

# Electron paramagnetic resonance: a powerful tool to support magnetic resonance imaging research

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The purpose of this paper is to describe some of the areas where electron paramagnetic resonance (EPR) has provided unique information to MRI developments. The field of application mainly encompasses the EPR characterization of MRI paramagnetic contrast agents (gadolinium and manganese chelates, nitroxides) and superparamagnetic agents (iron oxide particles). The combined use of MRI and EPR has also been used to qualify or disqualify sources of contrast in MRI. Illustrative examples are presented with attempts to qualify oxygen sensitive contrast (i.e.  $T