dysfunction and cardiac events in a relatively small population of both children and adults undergoing chemotherapy. In specific populations, genotype variant testing, along with cumulative chemotherapy dosage and traditional CV risk factors, may be useful to improve the identification of cancer patients with a higher risk for developing HF upon chemotherapy.⁶⁸

Other CV risk factors, such as hypertension and trace albuminuria, have been related to cancer development. Therefore, systemic risk factors likely exert effects on several damage pathways, and it is hypothesized that individual additional risk factors, such as genetic predisposition or pre-existing conditions, will also contribute to the risk of one or both conditions.

Third, cancer and CVD are both associated with profound changes in tissue structure, either growth of entirely new tissue or tissue deformation, remodelling, and scarring of pre-existing tissues, such as heart, endothelial cells, and matrix. Neoplasms are characterized by stroma, which is matrix tissue supporting the tumour, providing a scaffold, structure, and connections to adjacent organs. Further, most cancers, and especially metastases, rely on strong neovascularization requiring mitogenic endothelial cells and pericytes, where multiple growth factors play a role. In comparison, damaged cardiac tissue leads to dysfunctional cardiomyocytes, and also may develop extracellular matrix remodelling, fibrosis, and scar. Matrix is produced by activated fibroblasts and multiple cell types homing in, including monocytes, macrophages, and neutrophils. The cardiac scar is not a static structure, but rather is a dynamic and secreting structure.⁶⁹

4. Psychological convergence of HF and cancer

There is a well-established psychological impact on patients suffering from chronic conditions, notably HF. This is one of the main aims of cardiac rehabilitation programmes in these patients. Unfortunately, rehabilitation programmes have only recently been implemented in cancer patients, in a generic ‘one fits all’ umbrella rather than bespoke guidelines for specific cancers. It is notable, however, the recognition that both cardiac dysfunction syndromes and cancers have a significant impact in regards to neuronal changes. Whilst these have only just been thought of, the molecular and cellular mechanisms of neuro-biology change remain relatively unknown. It is likely that both neuronal changes *per se* and modifications in signalling and transmission underlie the clinical states of depression or cognitive changes in these patients. The most likely culprit remains the chronic systemic inflammatory state present in both,

probably responsible for an enhanced level of oxidative stress, DNA damage, mitochondrial dysfunction, and synaptic modifications.^{70,71}

Whilst there is available evidence to support a link between certain chemotherapies and peripheral neuropathy (for example cisplatin), the issue of clinical states of depression/cognitive changes and them *per se* being a basis for autonomic dysfunction seen in these patients is far more complex and yet undemonstrated. At this current time, it does not have the level of evidence and merits further exploration.

5. Inflammation at the crossroad between cancer, cardiotoxicity of anticancer therapies and HF

Abnormal inflammation is increasingly recognized as a common driver of CVD and cancer.^{72,73} HF is characterized by a state of mild chronic systemic inflammation, with increased circulating concentrations of pro-inflammatory cytokines, such as tumour necrosis factor- α (TNF- α), interleukin-1 (IL-1), and interleukin-6 (IL-6). Myocardial injury itself triggers the recruitment and the activation of immune cells, which in turn produce pro-inflammatory cytokines and contribute to a self-perpetuating inflammatory state that underlies adverse tissue remodelling, primarily associated with capillary dysfunction and fibrosis.⁷⁴ Doxorubicin-induced damage also involves inflammation (Figure 2), with upregulation of pro-inflammatory toll-like receptor 4 (TLR4) in macrophages,⁷⁵ higher levels as TNF- α and IL-6 and reduced levels of the anti-inflammatory cytokine IL-10.⁷⁶ Cardiac function was preserved and survival improved in TLR2 knock-out mice after DOXO exposure compared to wild-types.⁷⁷ DOXO also induces local modulators of inflammation and fibrosis, produced by both macrophages and fibroblasts. Increased production of the matricellular protein thrombospondin-2 (TSP2) is protective in mice treated with DOXO. Enhanced myocyte damage in the absence of TSP-2 was associated with impaired activation of the Akt signalling pathway. Inhibition of Akt phosphorylation in cardiomyocytes significantly reduced TSP-2 expression, unveiling a unique feedback loop between Akt and TSP-2.⁷⁸ Importantly, CCL2/CCR2-dependent recruitment of functional antigen-presenting cells into tumours is a desired therapeutic effect of anthracyclines.⁷⁹

Indeed, for decades oncologists have been developing strategies to modulate inflammation in order to achieve therapeutic anticancer immune responses.⁸⁰ The first attempts were not really successful, since cancer escapes T-cell-mediated cancer-specific immunity via inhibitory pathways mediated by cytotoxic T-lymphocyte-associated protein 4 (CTLA-4), programmed cell death protein 1 (PD-1), and programmed cell death ligand 1 (PD-L1) (all depressing the antineoplastic activity of T lymphocytes).⁸¹ On the opposite, in the last years, immune checkpoint inhibitors (ICIs), such as monoclonal antibodies (mAbs) targeting CTLA-4, PD-1, and PD-L1, have dramatically improved the outcome of many malignancies, but serious immune-related CV adverse events have been observed^{82–84} (Figure 2). Interfering with the CTLA-4 and PD-1 axes can bring to autoimmune myocarditis and dilated cardiomyopathy,⁸⁵ suggesting that these molecules play an important role in preventing autoimmunity.⁸⁶ Hence, immunosuppressive therapies may be necessary to halt immune-related adverse events (IRAEs) and major adverse CV events (MACE).^{87–89}

More recently, engineered T cells with chimeric antigen receptors (CAR-T cells) have been approved by the U.S. Food and Drug Administration (FDA) as the first genetically modified autologous T-cell

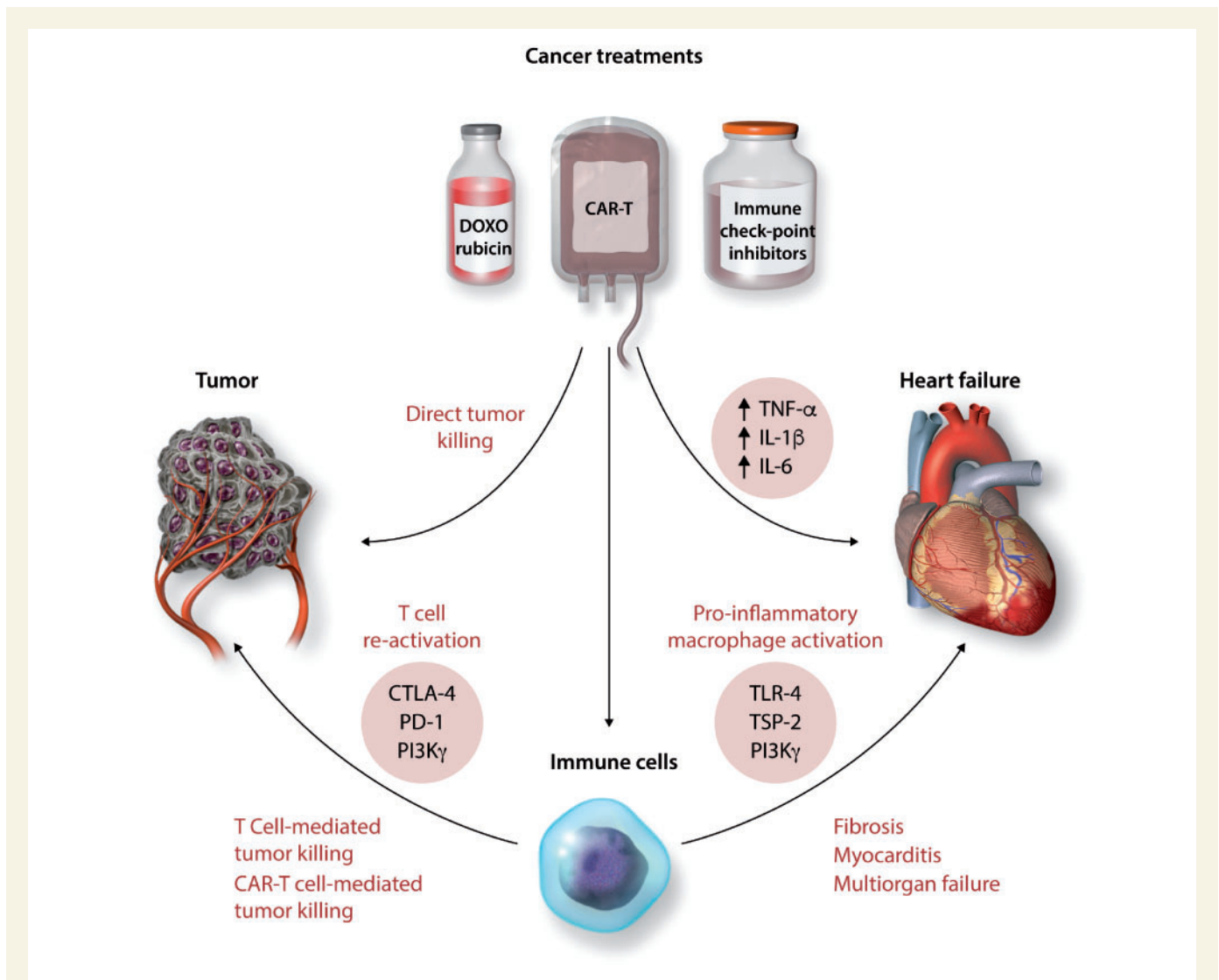


Figure 2 Inflammation at the intersection of the anticancer action and cardiac side effects of major oncological treatments. Besides directly killing tumour cells, doxorubicin triggers cardiac inflammation via activation of macrophages and fibroblasts and the ensuing release of local modulators of inflammation and fibrosis, such as TNF- α , IL-1 β , and IL-6. Major players of the inflammatory response induced by doxorubicin include macrophage TLR-4, the matricellular protein thrombospondin-2 (TSP-2), and leucocyte PI3K γ . On the other hand, immune check point inhibitors (ICIs) inhibit molecules, such as cytotoxic-T-lymphocyte-associated antigen 4 (CTLA-4), programmed cell death 1 (PD-1), and its ligand PD-L1. As a consequence, anti-tumour immune cell responses are reactivated and lead to tumour cell death, but concomitantly drives myocarditis. Although these new immunotherapies have notable anticancer effects, multiple mechanisms of immune resistance exist, and these might be overcome by using PI3K γ inhibitors that re-shape the tumour immune microenvironment. Finally, engineered T cells with chimeric antigen receptors (CAR-T cells) boosts T-cell-mediated tumour killing but are burdened by cytokine release syndrome (CRS) leading to extremely serious complications, including cardiac and vascular dysfunction, and ultimately to multiorgan failure.

immunotherapeutic agents that target CD-19. CD-19 is broadly expressed on most B-cell malignancies and has limited expression beyond B-cell lineage.^{90,91} Unfortunately, CAR-T cells are burdened by cytokine release syndrome (CRS) that is due to elevated levels of inflammatory cytokines released by activated CAR-T cells and other immune cells, such as macrophages, with fever and tachycardia that may be associated with hypotension and hypoxia. Also, cardiac dysfunction and extremely serious complications, such as vascular leak syndrome with circulatory collapse and multiorgan failure can be dreadful side effects of these therapies^{92,93} (Figure 2). Beside CAR-T cells, bispecific antibodies such as blinatumumab [that targets CD19 and CD3 and is increasingly used in the treatment of

Philadelphia chromosome negative B-cell acute lymphoblastic leukaemia (ALL)] can also lead to CRS and cardiomyopathy.⁹⁴

Interestingly, inflammation in cancer plays a dual role. On the one hand, it is essential to recognize and destroy cancer cells; on the other hand, it provides a fertile milieu for tumourigenesis and plays key roles in different steps of tumour development, from initiation and promotion to invasion and metastasis. Tumour-associated inflammation favours proliferation and survival of malignant cells, promotes angiogenesis and metastasis, undermines adaptive immune responses, and potentially interferes with responses to hormones and chemotherapeutic agents.^{95,96} The finding that anti-inflammatory agents are effective in the prevention of

cancer and CVD further advocates inflammation as a common contributor to both diseases. A 2019 study concluded that chronic systemic low-grade inflammation, measured by C reactive protein (CRP) levels <10 mg/L, is a risk factor for incident cancer, in particular, lung cancer, in patients with stable CVD. The relation between inflammation and incident cancer is seen in former and current smokers and is uncertain in never smokers.⁹⁷ Blockade of the pro-inflammatory cytokine IL-1 β with canakinumab was shown to significantly reduce the rate of recurrent CV events in patients with previous MI (CANTOS trial). At the same time, blocking IL-1 β appeared to protect from lung cancer mortality.^{98,99} Mice exposed to DOXO showed an increase in serum IL-1 β along with other inflammatory factors.¹⁰⁰ Moreover, the IL-1 β receptor antagonism protects against DOXO cardiotoxicity.¹⁰¹ Similarly, the IL-6 inhibitor tocilizumab can protect against MACE in CAR-T patients.⁹³

The experience of IL-1 β blockade highlights that the identification of key players of the inflammatory response is important to tackle both cancer and heart disease. Among intriguing candidates are PI3Ks, and more specifically the PI3K γ isoform that is enriched in both cardiomyocytes and leucocytes (Figure 2). This implies a key role for this isoform not only in the control of cardiomyocyte pathobiology but also in the orchestration of the inflammatory response associated with different types of CV injury.¹⁰² PI3K γ is up-regulated in patients and mouse models of atherosclerosis, and directs leucocyte infiltration of the arterial wall, which is a key pathogenic event in atherosclerosis.¹⁰³ PI3K γ -mediated inflammation is also pivotal to the cardiac response to pressure overload.¹⁰⁴

Besides directing the cardiac response to stress, macrophage PI3K γ expression critically contributes to tumour growth and progression. Intriguingly, macrophages play opposite roles in non-oncological inflammatory conditions and cancer. In response to pathogens or injury, macrophages express cytokines that stimulate cytotoxic T cells to clear infected or damaged cells. Conversely, in cancer macrophages express anti-inflammatory cytokines that induce immune suppression, inhibit T-cell-mediated tumour killing and promote resistance to immunotherapies (i.e. T-cell checkpoint inhibitors). PI3K γ has been recently proposed as the molecular switch controlling immune stimulation and suppression in cancer.¹⁰⁵ The unique feature of macrophage PI3K γ , playing a maladaptive role both in heart disease and in cancer, makes this enzyme the ideal pharmacological target to 'kill two birds with one stone', i.e. to halt the tumour and at the same time treat the heart.¹⁰⁶ This is particularly relevant for cancer patients treated with chemotherapy and suffering from iatrogenic cardiotoxicity.^{15,105,107} Results from clinical trials assessing the combined anticancer effect of such compounds in a context of cardiac protection are awaited.

6. The gut microbiome in Cardio-Oncology

HF has long been recognized to be associated with altered gut function.^{108,109} Low cardiac output in HF results in intestinal ischaemia, with congestion of the splanchnic circulation, bowel wall oedema, and impaired intestinal barrier function (Figure 3). This condition increases the overall inflammatory state and oxidative stress as a consequence of HF-induced ischaemia and congestion within the gut via enhanced bacterial translocation and the presence of bacterial products in the blood circulation. Increased leakiness modifies the gut environment and affects its resident microbial population.¹¹⁰

Among the conditions that can influence the gut composition, including individual genetic variability, lifestyle, colonization, and delivery at

birth^{111–113} also changes in diet, presence of diseases and relative treatments have to be considered.¹¹⁴ Interestingly, genetic composition of gut microbiota, defined as microbiome, also influences cancer development and progression in different ways.¹¹⁵ Several types of cancers (head and neck, lung, colorectal, and cervical carcinomas) promote a shift in microbiome composition.^{116–118} In addition, chemotherapy directly impacts the gut microbiota and its efficacy is strongly influenced by microbiome composition (Figure 3).^{119,120}

Metabolites generated by the gut microbiota derive from the fermentation of indigestible fibres to short-chain fatty acids, which have protective properties (reducing inflammation, oxidative stress,^{121,122} and improving vascular tone). Dietary sources of choline, phosphatidylcholine, l-carnitine, and other methylamine-containing nutrients provide substrates for microbiota-mediated generation of trimethylamine (TMA) that accesses the portal circulation and is converted by the hepatic flavin-containing monooxygenase (FMO) family of enzymes into trimethylamine N-oxide (TMAO, Table 3). TMAO can favour the development of atherosclerosis, thrombosis, kidney disease, and HF (Figure 3). High plasma levels of TMAO have been suggested to be predictive of CV events of mortality, independently from renal function and CV comorbidities.¹¹⁰

Additionally, the bacterial transformation of bile acids can result in altered bile acid profiles, which in turn can impact systemic inflammatory and fibrotic processes.¹¹⁰ Importantly, microbiota-derived peptide mimics may also drive HF, by inducing a lethal inflammatory cardiomyopathy. Cardiac myosin-specific TH17 cells are being imprinted in the intestine by a commensal *Bacteroides* species peptide mimic. These cells promote cardiac inflammation and dysfunction in genetically susceptible individuals.¹²³

Several studies reported SCFAs-producing bacteria perturbation in patients with CVDs.¹²⁴ Among these SCFA generated by the gut microbiota, butyrate (BUT) has multiple beneficial effects for our CV system through different mechanisms^{125–131,120} (Table 3). BUT exerts major epigenetic effects, acting as a potent inhibitor of histone deacetylase (HDACs) activity. Inhibition of HDACs is well-known to protect the heart from pathologic hypertrophy and ischaemia.^{132–135} Among HDAC inhibitors, BUT has been shown to exert anti-neoplastic properties *in vitro*^{120,136}, while its derivatives can enhance the anticancer cytotoxic effects of DOXO while protecting against cardiotoxicity¹³⁷ and can decrease cardiac apoptosis and myocardial dysfunction induced by DOXO, by lowering endoplasmic reticulum stress-initiated apoptotic signalling and HDAC-inhibition mechanisms.^{138,139} The cardioprotective effect of BUT and analogues is associated with the production of anti-inflammatory molecules, cytoprotection, modulation of angiogenesis, limiting the occurrence of cardiotoxic manifestations caused by DOXO treatments, with reduction of nitrosative and oxidative stress, counteracting mitochondrial dysfunction.¹²¹ In turn, DOXO is reported to induce GUT-microbiota dysbiosis in mice, while the administration of BUT attenuates the inflammation state induced by DOXO,¹⁴⁰ fuelling nutraceutical as a new promising area of research to cardio-oncology.

7. Opportunities and limitations in the use of non-coding RNAs in Cardio-Oncology

Multiple evidence seems to suggest an involvement of circulating microRNAs (miRNAs) in anthracyclines-induced cardiotoxicity both *in vivo* and in the clinical setting, evidencing a very heterogeneous situation. In particular, when focusing on DOXO, miR-1^{141,142} and miR-

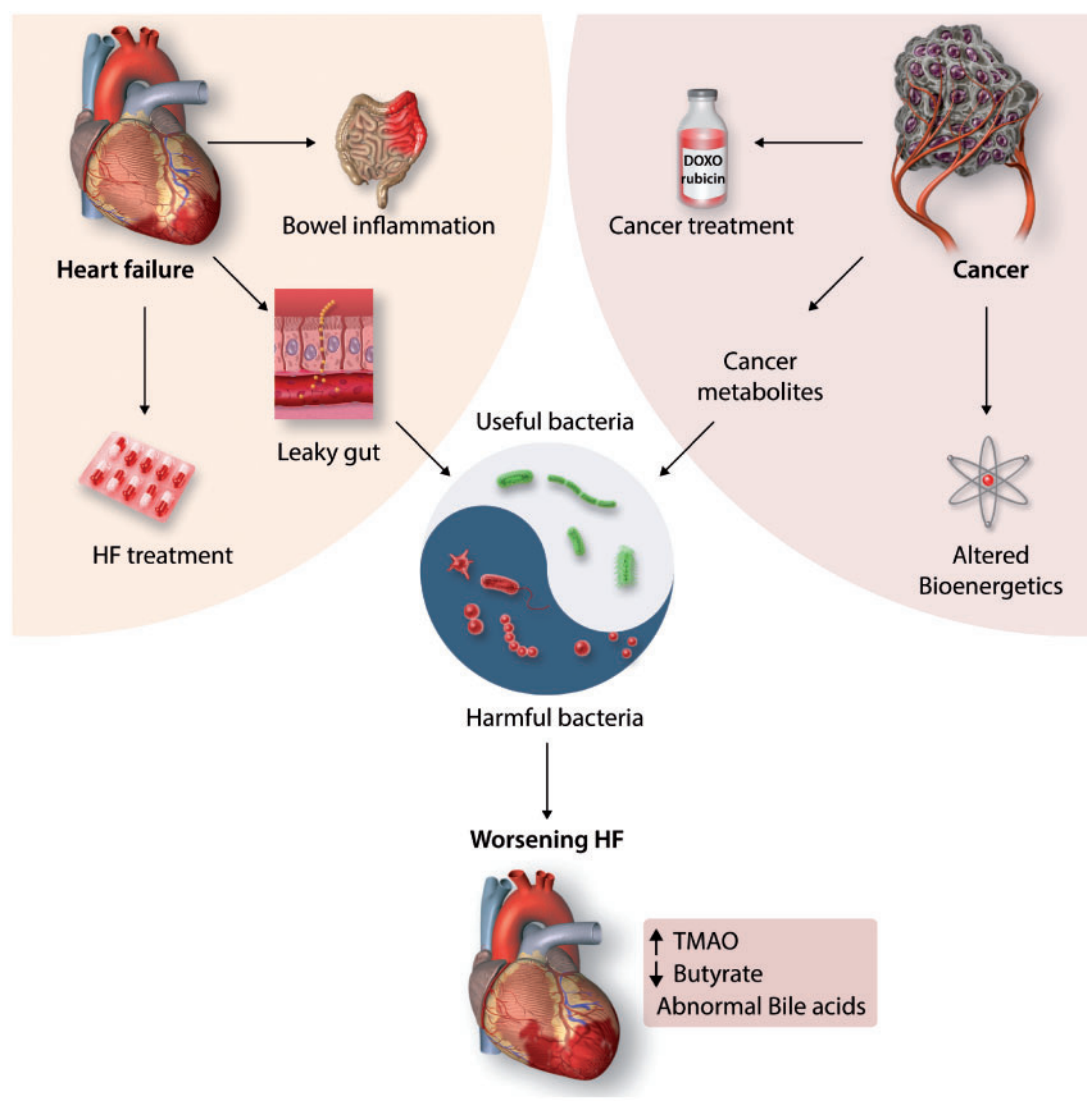


Figure 3 GUT microbiome dysbiosis can be influenced by both HF and cancer. HF has long been associated with congestion of splanchnic circulation, leading to bowel wall oedema, impaired intestinal barrier function and increased systemic inflammation, which drastically affect GUT microbiome composition and response to HF treatments. At the same time, cancer-mediated disruption of metabolism and the production of cancer-derived metabolites modifies the microbiome. Such altered gut microbiome generates cardiotoxic metabolites, such as TMAO and Bile Acids, eventually leading to HF worsening.

34a,^{143–146} showed a drug-induced regulation in tissues and plasma samples, both in patients and animal models. miR-1 is one of the most investigated and most highly expressed miRNAs in cardiac and skeletal muscle, both in physiological^{146,147} and pathologic^{148,149} condition. While many groups have indicated miR-1 as a specific circulating marker of heart disease, there is no clear indication about its unambiguous cardiac origin, particularly in anthracyclines-induced toxicity, which is a systemic phenomenon. Similarly, miR-34a was demonstrated to be modulated by anthracyclines both in experimental models^{144,145,150} and in breast cancer patients.¹⁵¹ Piegari *et al.*,¹⁴⁴ showed that tissue regulation of miR-34a by DOXO was not restricted only to the heart, hinting at a multi-tissue contribution to the circulating levels of this miRNA. Indeed, besides cardiomyocytes, smooth muscle cells, fibroblasts, cardiac progenitor cells, and endothelial cells may also play a role in DOXO-induced

cardiomyopathy.¹⁵² Acute DOXO treatment in mice was shown to reduce microvessel density and VEGF-A expression with a parallel increase in miR-320a.¹⁵³ Inhibition of miR320a improved cardiac function, decreased apoptosis, and increased microvessel density in DOXO-treated mice, while overexpression of miR-320a worsened DOXO-induced LV dysfunction.¹⁵⁴ Conversely, overexpression of the miR-320a target VEGF-A prevented detrimental effects of miR-320a in DOXO-cardiotoxicity experimental model confirming VEGF as a direct downstream target molecule.¹⁵³ Mechanistically, the overexpression of the pro-hypertrophic miR-212/132 cluster in primary rodent and human iPSC-derived cardiomyocytes and in *in vivo* models has been shown to inhibit doxorubicin-induced toxicity.¹⁵⁵ Also, another class of non-coding RNAs, circular RNAs, may play a crucial role in mediating cardiotoxicity of doxorubicin; indeed, overexpression of the RNA binding protein

Table 3 Role of TMAO and butyrate in cardio-oncology

Cardiovascular field		Cancer field	
TMAO	Butyrate	TMAO	Butyrate
• Prognostic 5-year follow-up marker in patients with heart failure • Prognostic biomarker in in chronic systolic HF • Predictive biomarker in patients with Acute Heart Failure • Prognostic biomarker in patients associated with NYHA III and IV ischaemic aetiology and adverse outcomes • Predictive biomarker for mortality and CV mortality in HFrEF but not HFpEF patients^{110,a}	• Inhibition of maladaptive hypertrophy and heart failure^{132,133,135} • Reduction of myocardial ischaemia–reperfusion injury¹³⁴ • Cardioprotective action against DOXO toxicity¹³⁷ • Alleviation of DOXO-induced ER stress^{138,139} • Prevention of DOXO-induced mitochondrial dysfunction and ROS/RNS production¹²¹	• Predictive biomarker of colorectal cancer¹⁵⁹ • Predictive biomarker of aggressive prostate cancer¹⁶⁰	• Induction of cytodifferentiation and inhibition leukaemic cells proliferation; inhibition of Lewis lung carcinoma cells growth¹³⁶ • Increase of the antineoplastic effect of DOXO¹³⁷

NYHA, New York Heart Association.
^a High levels of TMAO in patients have been suggested to be predictive of cardiovascular events of mortality, independently from renal function and cardiovascular comorbidities.¹¹⁰

Quaking 5 (*Qki5*) strongly attenuated the toxic effect of doxorubicin in a mouse model by regulating a set of circular RNAs including those derived from titin (*Ttn*¹⁵⁶).

The role of miRNAs as markers of cardiotoxicity has also been investigated. Ruggeri *et al.*¹⁵⁰ showed that after one month from DOXO administration, only a part of the drug-treated mice presented cardiac dysfunction, similarly to the clinical context. miR-1 was again among the circulating miRNAs regulated after cardiotoxicity onset, together with miR-499-5p. In an acute DOXO cardiotoxicity model, the same authors showed that miR-34a-5p and miR-451a were dysregulated in all cardiac chambers, with miR34a-5p showing opposite trends of regulation between the atria and the ventricles of treated mice. In another study using DOXO both *in vivo* and *in vitro* acutely and chronically treated cardiomyocytes, DOXO-dependent downregulation of miR-30 led to increased cardiomyocyte apoptosis and abnormalities of cardiomyocyte β-adrenergic receptor signalling.¹⁵⁷

Importantly, only part of circulating miRNAs overlapped with their cardiac counterparts, suggesting only a partial contribution of the heart to the variations in circulating levels of miRNAs upon drug administration. Limitations of the studies are the number of animals, the number of screened miRNAs (often only selected CV miRNAs), the acute phase observed, the absence of tumour in the experimental models, and the lack of additional cancer treatments.

Besides the few miRNAs showing a ‘reproducible sensitivity’ to anthracycline treatment, there is a highly heterogeneous picture composed by past and present investigations. While the discrepancies in terms of results could be in part explained by the different experimental models and by the different malignancies and therapies adopted in patients-based investigations, there are at least two fundamental issues that should be addressed in future works. A striking feature of many, if not all, published papers is that no study described a decline of left ventricular ejection fraction (LVEF) below the ‘normal’ threshold of 50%, possibly because of lack of a long-term follow-up. Moreover, the vast majority of human-based research studies concentrated on the acute

phase of cardiotoxicity,¹⁵⁸ and the same limitations often apply also to *experimental* researches, which rarely go beyond a few days’ time span from treatment to sacrifice.

Additional data on the main non-coding RNAs are summarized in Table 4.

8. Concluding remarks

We discussed several of the novel exciting insights that are emerging in the ever-expanding field of cardio-oncology. More research is required to identify and investigate the pathways and mechanisms underpinning the intimate relationship between CVD and cancer. Current studies focus on shared risk factors, both acquired/modifiable and genetic. The substantial structural changes in diseased organs prompt further studies in an effort to learn how disease in one organ may communicate with another organ. Learning from each disease mechanisms may help to combat both CVD disease and cancer.

Acknowledgements

We acknowledge the support from the Netherlands Cardiovascular Research Initiative, an initiative with support of the Dutch Heart Foundation, CVON2016-Early HFPEF, 2015–10, CVON She-PREDICTS, grant 2017–21, CVON Arena-PRIME, 2017–18. Furthermore, we acknowledge the support of FWO G091018N and FWO G0B5930N to S.H. I.F.P. was supported by Fundo Europeu de Desenvolvimento Regional (FEDER) through Compete 2020 – Programa Operacional Competitividade E Internacionalização (POCI), the project DOCNET (norte-01-0145-feder-000003) was supported by Norte Portugal regional operational programme (norte 2020), under the Portugal 2020 partnership agreement, through the European Regional Development Fund (ERDF).

Table 4 Non-coding RNAs in cardio-oncology

	Cardiovascular field		Cancer field	
	Circulating forms (biomarkers)	Intracellular forms (epigenetic activity)	Circulating forms (biomarkers)	Intracellular forms (epigenetic activity)
miR-1	Marker of myocardial infarction ¹⁶¹	Involved in cardiogenesis, heart function, cardiac pathology ¹⁶²	Non-invasive biomarkers in breast cancer: early diagnosis and metastasis prediction ¹⁶⁴	Inhibition of proliferation and metastasis of breast cancer ¹⁶⁶
	Biomarker of atherosclerosis, coronary artery disease, acute coronary syndrome ¹⁴⁸	Involved in heart disease and cardioprotection ¹⁶³	Correlation with clinico-pathologic characteristics and lung cancer detection ¹⁶⁵	Differential expression in different human cancers ¹⁶⁷
miR-34a	Biomarker of ageing ¹⁶⁸	Regulation of cardiac ageing and function ¹⁷⁰	Non-invasive biomarkers in breast cancer: early diagnosis and metastasis prediction ¹⁶⁴	Potential tumour suppressor and therapeutic candidate in cancer ¹⁷³
	Marker of anthracycline treatment ¹⁶⁹	Contribution to doxorubicin-induced cardiotoxicity ¹⁷¹	Potential biomarker for early diagnosis of oesophageal cancer ¹⁷²	Associated with aggressive breast cancer ¹⁷⁴
miR-320a	Biomarker of arrhythmogenic cardiomyopathy ¹⁷⁵	Mediator of doxorubicin-induced cardiotoxicity ¹⁵³	Early detection of pancreatic neoplasia ¹⁷⁸ circulating biomarker of melanoma ¹⁷⁹	Regulation of cell metastasis and invasion in non-small-cell lung cancer ¹⁸⁰
	Predictive biomarker for left ventricular remodelling ¹⁷⁶	Involved in human myocardial infarction ¹⁷⁷		Modulation of cell growth and chemosensitivity in gastric cancer ¹⁸¹
miR-212/132 cluster	Estimation of atherosclerosis presence ¹⁸²	Prevention of doxorubicin-mediated atrophy and cardiotoxicity ¹⁵⁵	Biomarker of pancreatic cancer risk ¹⁸⁵	Suppression of proliferation of human lung cancer cells ¹⁸⁷
	Risk prediction for heart failure ¹⁸³	Regulation of cardiac hypertrophy and cardiomyocyte autophagy ¹⁸⁴	Diagnostic biomarker of malignant mesothelioma ¹⁸⁶	Regulated in ovarian cancer cells ¹⁸⁸
miR-451a	Modulated in acute model of doxorubicin-induced cardiotoxicity ¹⁴³	Regulation of doxorubicin-induced cardiotoxicity ¹⁹⁰	Biomarker and potential therapeutic target for cancer ¹⁹²	Regulation of papillary thyroid carcinoma cells ¹⁹⁴
	Marker of type 2 diabetes ¹⁸⁹	Decreased in hypertrophic cardiomyopathy and regulates autophagy ¹⁹¹	Biomarker for the diagnosis of oesophageal squamous cell carcinoma and squamous dysplasia ¹⁹³	Involved in breast cancer ¹⁹⁵
miR-30	Biomarkers of acute coronary syndrome and stable coronary artery disease ¹⁹⁶	Cardioprotection in doxorubicin-induced heart failure and inhibition of cardiomyocytes autophagy ¹⁹⁸	Prognosis of ovarian cancer ²⁰⁰	Involved in cancer progression ²⁰²
	Marker of stable atherosclerotic disease ¹⁹⁷	Involvement in ventricular remodelling: the miR-30 family ¹⁹⁹	Prognosis of bladder cancer ²⁰¹	Involved in breast cancer invasion, osteomimicry, and bone destruction ²⁰³

Funding

C.G.T. was supported by a Ricerca di Ateneo/Federico II University grant, P.A. is supported by the Italian Ministry of Health (GR-2018-12365661, CHANGE Study), P.F. was supported by the National Research, Development and Innovation Office of Hungary (Research Excellence Program - TKP, National Heart Program NVKP 16-1-2016-0017; 2019-1.1.1-PIACI-KFI-2019-00367) and by the Higher Education Institutional Excellence Program of the Ministry of Human Capacities in Hungary, within the framework of the Therapeutic Development thematic programme of the Semmelweis University. T.T. received funding from the EU-ERANET grant EXPERT. L.B. is Research director of FRS-FNRS, Belgium, R.M. received funding from Incyte Biosciences. R.A.d.B. was supported by the European Research Council (ERC CoG 818715, SECRETE-HF) and the CVON grants from Netherlands Heart Foundation (DOSIS, 2014-40; SHE-PREDICTS-HF, 2017-21; RED-CVD, 2017-11; and PREDICT2, 2018-30). This manuscript has been possible thanks to the support of the ERA-Net-CVD project MacroERA, 01KL1706, and IMI2-CARDIATEAM (N° 821508) to S.H., Associazione Italiana per la Ricerca sul Cancro (AIRC) IG 2016 Id.19032 to SZ, Leducq 09CVD01 "Targeted Approaches for Prevention and Treatment of Anthracycline-Induced Cardiotoxicity" to E.H. and A.G.

Conflict of interest: P.F. is the founder and CEO of Pharmahungary Group, a group of R&D companies. T.T. has filed and licensed patents in the field of non-coding RNAs. T.T. is founder and shareholder of Cardior Pharmaceuticals GmbH. The UMCG, which employs RAdB has received research grants and/or fees from AstraZeneca, Abbott, Bristol-Myers Squibb, Novartis, Novo Nordisk, and Roche. T.T. received grants/fees from Novo Nordisk, Sanofi-Genzyme, Amicus Therapeutics. R.A.d.B. received speaker fees from Abbott, AstraZeneca, Novartis, and Roche.

References

- Anker MS, von Haehling S, Landmesser U, Coats AJS, Anker SD. Cancer and heart failure-more than meets the eye: common risk factors and co-morbidities. *Eur J Heart Fail* 2018;**20**:1382–1384.
- Strongman H, Gadd S, Matthews A, Mansfield KE, Stanway S, Lyon AR, Dos-Santos-Silva I, Smeeth L, Bhaskaran K. Medium and long-term risks of specific cardiovascular diseases in survivors of 20 adult cancers: a population-based cohort study using multiple linked UK electronic health records databases. *Lancet* 2019;**394**:1041–1054.
- Ameri P, Canepa M, Anker MS, Belenkov Y, Bergler-Klein J, Cohen-Solal A, Farmakis D, Lopez-Fernandez T, Lainscak M, Pudil R, Ruschitska F, Seferovic P, Filippatos G, Coats A, Suter T, Von Haehling S, Ciardiello F, de Boer RA, Lyon AR, Tocchetti CG; for the Heart Failure Association Cardio-Oncology Study Group of the European Society of Cardiology. Cancer diagnosis in patients with heart failure: epidemiology, clinical implications and gaps in knowledge. *Eur J Heart Fail* 2018;**20**: 879–887.
- Meijers WC, de Boer RA. Common risk factors for heart failure and cancer. *Cardiovasc Res* 2019;**115**:844–853.
- Meijers WC, Maglione M, Bakker SJL, Oberhuber R, Kieneker LM, de Jong S, Haubner BJ, Nagengast WB, Lyon AR, van der Vegt B, van Veldhuisen DJ, Westenbrink BD, van der Meer P, Silljé HHV, de Boer RA. Heart failure stimulates tumor growth by circulating factors. *Circulation* 2018;**138**:678–691.
- Banke A, Schou M, Videbaek L, Moller JE, Torp-Pedersen C, Gustafsson F, Dahl JS, Kober L, Hildebrandt PR, Gislason GH. Incidence of cancer in patients with chronic heart failure: a long-term follow-up study. *Eur J Heart Fail* 2016;**18**:260–266.
- Hasin T, Gerber Y, McNallan SM, Weston SA, Kushwaha SS, Nelson TJ, Cerhan JR, Roger VL. Patients with heart failure have an increased risk of incident cancer. *J Am Coll Cardiol* 2013;**62**:881–886.
- Hasin T, Gerber Y, Weston SA, Jiang R, Killian JM, Manemann SM, Cerhan JR, Roger VL. Heart failure after myocardial infarction is associated with increased risk of cancer. *J Am Coll Cardiol* 2016;**68**:265–271.
- Cramer L, Hildebrandt B, Kung T, Wichmann K, Springer J, Doeberner W, Sandek A, Valentova M, Stojakovic T, Scharnagl H, Riess H, Anker SD, von Haehling S. Cardiovascular function and predictors of exercise capacity in patients with colorectal cancer. *J Am Coll Cardiol* 2014;**64**:1310–1319.
- Antoniades C, Small HY, Guzik T. The evolution of Cardiovascular Research Online: online and on demand. *Cardiovasc Res* 2018;**114**:e9.
- Moslehi J, Fujiwara K, Guzik T. Cardio-oncology: a novel platform for basic and translational cardiovascular investigation driven by clinical need. *Cardiovasc Res* 2019; **115**:819–823.
- Molinaro M, Ameri P, Marone G, Petretta M, Abete P, Di Lisa F, De Placido S, Bonaduce D, Tocchetti CG. Recent advances on pathophysiology, diagnostic and therapeutic insights in cardiac dysfunction induced by antineoplastic drugs. *Biomed Res Int* 2015;**2015**:1–14.
- Bertero E, Ameri P, Maack C. Bidirectional relationship between cancer and heart failure: old and new issues in cardio-oncology. *Card Fail Rev* 2019;**5**:106–111.
- Pinto AR, Ilinykh A, Ivey MJ, Kuwabara JT, D'Antoni ML, Debuque R, Chandran A, Wang L, Arora K, Rosenthal NA, Tallquist MD. Revisiting cardiac cellular composition. *Circ Res* 2016;**118**:400–409.
- Skelly DA, Squiers GT, McLellan MA, Bolisetty MT, Robson P, Rosenthal NA, Pinto AR. Single-cell transcriptional profiling reveals cellular diversity and intercommunication in the mouse heart. *Cell Rep* 2018;**22**:600–610.
- Ky B, Vejpongsa P, Yeh ET, Force T, Moslehi JJ. Emerging paradigms in cardiomyopathies associated with cancer therapies. *Circ Res* 2013;**113**:754–764.
- Ewer MS, Von Hoff DD, Benjamin RS. A historical perspective of anthracycline cardiotoxicity. *Heart Fail Clin* 2011;**7**:363–372.
- Varricchi G, Ameri P, Cadeddu C, Ghigo A, Madonna R, Marone G, Mercurio V, Monte I, Novo G, Parrella P, Pirozzi F, Pecoraro A, Spallarossa P, Zito C, Mercurio G, Pagliaro P, Tocchetti CG. Antineoplastic drug-induced cardiotoxicity: a redox perspective. *Front Physiol* 2018;**9**:167.
- Zhang S, Liu X, Bawa-Khalfe T, Lu LS, Lyu YL, Liu LF, Yeh ET. Identification of the molecular basis of doxorubicin-induced cardiotoxicity. *Nat Med* 2012;**18**: 1639–1642.
- Amgalan D, Garner TP, Pekson R, Jia XF, Yanamandala M, Paulino V, Liang FG, Corbalan JJ, Lee J, Chen Y, Karagiannis GS, Sanchez LR, Liang H, Narayanagari S-R, Mitchell K, Lopez A, Margulets V, Scarlata M, Santulli G, Asnani A, Peterson RT, Hazan RB, Condeelis JS, Oktay MH, Steidl U, Kirshenbaum LA, Gavathiotis E, Kitsis RN. A small-molecule allosteric inhibitor of BAX protects against doxorubicin-induced cardiomyopathy. *Nat Cancer* 2020;**1**:315–328.
- Maslov MY, Chacko VP, Hirsch GA, Akki A, Leppo MK, Steenbergen C, Weiss RG. Reduced in vivo high-energy phosphates precede adriamycin-induced cardiac dysfunction. *Am J Physiol Heart Circ Physiol* 2010;**299**:H332–H337.
- Gupta A, Rohlfen C, Leppo MK, Chacko VP, Wang Y, Steenbergen C, Weiss RG. Creatine kinase-overexpression improves myocardial energetics, contractile dysfunction and survival in murine doxorubicin cardiotoxicity. *PLoS One* 2013;**8**:e74675.
- Suter TM, Ewer MS. Cancer drugs and the heart: importance and management. *Eur Heart J* 2013;**34**:1102–1111.
- De Keulenaer GW, Doggen K, Lemmens K. The vulnerability of the heart as a pluricellular paracrine organ: lessons from unexpected triggers of heart failure in targeted ErbB2 anticancer therapy. *Circ Res* 2010;**106**:35–46.
- Eschenhagen T, Force T, Ewer MS, de Keulenaer GW, Suter TM, Anker SD, Avkiran M, de Azavedo E, Balligand JL, Brutsaert DL, Condorelli G, Hansen A, Heymans S, Hill JA, Hirsch E, Hilfiker-Kleiner D, Janssens S, de Jong S, Neubauer G, Pieske B, Ponikowski P, Pirmohamed M, Rauchhaus M, Sawyer D, Sugden PH, Wojta J, Zannad F, Shah AM. Cardiovascular side effects of cancer therapies: a position statement from the heart failure association of the European Society of Cardiology. *Eur J Heart Fail* 2011;**13**:1–10.
- Moslehi JJ. Cardiovascular toxic effects of targeted cancer therapies. *N Engl J Med* 2016;**375**:1457–1467.
- Luu AZ, Chowdhury B, Al-Omran M, Teoh H, Hess DA, Verma S. Role of endothelium in doxorubicin-induced cardiomyopathy. *JACC Basic Transl Sci* 2018;**3**:861–870.
- Zhang W, St Clair D, Butterfield A, Vore M. Loss of Mrp1 potentiates doxorubicin-induced cytotoxicity in neonatal mouse cardiomyocytes and cardiac fibroblasts. *Toxicol Sci* 2016;**151**:44–56.
- Narikawa M, Umemura M, Tanaka R, Hikichi M, Nagasako A, Fujita T, Yokoyama U, Ishigami T, Kimura K, Tamura K, Ishikawa Y. Doxorubicin induces trans-differentiation and MMP1 expression in cardiac fibroblasts via cell death-independent pathways. *PLoS One* 2019;**14**:e0221940.
- Ghosh AK, Rai R, Park KE, Eren M, Miyata T, Wilsbacher LD, Vaughan DE. A small molecule inhibitor of PAI-1 protects against doxorubicin-induced cellular senescence. *Oncotarget* 2016;**7**:72443–72457.
- Mercurio V, Pirozzi F, Lazzarini E, Marone G, Rizzo P, Agnetti G, Tocchetti CG, Ghigo A, Ameri P. Models of heart failure based on the cardiotoxicity of anticancer drugs. *J Card Fail* 2016;**22**:449–458.
- Vermeulen Z, Segers VF, De Keulenaer GW. ErbB2 signaling at the crossing between heart failure and cancer. *Basic Res Cardiol* 2016;**111**:60.
- Wilkinson EL, Sidaway JE, Cross MJ. Cardiotoxic drugs Herceptin and doxorubicin inhibit cardiac microvascular endothelial cell barrier formation resulting in increased drug permeability. *Biol Open* 2016;**5**:1362–1370.
- Kitani T, Ong SG, Lam CK, Rhee JW, Zhang JZ, Oikonomopoulos A, Ma N, Tian L, Lee J, Telli ML, Witteles RM, Sharma A, Sayed N, Wu JC. Human-induced pluripotent stem cell model of trastuzumab-induced cardiac dysfunction in patients with breast cancer. *Circulation* 2019;**139**:2451–2465.
- Chintalgattu V, Rees ML, Culver JC, Goel A, Jiffar T, Zhang J, Dunner K Jr, Pati S, Bankson JA, Pasqualini R, Arap W, Bryan NS, Taegtmeier H, Langley RR, Yao H, Kupferman ME, Entman ML, Dickinson ME, Khakoo AY. Coronary microvascular pericytes are the cellular target of sunitinib malate-induced cardiotoxicity. *Sci Transl Med* 2013;**5**:187ra69.
- Moslehi J, Minamishima YA, Shi J, Neuberger D, Charytan DM, Padera RF, Signoretti S, Liao R, Kaelin WG Jr. Loss of hypoxia-inducible factor prolyl hydroxylase activity

- in cardiomyocytes phenocopies ischemic cardiomyopathy. *Circulation* 2010;**122**:1004–1016.
37. Bekereditan R, Walton CB, MacCannell KA, Ecker J, Kruse F, Outten JT, Sutcliffe D, Gerard RD, Bruick RK, Shohet RV. Conditional HIF-1 α expression produces a reversible cardiomyopathy. *PLoS One* 2010;**5**:e11693.
 38. Dobbin SJH, Mangion K, Berry C, Roditi G, Basak S, Sourbron S, White J, Venugopal B, Touyz RM, Jones RJ, Petrie MC, Lang NN. Cardiotoxicity and myocardial hypoperfusion associated with anti-vascular endothelial growth factor therapies: prospective cardiac magnetic resonance imaging in patients with cancer. *Eur J Heart Fail* 2020;doi: 10.1002/ehf.1847.
 39. Urazei I, Cheng S, Moslehi J. Reversible cardiomyopathy associated with sunitinib and sorafenib. *N Engl J Med* 2011;**365**:1649–1650.
 40. Kivela R, Hemanthakumar KA, Vaparenta K, Robciuc M, Izumiya Y, Kidoya H, Takakura N, Peng X, Sawyer DB, Elenius K, Walsh K, Alitalo K. Endothelial cells regulate physiological cardiomyocyte growth via VEGFR2-mediated paracrine signaling. *Circulation* 2019;**139**:2570–2584.
 41. Chiusa M, Hool SL, Truetsch P, Djafarzadeh S, Jakob SM, Seifriz F, Scherer SJ, Suter TM, Zuppinger C, Zbinden S. Cancer therapy modulates VEGF signaling and viability in adult rat cardiac microvascular endothelial cells and cardiomyocytes. *J Mol Cell Cardiol* 2012;**52**:1164–1175.
 42. Tocchetti CG, Gallucci G, Coppola C, Piscopo G, Cipresso C, Maurea C, Giudice A, Iaffaioli RV, Arra C, Maurea N. The emerging issue of cardiac dysfunction induced by antineoplastic angiogenesis inhibitors. *Eur J Heart Fail* 2013;**15**:482–489.
 43. Sharma A, Burrige PW, McKeithan WL, Serrano R, Shukla P, Sayed N, Churko JM, Kitani T, Wu H, Holmstrom A, Matsa E, Zhang Y, Kumar A, Fan AC, Del Alamo JC, Wu SM, Moslehi JJ, Mercola M, Wu JC. High-throughput screening of tyrosine kinase inhibitor cardiotoxicity with human induced pluripotent stem cells. *Sci Transl Med* 2017;**9**:eaaf2584.
 44. Paulus WJ, Tschope C. A novel paradigm for heart failure with preserved ejection fraction: comorbidities drive myocardial dysfunction and remodeling through coronary microvascular endothelial inflammation. *J Am Coll Cardiol* 2013;**62**:263–271.
 45. Triposkiadis F, Butler J, Abboud FM, Armstrong PV, Adamopoulos S, Atherton JJ, Backs J, Bauersachs J, Burkhardt D, Bonow RO, Chopra VK, de Boer RA, de Windt L, Hamdani N, Hasenfuss G, Heymans S, Hulot JS, Konstam M, Lee RT, Linke WA, Lunde IG, Lyon AR, Maack C, Mann DL, Mebazaa A, Mentz RJ, Nihoyannopoulos P, Papp Z, Parisiss J, Pedrazzini T, Rosano G, Rouleau J, Seferovic PM, Shah AM, Starling RC, Tocchetti CG, Trochu JN, Thum T, Zannad F, Brutsaert DL, Segers VF, De Keulenaer GW. The continuous heart failure spectrum: moving beyond an ejection fraction classification. *Eur Heart J* 2019;**40**:2155–2163.
 46. Lourenco AP, Leite-Moreira AF, Balligand JL, Bauersachs J, Dawson D, de Boer RA, de Windt LJ, Falcao-Pires I, Fontes-Carvalho R, Franz S, Giacca M, Hilfinger-Kleiner D, Hirsch E, Maack C, Mayr M, Pieske B, Thum T, Tocchetti CG, Brutsaert DL, Heymans S. An integrative translational approach to study heart failure with preserved ejection fraction: a position paper from the working group on myocardial function of the European Society of Cardiology. *Eur J Heart Fail* 2018;**20**:216–227.
 47. Saiki H, Petersen IA, Scott CG, Bailey KR, Dunlay SM, Finley RR, Ruddy KJ, Yan E, Redfield MM. Risk of heart failure with preserved ejection fraction in older women after contemporary radiotherapy for breast cancer. *Circulation* 2017;**135**:1388–1396.
 48. Saiki H, Moulay G, Guenzel AJ, Liu W, Decklever TD, Classic KL, Pham L, Chen HH, Burnett JC, Russell SJ, Redfield MM. Experimental cardiac radiation exposure induces ventricular diastolic dysfunction with preserved ejection fraction. *Am J Physiol Heart Circ Physiol* 2017;**313**:H392–H407.
 49. Mercurio V, Cuomo A, Della Pepa R, Ciervo D, Cella L, Pirozzi F, Parrella P, Campi G, Franco R, Varricchi G, Abete P, Marone G, Petretta M, Bonaduce D, Pacelli R, Picardi M, Tocchetti CG. What is the cardiac impact of chemotherapy and subsequent radiotherapy in lymphoma patients? *Antioxid Redox Signal* 2019;**31**:1166–1174.
 50. Hirsch E, Nagai R, Thum T. Heterocellular signalling and crosstalk in the heart in ischaemia and heart failure. *Cardiovasc Res* 2014;**102**:191–193.
 51. Bang C, Batkai S, Dangwal S, Gupta SK, Foinquinos A, Holzmann A, Just A, Remke J, Zimmer K, Zeug A, Ponimaskin E, Schmiedl A, Yin X, Mayr M, Halder R, Fischer A, Engelhardt S, Wei Y, Schober A, Fiedler J, Thum T. Cardiac fibroblast-derived microRNA passenger strand-enriched exosomes mediate cardiomyocyte hypertrophy. *J Clin Invest* 2014;**124**:2136–2146.
 52. Zhan H, Aizawa K, Sun J, Tomida S, Otsu K, Conway SJ, McKinnon PJ, Manabe I, Komuro I, Miyagawa K, Nagai R, Suzuki T. Ataxia telangiectasia mutated in cardiac fibroblasts regulates doxorubicin-induced cardiotoxicity. *Cardiovasc Res* 2016;**110**:85–95.
 53. Meyer K, Hodwin B, Ramanujam D, Engelhardt S, Sarikas A. Essential role for premature senescence of myofibroblasts in myocardial fibrosis. *J Am Coll Cardiol* 2016;**67**:2018–2028.
 54. Sayed N, Ameen M, Wu JC. Personalized medicine in cardio-oncology: the role of induced pluripotent stem cell. *Cardiovasc Res* 2019;**115**:949–959.
 55. Archer CR, Sergeant R, Basak J, Pilling J, Barnes JR, Pointon A. Characterization and validation of a human 3D cardiac microtissue for the assessment of changes in cardiac pathology. *Sci Rep* 2018;**8**:10160.
 56. Ferdinandy P, Baczkó I, Bencsik P, Giricz Z, Gorbé A, Pachter P, Varga ZV, Varro A, Schulz R. Definition of hidden drug cardiotoxicity: paradigm change in cardiac safety testing and its clinical implications. *Eur Heart J* 2019;**40**:1771–1777.
 57. Ferdinandy P, Hausenloy DJ, Heusch G, Baxter GF, Schulz R. Interaction of risk factors, comorbidities, and comedications with ischemia/reperfusion injury and cardioprotection by preconditioning, postconditioning, and remote conditioning. *Pharmacol Rev* 2014;**66**:1142–1174.
 58. Perrino C, Barabasi AL, Condorelli G, Davidson SM, De Windt L, Dimmeler S, Engel FB, Hausenloy DJ, Hill JA, Van Laake LW, Lecour S, Leor J, Madonna R, Mayr M, Prunier F, Sluijter JGP, Schulz R, Thum T, Ytrehus K, Ferdinandy P. Epigenomic and transcriptomic approaches in the post-genomic era: path to novel targets for diagnosis and therapy of the ischaemic heart? Position paper of the European Society of Cardiology Working Group on Cellular Biology of the Heart. *Cardiovasc Res* 2017;**113**:725–736.
 59. Makkos A, Szantai A, Paloczi J, Pipis J, Kiss B, Poggi P, Ferdinandy P, Chatgililoglu A, Gorbé A. A comorbidity model of myocardial ischemia/reperfusion injury and hypercholesterolemia in rat cardiac myocyte cultures. *Front Physiol* 2020;**10**:1564.
 60. Boer RA, Meijers WC, Meer P, Veldhuisen DJ. Cancer and heart disease: associations and relations. *Eur J Heart Fail* 2019;**21**:1515–1525.
 61. Genovese G, Kahler AK, Handsaker RE, Lindberg J, Rose SA, Bakhoum SF, Chambert K, Mick E, Neale BM, Fromer M, Purcell SM, Svantesson O, Landen M, Hoglund M, Lehmann S, Gabriel SB, Moran JL, Lander ES, Sullivan PF, Sklar P, Gronberg H, Hultman CM, McCarroll SA. Clonal hematopoiesis and blood-cancer risk inferred from blood DNA sequence. *N Engl J Med* 2014;**371**:2477–2487.
 62. Dorsheimer L, Assmus B, Rasper T, Ortmann CA, Ecke A, Abou-El-Ardat K, Schmid T, Brune B, Wagner S, Serve H, Hoffmann J, Seeger F, Dimmeler S, Zeiher AM, Rieger MA. Association of mutations contributing to clonal hematopoiesis with prognosis in chronic ischemic heart failure. *JAMA Cardiol* 2019;**4**:25–33.
 63. Fuster JJ, Walsh K. Somatic mutations and clonal hematopoiesis: unexpected potential new drivers of age-related cardiovascular disease. *Circ Res* 2018;**122**:523–532.
 64. Jaiswal S, Natarajan P, Silver AJ, Gibson CJ, Bick AG, Shvartz E, McConkey M, Gupta N, Gabriel S, Ardissino D, Baber U, Mehran R, Fuster V, Danesh J, Frossard P, Saleheen D, Melander O, Sukhova GK, Neuberg D, Libby P, Kathiresan S, Ebert BL. Clonal hematopoiesis and risk of atherosclerotic cardiovascular disease. *N Engl J Med* 2017;**377**:111–121.
 65. Jaiswal S, Fontanillas P, Flannick J, Manning A, Grauman PV, Mar BG, Lindsley RC, Mermel CH, Burt N, Chavez A, Higgins JM, Moltchanov V, Kuo FC, Kluk MJ, Henderson B, Kinnunen L, Koistinen HA, Ladvall C, Getz G, Correa A, Banahan BF, Gabriel S, Kathiresan S, Stringham HM, McCarthy MI, Boehnke M, Tuomilehto J, Haiman C, Groop L, Atzmon G, Wilson JG, Neuberg D, Altshuler D, Ebert BL. Age-related clonal hematopoiesis associated with adverse outcomes. *N Engl J Med* 2014;**371**:2488–2498.
 66. Libby P, Sidlow R, Lin AE, Gupta D, Jones LW, Moslehi J, Zeiher A, Jaiswal S, Schulz C, Blankstein R, Bolton KL, Steensma D, Levine RL, Ebert BL. Clonal hematopoiesis: crossroads of aging, cardiovascular disease, and cancer: JACC review topic of the week. *J Am Coll Cardiol* 2019;**74**:567–577.
 67. Yura Y, Sano S, Walsh K. Clonal hematopoiesis: a new step linking inflammation to heart failure. *JACC Basic Transl Sci* 2020;**5**:196–207.
 68. Garcia-Pavia P, Kim Y, Restrepo-Cordoba MA, Lunde IG, Wakimoto H, Smith AM, Toepfer CN, Getz K, Gorham J, Patel P, Ito K, Willcox JA, Arany Z, Li J, Owens AT, Govind R, Nuñez B, Mazaika E, Bayes-Genis A, Walsh R, Finkelman B, Lupon J, Whiffin N, Serrano I, Midwinter W, Wilk A, Bardaji A, Ingold N, Buchan R, Tayal U, Pascual-Figal DA, de Marvao A, Ahmad M, Garcia-Pinilla JM, Pantazis A, Dominguez F, John Baksi A, O'Regan DP, Rosen SD, Prasad SK, Lara-Pezzi E, Provencio M, Lyon AR, Alonso-Pulpon L, Cook SA, DePalma SR, Barton PJR, Aplenc R, Seidman JG, Ky B, Ware JS, Seidman CE. Genetic variants associated with cancer therapy-induced cardiomyopathy. *Circulation* 2019;**140**:31–41.
 69. de Boer RA, De Keulenaer G, Bauersachs J, Brutsaert D, Cleland JG, Diez J, Du XJ, Ford P, Heinzel FR, Lipson KE, McDonagh T, Lopez-Andres N, Lunde IG, Lyon AR, Pollesello P, Prasad SK, Tocchetti CG, Mayr M, Sluijter JGP, Thum T, Tschope C, Zannad F, Zimmermann WH, Ruschitzka F, Filippatos G, Lindsey ML, Maack C, Heymans S. Towards better definition, quantification and treatment of fibrosis in heart failure. A scientific roadmap by the committee of translational research of the Heart Failure Association (HFA) of the European Society of Cardiology. *Eur J Heart Fail* 2019;**21**:272–285.
 70. Li M, Caeyenberghs K. Longitudinal assessment of chemotherapy-induced changes in brain and cognitive functioning: a systematic review. *Neurosci Biobehav Rev* 2018;**92**:304–317.
 71. Ogren JA, Fonarow GC, Woo MA. Cerebral impairment in heart failure. *Curr Heart Fail Rep* 2014;**11**:321–329.
 72. Bertero E, Canepa M, Maack C, Ameri P. Linking heart failure to cancer. *Circulation* 2018;**138**:735–742.
 73. Libby P, Koblold S. Inflammation: a common contributor to cancer, aging, and cardiovascular diseases-expanding the concept of cardio-oncology. *Cardiovasc Res* 2019;**115**:824–829.
 74. Prabhu SD, Frangogiannis NG. The biological basis for cardiac repair after myocardial infarction: from inflammation to fibrosis. *Circ Res* 2016;**119**:91–112.
 75. Wang L, Chen Q, Qi H, Wang C, Wang C, Zhang J, Dong L. Doxorubicin-induced systemic inflammation is driven by upregulation of toll-like receptor TLR4 and endotoxin leakage. *Cancer Res* 2016;**76**:6631–6642.

76. Pecoraro M, Del Pizzo M, Marzocco S, Sorrentino R, Ciccarelli M, Iaccarino G, Pinto A, Popolo A. Inflammatory mediators in a short-time mouse model of doxorubicin-induced cardiotoxicity. *Toxicol Appl Pharmacol* 2016;**293**:44–52.
77. Nozaki N, Shishido T, Takeishi Y, Kubota I. Modulation of doxorubicin-induced cardiac dysfunction in toll-like receptor-2-knockout mice. *Circulation* 2004;**110**:2869–2874.
78. van Almen GC, Swinnen M, Carai P, Verhesen W, Cleutjens JP, D'Hooge J, Verheyen FK, Pinto YM, Schroen B, Carmeliet P, Heymans S. Absence of thrombospondin-2 increases cardiomyocyte damage and matrix disruption in doxorubicin-induced cardiomyopathy. *J Mol Cell Cardiol* 2011;**51**:318–328.
79. Ma Y, Mattarollo SR, Adjemian S, Yang H, Aymeric L, Hannani D, Portela Catani JP, Duret H, Teng MW, Kepp O, Wang Y, Sistig A, Schultze JL, Stoll G, Galluzzi L, Zitvogel L, Smyth MJ, Kroemer G. CCL2/CCR2-dependent recruitment of functional antigen-presenting cells into tumors upon chemotherapy. *Cancer Res* 2014;**74**:436–445.
80. Lesterhuis WJ, Haanen JB, Punt CJ. Cancer immunotherapy—revisited. *Nat Rev Drug Discov* 2011;**10**:591–600.
81. Varricchi G, Galdiero MR, Tocchetti CG. Cardiac toxicity of immune checkpoint inhibitors: cardio-oncology meets immunology. *Circulation* 2017;**136**:1989–1992.
82. Hu JR, Florido R, Lipson EJ, Naidoo J, Ardehali R, Tocchetti CG, Lyon AR, Padera RF, Johnson DB, Moslehi J. Cardiovascular toxicities associated with immune checkpoint inhibitors. *Cardiovasc Res* 2019;**115**:854–868.
83. Lyon AR, Yousaf N, Battisti NML, Moslehi J, Larkin J. Immune checkpoint inhibitors and cardiovascular toxicity. *Lancet Oncol* 2018;**19**:e447–e458.
84. Escudier M, Cautela J, Malissen N, Ancedy Y, Orabona N, Pinto J, Monestier S, Grob JJ, Scemama U, Jacquier A, Lavee N, Barraud J, Peyrol M, Laine M, Bonello L, Paganelli F, Cohen A, Barlesi F, Ederhy S, Thuny F. Clinical features, management, and outcomes of immune checkpoint inhibitor-related cardiotoxicity. *Circulation* 2017;**136**:2085–2087.
85. Nishimura H, Okazaki T, Tanaka Y, Nakatani K, Hara M, Matsumori A, Sasayama S, Mizoguchi A, Hiai H, Minato N, Honjo T. Autoimmune dilated cardiomyopathy in PD-1 receptor-deficient mice. *Science* 2001;**291**:319–322.
86. Varricchi G, Marone G, Mercurio V, Galdiero MR, Bonaduce D, Tocchetti CG. Immune checkpoint inhibitors and cardiac toxicity: an emerging issue. *Curr Med Chem* 2018;**25**:1327–1339.
87. Awadalla M, Mahmood SS, Groarke JD, Hassan MZO, Nohria A, Rokicki A, Murphy SP, Mercaldo ND, Zhang L, Zlotoff DA, Reynolds KL, Alvi RM, Banerji D, Liu S, Heinzerling LM, Jones-O'Connor M, Bakar RB, Cohen JV, Kirchberger MC, Sullivan RJ, Gupta D, Mulligan CP, Shah SP, Ganatra S, Rizvi MA, Sahni G, Tocchetti CG, Lawrence DP, Mahmoudi M, Devereux RB, Forrester BJ, Mandawat A, Lyon AR, Chen CL, Barac A, Hung J, Thavendiranathan P, Picard MH, Thuny F, Ederhy S, Fradley MG, Neilan TG. Global longitudinal strain and cardiac events in patients with immune checkpoint inhibitor-related myocarditis. *J Am Coll Cardiol* 2020;**75**:467–478.
88. Salem JE, Allenbach Y, Vozy A, Brechot N, Johnson DB, Moslehi JJ, Kerneis M. Abatacept for severe immune checkpoint inhibitor-associated myocarditis. *N Engl J Med* 2019;**380**:2377–2379.
89. Zhang L, Zlotoff DA, Awadalla M, Mahmood SS, Nohria A, Hassan MZO, Thuny F, Zubiri L, Chen CL, Sullivan RJ, Alvi RM, Rokicki A, Murphy SP, Jones-O'Connor M, Heinzerling LM, Barac A, Forrester BJ, Yang EH, Gupta D, Kirchberger MC, Shah SP, Rizvi MA, Sahni G, Mandawat A, Mahmoudi M, Ganatra S, Ederhy S, Zatarain-Nicolas E, Groarke JD, Tocchetti CG, Lyon AR, Thavendiranathan P, Cohen JV, Reynolds KL, Fradley MG, Neilan TG. Major adverse cardiovascular events and the timing and dose of corticosteroids in immune checkpoint inhibitor-associated myocarditis. *Circulation* 2020;**141**:2031–2034.
90. Ganatra S, Carver JR, Hayek SS, Ky B, Leja MJ, Lenihan DJ, Lenneman C, Mousavi N, Park JH, Perales MA, Ryan TD, Scherrer-Crosbie M, Steingart RM, Yang EH, Zaha V, Barac A, Liu JE. Chimeric antigen receptor T-cell therapy for cancer and heart: JACC Council Perspectives. *J Am Coll Cardiol* 2019;**74**:3153–3163.
91. June CH, Sadelain M. Chimeric antigen receptor therapy. *N Engl J Med* 2018;**379**:64–73.
92. Riegler LL, Jones GP, Lee DW. Current approaches in the grading and management of cytokine release syndrome after chimeric antigen receptor T-cell therapy. *Ther Clin Risk Manag* 2019;**15**:323–335.
93. Alvi RM, Frigault MJ, Fradley MG, Jain MD, Mahmood SS, Awadalla M, Lee DH, Zlotoff DA, Zhang L, Drobni ZD, Hassan MZO, Bassily E, Rhea I, Ismail-Khan R, Mulligan CP, Banerji D, Lazaryan A, Shah BD, Rokicki A, Raju N, Chavez JC, Abramson J, Locke FL, Neilan TG. Cardiovascular events among adults treated with chimeric antigen receptor T-cells (CAR-T). *J Am Coll Cardiol* 2019;**74**:3099–3108.
94. Aldoss I, Khaled SK, Budde E, Stein AS. Cytokine release syndrome with the novel treatments of acute lymphoblastic leukemia: pathophysiology, prevention, and treatment. *Curr Oncol Rep* 2019;**21**:4.
95. Mantovani A, Allavena P, Sica A, Balkwill F. Cancer-related inflammation. *Nature* 2008;**454**:436–444.
96. Galdiero MR, Garlanda C, Jaillon S, Marone G, Mantovani A. Tumor associated macrophages and neutrophils in tumor progression. *J Cell Physiol* 2013;**228**:1404–1412.
97. van't Klooster CC, Ridker PM, Hjortnaes J, van der Graaf Y, Asselbergs FW, Westerink J, Aerts JGJV, Visseren FLJ. The relation between systemic inflammation and incident cancer in patients with stable cardiovascular disease: a cohort study. *Eur Heart J* 2019;**40**:3901–3909.
98. Ridker PM, MacFadyen JG, Thuren T, Everett BM, Libby P, Glynn RJ; CANTOS Trial Group. Effect of interleukin-1 β inhibition with canakinumab on incident lung cancer in patients with atherosclerosis: exploratory results from a randomised, double-blind, placebo-controlled trial. *Lancet* 2017;**390**:1833–1842.
99. Ridker PM, Everett BM, Thuren T, MacFadyen JG, Chang WH, Ballantyne C, Fonseca F, Nicolau J, Koenig W, Anker SD, Kastelein JJP, Cornel JH, Pais P, Pella D, Genest J, Cifkova R, Lorenzatti A, Forster T, Kobalava Z, Vida-Simiti L, Flather M, Shimokawa H, Ogawa H, Dellborg M, Rossi PRF, Troquay RPT, Libby P, Glynn RJ; CANTOS Trial Group. Antiinflammatory therapy with canakinumab for atherosclerotic disease. *N Engl J Med* 2017;**377**:1119–1131.
100. Sauter KA, Wood LJ, Wong J, Jordanov M, Magun BE. Doxorubicin and daunorubicin induce processing and release of interleukin-1 β through activation of the NLRP3 inflammasome. *Cancer Biol Ther* 2011;**11**:1008–1016.
101. Zhu J, Zhang J, Xiang D, Zhang Z, Zhang L, Wu M, Zhu S, Zhang R, Han W. Recombinant human interleukin-1 receptor antagonist protects mice against acute doxorubicin-induced cardiotoxicity. *Eur J Pharmacol* 2010;**643**:247–253.
102. Ghigo A, Damilano F, Braccini L, Hirsch E. PI3K inhibition in inflammation: toward tailored therapies for specific diseases. *Bioessays* 2010;**32**:185–196.
103. Fougerat A, Gayral S, Gourdy P, Schambourg A, Ruckl T, Schwarz MK, Rommel C, Hirsch E, Arnal J-F, Salles J-P, Perret B, Breton-Douillon M, Wymann MP, Laffargue M. Genetic and pharmacological targeting of phosphoinositide 3-kinase-gamma reduces atherosclerosis and favors plaque stability by modulating inflammatory processes. *Circulation* 2008;**117**:1310–1317.
104. Damilano F, Franco I, Perrino C, Schaefer K, Azzolino O, Carnevale D, Cifelli G, Carullo P, Ragona R, Ghigo A, Perino A, Lembo G, Hirsch E. Distinct effects of leukocyte and cardiac phosphoinositide 3-kinase gamma activity in pressure overload-induced cardiac failure. *Circulation* 2011;**123**:391–399.
105. Kaneda MM, Messer KS, Ralainirina N, Li H, Leem CJ, Gorjestani S, Woo G, Nguyen AV, Figueiredo CC, Foubert P, Schmid MC, Pink M, Winkler DG, Rausch M, Palombella VJ, Kutok J, McGovern K, Frazer KA, Wu X, Karin M, Sasik R, Cohen EE, Varner JA. PI3K γ is a molecular switch that controls immune suppression. *Nature* 2016;**539**:437–442.
106. Li M, Sala V, De Santis MC, Cimino J, Cappello P, Pianca N, Di Bona A, Margaria JP, Martini M, Lazzarini E, Pirozzi F, Rossi L, Franco I, Bornbaum J, Heger J, Rohrbach S, Perino A, Tocchetti CG, Lima BHF, Teixeira MM, Porporato PE, Schulz R, Angelini A, Sandri M, Ameri P, Sciarretta S, Lima-Junior RCP, Mongillo M, Zaglia T, Morello F, Novelli F, Hirsch E, Ghigo A. Phosphoinositide 3-kinase gamma inhibition protects from anthracycline cardiotoxicity and reduces tumor growth. *Circulation* 2018;**138**:696–711.
107. Gangadhara G, Dahl G, Bohnacker T, Rae R, Gunnarsson J, Blaho S, Oster L, Lindmark H, Karabelas K, Pemberton N, Tyrchan C, Mogemark M, Wymann MP, Williams RL, Perry MWD, Papavoine T, Petersen J. A class of highly selective inhibitors bind to an active state of PI3K γ . *Nat Chem Biol* 2019;**15**:348–357.
108. Anker SD, Egerer KR, Volk HD, Kox WJ, Poole-Wilson PA, Coats AJ. Elevated soluble CD14 receptors and altered cytokines in chronic heart failure. *Am J Cardiol* 1997;**79**:1426–1430.
109. Krack A, Sharma R, Figulla HR, Anker SD. The importance of the gastrointestinal system in the pathogenesis of heart failure. *Eur Heart J* 2005;**26**:2368–2374.
110. Tang WHW, Li DY, Hazen SL. Dietary metabolism, the gut microbiome, and heart failure. *Nat Rev Cardiol* 2019;**16**:137–154.
111. Backhed F, Roswall J, Peng Y, Feng Q, Jia H, Kovatcheva-Datchary P, Li Y, Xia Y, Xie H, Zhong H, Khan MT, Zhang J, Li J, Xiao L, Al-Aama J, Zhang D, Lee YS, Kotowska D, Colding C, Tremaroli V, Yin Y, Bergman S, Xu X, Madsen L, Kristiansen K, Dahlgren J, Wang J. Dynamics and stabilization of the human gut microbiome during the first year of life. *Cell Host Microbe* 2015;**17**:690–703.
112. Dominguez-Bello MG, Costello EK, Contreras M, Magris M, Hidalgo G, Fierer N, Knight R. Delivery mode shapes the acquisition and structure of the initial microbiota across multiple body habitats in newborns. *Proc Natl Acad Sci USA* 2010;**107**:11971–11975.
113. Goodrich JK, Waters JL, Poole AC, Sutter JL, Koren O, Blekhan R, Beaumont M, Van Treuren W, Knight R, Bell JT, Spector TD, Clark AG, Ley RE. Human genetics shape the gut microbiome. *Cell* 2014;**159**:789–799.
114. Kinross JM, Darzi AW, Nicholson JK. Gut microbiome-host interactions in health and disease. *Genome Med* 2011;**3**:14.
115. Zitvogel L, Dailere R, Roberti MP, Routy B, Kroemer G. Anticancer effects of the microbiome and its products. *Nat Rev Microbiol* 2017;**15**:465–478.
116. Nakatsu G, Li X, Zhou H, Sheng J, Wong SH, Wu WK, Ng SC, Tsoi H, Dong Y, Zhang N, He Y, Kang Q, Cao L, Wang K, Zhang J, Liang Q, Yu J, Sung JJ. Gut mucosal microbiome across stages of colorectal carcinogenesis. *Nat Commun* 2015;**6**:8727.
117. Mitra A, MacIntyre DA, Lee YS, Smith A, Marchesi JR, Lehne B, Bhatia R, Lyons D, Paraskevidis E, Li JV, Holmes E, Nicholson JK, Bennett PR, Kyrgiou M. Cervical intraepithelial neoplasia disease progression is associated with increased vaginal microbiome diversity. *Sci Rep* 2015;**5**:16865.
118. Yu G, Gail MH, Consonni D, Carugno M, Humphrys M, Pesatori AC, Caporaso NE, Goedert JJ, Ravel J, Landi MT. Characterizing human lung tissue microbiota and its relationship to epidemiological and clinical features. *Genome Biol* 2016;**17**:163.

119. Karin M, Jobin C, Balkwill F. Chemotherapy, immunity and microbiota—a new triumvirate? *Nat Med* 2014;**20**:126–127.
120. Roy S, Trinchieri G. Microbiota: a key orchestrator of cancer therapy. *Nat Rev Cancer* 2017;**17**:271–285.
121. Russo M, Guida F, Paparo L, Trinchese G, Aitoro R, Avagliano C, Fiordelisi A, Napolitano F, Mercurio V, Sala V, Li M, Sorriento D, Ciccarelli M, Ghigo A, Hirsch E, Bianco R, Iaccarino G, Abete P, Bonaduce D, Calignano A, Berni Canani R, Tocchetti CG. The novel butyrate derivative phenylalanine-butyramide protects from doxorubicin-induced cardiotoxicity. *Eur J Heart Fail* 2019;**21**:519–528.
122. Paparo L, Calignano A, Tocchetti CG, Di Scala C, Russo R, Bonaduce D, Canani RB. The influence of fiber on gut microbiota: butyrate as molecular player involved in the beneficial interplay between dietary fiber and cardiovascular health. In Saman R (ed). *Dietary Fiber for the Prevention of Cardiovascular Disease: Fiber's Interaction between Gut Microflora, Sugar Metabolism, Weight Control and Cardiovascular Health*. 1st ed, Academic Press, 2017. pp. 61–71.
123. Gil-Cruz C, Perez-Shibayama C, De Martin A, Ronchi F, van der Borght K, Niederer R, Onder L, Lutge M, Novkovic M, Nindl V, Ramos G, Arnoldini M, Slack EMC, Boivin-Jahns V, Jahns R, Wyss M, Mooser C, Lambrecht BN, Maeder MT, Rickli H, Flatz L, Eriksson U, Geuking MB, McCoy KD, Ludewig B. Microbiota-derived peptide mimics drive lethal inflammatory cardiomyopathy. *Science* 2019;**366**:881–886.
124. Tang WH, Kitai T, Hazen SL. Gut microbiota in cardiovascular health and disease. *Circ Res* 2017;**120**:1183–1196.
125. Kim MH, Kang SG, Park JH, Yanagisawa M, Kim CH. Short-chain fatty acids activate GPR41 and GPR43 on intestinal epithelial cells to promote inflammatory responses in mice. *Gastroenterology* 2013;**145**:396–406.e1–10.
126. Maslowski KM, Vieira AT, Ng A, Kranich J, Sierro F, Yu D, Schilter HC, Rolph MS, Mackay F, Artis D, Xavier RJ, Teixeira MM, Mackay CR. Regulation of inflammatory responses by gut microbiota and chemoattractant receptor GPR43. *Nature* 2009;**461**:1282–1286.
127. Brown AJ, Goldsworthy SM, Barnes AA, Eilert MM, Tcheang L, Daniels D, Muir AI, Wigglesworth MJ, Kinghorn I, Fraser NJ, Pike NB, Strum JC, Stepelwski KM, Murdock PR, Holder JC, Marshall FH, Szekeres PG, Wilson S, Ignar DM, Foord SM, Wise A, Dowell SJ. The orphan G protein-coupled. *J Biol Chem* 2003;**278**:11312–11319.
128. Le Poul E, Loison C, Struyf S, Springael JY, Lannoy V, Decobecq ME, Brezillon S, Dupriez V, Vassart G, Van Damme J, Parmentier M, Dethieux M. Functional characterization of human receptors for short chain fatty acids and their role in polymorphonuclear cell activation. *J Biol Chem* 2003;**278**:25481–25489.
129. Li L, Hua Y, Ren J. Short-chain fatty acid propionate alleviates Akt2 knockout-induced myocardial contractile dysfunction. *Exp Diabetes Res* 2012;**2012**:851717.
130. Mujumdar VS, Tummalaipalli CM, Aru GM, Tyagi SC. Mechanism of constrictive vascular remodeling by homocysteine: role of PPAR. *Am J Physiol Cell Physiol* 2002;**282**:C1009–C1015.
131. Ali FY, Armstrong PC, Dhanji AR, Tucker AT, Paul-Clark MJ, Mitchell JA, Warner TD. Antiplatelet actions of statins and fibrates are mediated by PPARs. *Arterioscler Thromb Vasc Biol* 2009;**29**:706–711.
132. Antos CL, McKinsey TA, Dreitz M, Hollingsworth LM, Zhang CL, Schreiber K, Rindt H, Gorkczynski RJ, Olson EN. Dose-dependent blockade to cardiomyocyte hypertrophy by histone deacetylase inhibitors. *J Biol Chem* 2003;**278**:28930–28937.
133. Gallo P, Latronico MV, Gallo P, Grimaldi S, Borgia F, Todaro M, Jones P, Gallinari P, De Francesco R, Ciliberto G, Steinkuhler C, Esposito G, Condorelli G. Inhibition of class I histone deacetylase with an apicidin derivative prevents cardiac hypertrophy and failure. *Cardiovasc Res* 2008;**80**:416–424.
134. Granger A, Abdullah I, Huebner F, Stout A, Wang T, Huebner T, Epstein JA, Gruber PJ. Histone deacetylase inhibition reduces myocardial ischemia-reperfusion injury in mice. *FASEB J* 2008;**22**:3549–3560.
135. Kong Y, Tannous P, Lu G, Berenji K, Rothermel BA, Olson EN, Hill JA. Suppression of class I and II histone deacetylases blunts pressure-overload cardiac hypertrophy. *Circulation* 2006;**113**:2579–2588.
136. Rephaeli A, Rabizadeh E, Aviram A, Shklai M, Ruse M, Nudelman A. Derivatives of butyric acid as potential anti-neoplastic agents. *Int J Cancer* 1991;**49**:66–72.
137. Rephaeli A, Waks-Yona S, Nudelman A, Tarasenko I, Tarasenko N, Phillips DR, Cutts SM, Kessler-Ickson G. Anticancer prodrugs of butyric acid and formaldehyde protect against doxorubicin-induced cardiotoxicity. *Br J Cancer* 2007;**96**:1667–1674.
138. Fu HY, Sanada S, Matsuzaki T, Liao Y, Okuda K, Yamato M, Tsuchida S, Araki R, Asano Y, Asanuma H, Asakura M, French BA, Sakata Y, Kitakaze M, Minamino T. Chemical endoplasmic reticulum chaperone alleviates doxorubicin-induced cardiac dysfunction. *Circ Res* 2016;**118**:798–809.
139. Daosukho C, Chen Y, Noel T, Sompol P, Nithipongvanitch R, Velez JM, Oberley TD, St Clair DK. Phenylbutyrate, a histone deacetylase inhibitor, protects against Adriamycin-induced cardiac injury. *Free Radic Biol Med* 2007;**42**:1818–1825.
140. Huang K, Liu Y, Tang H, Qiu M, Li C, Duan C, Wang C, Yang J, Zhou X. Glabridin prevents doxorubicin-induced cardiotoxicity through gut microbiota modulation and colonic macrophage polarization in mice. *Front Pharmacol* 2019;**10**:107.
141. Nishimura Y, Kondo C, Morikawa Y, Tonomura Y, Torii M, Yamate J, Uehara T. Plasma miR-208 as a useful biomarker for drug-induced cardiotoxicity in rats. *J Appl Toxicol* 2015;**35**:173–180.
142. Rigaud VR, Ferreira LR, Ayub-Ferreira SM, Avila MS, Brandao SM, Cruz FD, Santos MH, Cruz CB, Alves MS, Issa VS, Guimaraes GV, Cunha-Neto E, Bocchi EA. Circulating miR-1 as a potential biomarker of doxorubicin-induced cardiotoxicity in breast cancer patients. *Oncotarget* 2017;**8**:6994–7002.
143. Gioffre S, Ricci V, Vavassori C, Ruggeri C, Chiesa M, Alfieri I, Zorzan S, Buzzetti M, Milano G, Scopece A, Castiglioni L, Sironi L, Pompilio G, Colombo GI, D'Alessandra Y. Plasmatic and chamber-specific modulation of cardiac microRNAs in an acute model of dox-induced cardiotoxicity. *Biomed Pharmacother* 2019;**110**:1–8.
144. Piegari E, Russo R, Cappetta D, Esposito G, Urbanek K, Dell'Aversana C, Altucci L, Berrino L, Rossi F, De Angelis A. MicroRNA-34a regulates doxorubicin-induced cardiotoxicity in rat. *Oncotarget* 2016;**7**:62312–62326.
145. Desai VG, J CK, Vijay V, Moland CL, Herman EH, Lee T, Han T, Lewis SM, Davis KJ, Muskhelishvili L, Kerr S, Fuscoe JC. Early biomarkers of doxorubicin-induced heart injury in a mouse model. *Toxicol Appl Pharmacol* 2014;**281**:221–229.
146. Horak M, Novak J, Bienertova-Vasku J. Muscle-specific microRNAs in skeletal muscle development. *Develop Biol* 2016;**410**:1–13.
147. Gomes CP, Oliveira GP Jr, Madrid B, Almeida JA, Franco OL, Pereira RW. Circulating miR-1, miR-133a, and miR-206 levels are increased after a half-marathon run. *Biomarkers* 2014;**19**:585–589.
148. Navicks R, Gal D, Laucevicius A, Taparuskaitė A, Zdanytė M, Holvoet P. Identifying circulating microRNAs as biomarkers of cardiovascular disease: a systematic review. *Cardiovasc Res* 2016;**111**:322–337.
149. Coenen-Stass AML, Wood MJA, Roberts TC. Biomarker potential of extracellular miRNAs in Duchenne muscular dystrophy. *Trends Mol Med* 2017;**23**:989–1001.
150. Ruggeri C, Bouffré S, Chiesa M, Buzzetti M, Milano G, Scopece A, Castiglioni L, Pontremoli M, Sironi L, Pompilio G, Colombo GI, D'Alessandra Y. A specific circulating microRNA cluster is associated to late differential cardiac response to doxorubicin-induced cardiotoxicity in vivo. *Dis Markers* 2018;**2018**:1–9.
151. Freres P, Bouznad N, Servais L, Josse C, Wenric S, Poncin A, Thiry J, Moonen M, Oury C, Lancellotti P, Bours V, Jerusalem G. Variations of circulating cardiac biomarkers during and after anthracycline-containing chemotherapy in breast cancer patients. *BMC Cancer* 2018;**18**:102.
152. Fiedler J, Gupta SK, Thum T. Identification of cardiovascular microRNA targetomes. *J Mol Cell Cardiol* 2011;**51**:674–681.
153. Yin Z, Zhao Y, Li H, Yan M, Zhou L, Chen C, Wang DW. MiR-320a mediates doxorubicin-induced cardiotoxicity by targeting VEGF signal pathway. *Aging* 2016;**8**:192–207.
154. Chatterjee S, Gupta SK, Bar C, Thum T. Noncoding RNAs: potential regulators in cardiology. *Am J Physiol Heart Circ Physiol* 2019;**316**:H160–H168.
155. Gupta SK, Garg A, Avramopoulos P, Engelhardt S, Streckfuss-Bomeke K, Batkai S, Thum T. miR-212/132 cluster modulation prevents doxorubicin-mediated atrophy and cardiotoxicity. *Mol Ther* 2019;**27**:17–28.
156. Gupta SK, Garg A, Bär C, Chatterjee S, Foinquinos A, Milting H, Streckfuß-Bömeke K, Fiedler J, Thum T. Quaking inhibits doxorubicin-mediated cardiotoxicity through regulation of cardiac circular RNA expression. *Circ Res* 2018;**122**:246–254.
157. Roca-Alonso L, Castellano L, Mills A, Dabrowska AF, Sikkel MB, Pellegrino L, Jacob J, Frampton AE, Krell J, Coombes RC, Harding SE, Lyon AR, Stebbing J. Myocardial miR-30 downregulation triggered by doxorubicin drives alterations in beta-adrenergic signaling and enhances apoptosis. *Cell Death Dis* 2015;**6**:e1754.
158. Oatmen KE, Toro-Salazar OH, Hauser K, Zellars KN, Mason KC, Hor K, Gillan E, Zeiss CJ, Gatti DM, Spinale FG. Identification of a novel microRNA profile in pediatric patients with cancer treated with anthracycline chemotherapy. *Am J Physiol Heart Circ Physiol* 2018;**315**:H1443–H1452.
159. Bae S, Ulrich CM, Neuhauser ML, Malysheva O, Bailey LB, Xiao L, Brown EC, Cushing-Haugen KL, Zheng Y, Cheng TY, Miller JW, Green R, Lane DS, Beresford SA, Caudill MA. Plasma choline metabolites and colorectal cancer risk in the women's health initiative observational study. *Cancer Res* 2014;**74**:7442–7452.
160. Mondul AM, Moore SC, Weinstein SJ, Karoly ED, Sampson JN, Albanes D. Metabolomic analysis of prostate cancer risk in a prospective cohort: the alpha-tocopherol, beta-carotene cancer prevention (ATBC) study. *Int J Cancer* 2015;**137**:2124–2132.
161. Wang C, Jing Q. Non-coding RNAs as biomarkers for acute myocardial infarction. *Acta Pharmacol Sin* 2018;**39**:1110–1119.
162. Chistiakov DA, Orekhov AN, Bobryshev YV. Cardiac-specific miRNA in cardiogenesis, heart function, and cardiac pathology (with focus on myocardial infarction). *J Mol Cell Cardiol* 2016;**94**:107–121.
163. Kura B, Kalocayova B, Devaux Y, Bartekova M. Potential clinical implications of miR-1 and miR-21 in heart disease and cardioprotection. *Int J Mol Sci* 2020;**21**:700.
164. Kashyap D, Kaur H. Cell-free miRNAs as non-invasive biomarkers in breast cancer: significance in early diagnosis and metastasis prediction. *Life Sci* 2020;**246**:117417.
165. Sheervailou R, Lotfi H, Shirvaliloo M, Sharifi A, Nazemiyeh M, Zarghami N. Circulating miR-10b, miR-1 and miR-30a expression profiles in lung cancer: possible correlation with clinico-pathologic characteristics and lung cancer detection. *Int J Mol Cell Med* 2019;**8**:118–129.
166. Peng J, Yuan C, Wu Z, Wang Y, Yin W, Zhou L, Lu J. Upregulation of microRNA1 inhibits proliferation and metastasis of breast cancer. *Mol Med Rep* 2020;**22**:454–464.
167. Han C, Shen JK, Hornicek FJ, Kan Q, Duan Z. Regulation of microRNA-1 (miR-1) expression in human cancer. *Biochim Biophys Acta Gene Regul Mech* 2017;**1860**:227–232.

168. Fulzele S, Mendhe B, Khayrullin A, Johnson M, Kaiser H, Liu Y, Isaacs CM, Hamrick MW. Muscle-derived miR-34a increases with age in circulating extracellular vesicles and induces senescence of bone marrow stem cells. *Aging* 2019;**11**:1791–1803.
169. Gioffre S, Chiesa M, Cardinale DM, Ricci V, Vavassori C, Cipolla CM, Masson S, Sandri MT, Salvatici M, Ciceri F, Latini R, Staszewsky LI, Pompilio G, Colombo GI, D'Alessandra Y. Circulating microRNAs as potential predictors of anthracycline-induced troponin elevation in breast cancer patients: diverging effects of doxorubicin and epirubicin. *J Clin Med* 2020;**9**:1418. doi: 10.3390/jcm9051418.
170. Boon RA, Iekushi K, Lechner S, Seeger T, Fischer A, Heydt S, Kaluza D, Treguer K, Carmona G, Bonauer A, Horrevoets AJ, Didier N, Girmatsion Z, Biliczki P, Ehrlich JR, Katus HA, Muller OJ, Potente M, Zeiher AM, Hermeking H, Dimmeler S. MicroRNA-34a regulates cardiac ageing and function. *Nature* 2013;**495**:107–110.
171. Zhu JN, Fu YH, Hu ZQ, Li WY, Tang CM, Fei HW, Yang H, Lin QX, Gou DM, Wu SL, Shan ZX. Activation of miR-34a-5p/sirt1/p66shc pathway contributes to doxorubicin-induced cardiotoxicity. *Sci Rep* 2017;**7**:11879.
172. Lin Y, Lin Z, Fang Z, Li H, Zhi X, Zhang Z. Plasma microRNA-34a as a potential biomarker for early diagnosis of esophageal cancer. *Clin Lab* 2019;**65**.
173. Zhang L, Liao Y, Tang L. MicroRNA-34 family: a potential tumor suppressor and therapeutic candidate in cancer. *J Exp Clin Cancer Res* 2019;**38**:53.
174. Tokumaru Y, Katsuta E, Oshi M, Sporn JC, Yan L, Le L, Matsuhashi N, Futamura M, Akao Y, Yoshida K, Takabe K. High expression of miR-34a associated with less aggressive cancer biology but not with survival in breast cancer. *Int J Med Sci* 2020;**21**:3045.
175. Sommariva E, D'Alessandra Y, Farina FM, Casella M, Cattaneo F, Catto V, Chiesa M, Stadiotti I, Brambilla S, Dello Russo A, Carbuicchio C, Vettor G, Riggio D, Sandri MT, Barbuti A, Vernillo G, Muratori M, Dal Ferro M, Sinagra G, Moimas S, Giacca M, Colombo GI, Pompilio G, Tondo C. MiR-320a as a potential novel circulating biomarker of arrhythmic cardiomyopathy. *Sci Rep* 2017;**7**:4802. doi: 10.1038/s41598-017-05001-z.
176. Galeano-Otero I, Del Toro R, Guisado A, Díaz I, Mayoral-González I, Guerrero-Márquez F, Gutiérrez-Carretero E, Casquero-Domínguez S, Díaz-de la Llera L, Barón-Esquivias G, Jiménez-Navarro M, Smani T, Ordóñez-Fernández A. Circulating miR-320a as a predictive biomarker for left ventricular remodelling in STEMI patients undergoing primary percutaneous coronary intervention. *J Clin Med* 2020;**9**:1051.
177. Bostjancic E, Zidar N, Glavac D. MicroRNAs and cardiac sarcoplasmic reticulum calcium ATPase-2 in human myocardial infarction: expression and bioinformatic analysis. *BMC Genomics* 2012;**13**:552.
178. Vila-Navarro E, Duran-Sanchon S, Vila-Casadesús M, Moreira L, Ginès À, Cuatrecasas M, Lozano JJ, Bujanda L, Castells A, Gironella ML. Novel circulating miRNA signatures for early detection of pancreatic neoplasia. *Clin Transl Gastroenterol* 2019;**10**:e00029.
179. Sole C, Tramonti D, Schramm M, Goicoechea I, Armesto M, Hernandez LI, Manterola L, Fernandez-Mercado M, Mujika K, Tuneu A, Jaka A, Tellaetxe M, Friedlander MR, Estivill X, Piazza P, Ortiz-Romero PL, Middleton MR, Lawrie CH. The circulating transcriptome as a source of biomarkers for melanoma. *Cancers* 2019;**11**:70.
180. Zhao W, Sun Q, Yu Z, Mao S, Jin Y, Li J, Jiang Z, Zhang Y, Chen M, Chen P, Chen D, Xu H, Ding S, Yu Z. MiR-320a-3p/ELF3 axis regulates cell metastasis and invasion in non-small cell lung cancer via PI3K/Akt pathway. *Gene* 2018;**670**:31–37.
181. Ge X, Cui H, Zhou Y, Yin D, Feng Y, Xin Q, Xu X, Liu W, Liu S, Zhang Q. MiR-320a modulates cell growth and chemosensitivity via regulating ADAM10 in gastric cancer. *Mol Med Rep* 2017;**16**:9664–9670.
182. Jeong HS, Kim JY, Lee SH, Hwang J, Shin JW, Song KS, Lee S, Kim J. Synergy of circulating miR-212 with markers for cardiovascular risks to enhance estimation of atherosclerosis presence. *PLoS One* 2017;**12**:e0177809.
183. Masson S, Batkai S, Beermann J, Bar C, Pfanne A, Thum S, Magnoli M, Balconi G, Nicolosi GL, Tavazzi L, Latini R, Thum T. Circulating microRNA-132 levels improve risk prediction for heart failure hospitalization in patients with chronic heart failure. *Eur J Heart Fail* 2018;**20**:78–85.
184. Ucar A, Gupta SK, Fiedler J, Eriki E, Kardasinski M, Batkai S, Dangwal S, Kumarswamy R, Bang C, Holzmann A, Remke J, Caprio M, Jentsch C, Engelhardt S, Geisendorf S, Glas C, Hofmann TG, Nessling M, Richter K, Schiffer M, Carrier L, Napp LC, Bauersachs J, Chowdhury K, Thum T. The miRNA-212/132 family regulates both cardiac hypertrophy and cardiomyocyte autophagy. *Nat Commun* 2012;**3**:1078.
185. Duell EJ, Lujan-Barroso L, Sala N, Deitz McElyea S, Overvad K, Tjonneland A, Olsen A, Weiderpass E, Busund LT, Moi L, Muller D, Vineis P, Aune D, Matullo G, Naccarati A, Panico S, Tagliabue G, Tumino R, Palli D, Kaaks R, Katzke VA, Boeing H, Bueno-de-Mesquita HBA, Peeters PH, Trichopoulou A, Lagiou P, Kotanidou A, Travis RC, Wareham N, Khaw KT, Ramon Quirós J, Rodríguez-Barranco M, Dorronsoro M, Chirlaque MD, Ardanaz E, Severi G, Boutron-Ruault MC, Rebours V, Brennan P, Gunter M, Scelo G, Cote G, Sherman S, Koric M. Plasma microRNAs as biomarkers of pancreatic cancer risk in a prospective cohort study. *Int J Cancer* 2017;**141**:905–915.
186. Weber DG, Gawrych K, Casjens S, Brik A, Lehnert M, Taeger D, Pesch B, Kollmeier J, Bauer TT, Johnen G, Bruning T. Circulating miR-132-3p as a candidate diagnostic biomarker for malignant mesothelioma. *Dis Markers* 2017;**2017**:1–15.
187. Jiang X, Chen X, Chen L, Ma Y, Zhou L, Qi Q, Liu Y, Zhang S, Luo J, Zhou X. Upregulation of the miR-212/132 cluster suppresses proliferation of human lung cancer cells. *Oncol Rep* 2015;**33**:705–712.
188. Lin L, Wang Z, Jin H, Shi H, Lu Z, Qi Z. MiR-212/132 is epigenetically downregulated by SOX4/EZH2-H3K27me3 feedback loop in ovarian cancer cells. *Tumour Biol* 2016;**37**:15719–15727.
189. Oliosio D, Dauriz M, Bacchi E, Negri C, Santi L, Bonora E, Moghetti P. Effects of aerobic and resistance training on circulating micro-RNA expression profile in subjects with type 2 diabetes. *J Clin Endocrinol Metab* 2019;**104**:1119–1130.
190. Li J, Wan W, Chen T, Tong S, Jiang X, Liu W. MiR-451 silencing inhibited doxorubicin exposure-induced cardiotoxicity in mice. *Biomed Res Int* 2019;**2019**:1–11.
191. Song L, Su M, Wang S, Zou Y, Wang X, Wang Y, Cui H, Zhao P, Hui R, Wang J. MiR-451 is decreased in hypertrophic cardiomyopathy and regulates autophagy by targeting TSC1. *J Cell Mol Med* 2014;**18**:2266–2274.
192. Bai H, Wu S. MiR-451: a novel biomarker and potential therapeutic target for cancer. *Oncol Targets Ther* 2019;**12**:11069–11082.
193. Shen Y, Ding Y, Ma Q, Zhao L, Guo X, Shao Y, Niu C, He Y, Zhang F, Zheng D, Wei W, Liu F. Identification of novel circulating miRNA biomarkers for the diagnosis of esophageal squamous cell carcinoma and squamous dysplasia. *Cancer Epidemiol Biomarkers Prev* 2019;**28**:1212–1220.
194. Wang Q, Shang J, Zhang Y, Zhou Y, Tang L. MiR-451a restrains the growth and metastatic phenotypes of papillary thyroid carcinoma cells via inhibiting ZEB1. *Biomed Pharmacother* 2020;**127**:109901.
195. Dong Y, Wang G. Knockdown of lncRNA SNHG12 suppresses cell proliferation, migration and invasion in breast cancer by sponging miR-451a. *Int J Clin Exp Pathol* 2020;**13**:393–402.
196. Kaur A, Mackin ST, Schlosser K, Wong FL, Elharham M, Delles C, Stewart DJ, Dayan N, Landry T, Pilote L. Systematic review of microRNA biomarkers in acute coronary syndrome and stable coronary artery disease. *Cardiovasc Res* 2020;**116**:1113–1124.
197. Pereira-da-Silva T, Coutinho Cruz M, Carrusca C, Cruz Ferreira R, Napoleao P, Carmo MM. Circulating microRNA profiles in different arterial territories of stable atherosclerotic disease: a systematic review. *Am J Cardiovasc Dis* 2018;**8**:1–13.
198. Lai L, Chen J, Wang N, Zhu G, Duan X, Ling F. MiRNA-30e mediated cardioprotection of ACE2 in rats with doxorubicin-induced heart failure through inhibiting cardiomyocytes autophagy. *Life Sci* 2017;**169**:69–75.
199. Zhang X, Dong S, Jia Q, Zhang A, Li Y, Zhu Y, Lv S, Zhang J. The microRNA in ventricular remodeling: the miR-30 family. *Biosci Rep* 2019;**39**:BSR20190788. doi: 10.1042/BSR20190788.
200. Shi M, Mu Y, Zhang H, Liu M, Wan J, Qin X, Li C. MicroRNA-200 and microRNA-30 family as prognostic molecular signatures in ovarian cancer: a meta-analysis. *Medicine* 2018;**97**:e11505.
201. Wang C, Chen L, Yang Y, Zhang M, Wong G. Identification of bladder cancer prognostic biomarkers using an ageing gene-related competitive endogenous RNA network. *Oncotarget* 2017;**8**:111742–111753.
202. Han W, Cui H, Liang J, Su X. Role of microRNA-30c in cancer progression. *J Cancer* 2020;**11**:2593–2601.
203. Croset M, Pantano F, Kan CWS, Bonnellye E, Descotes F, Alix-Panabières C, Lecellier C-H, Bachelier R, Alloli N, Hong S-S, Bartkowiak K, Pantel K, Clézardin P. MiRNA-30 family members inhibit breast cancer invasion, osteomimicry, and bone destruction by directly targeting multiple bone metastasis-associated genes. *Cancer Res* 2018;**78**:5259–5273.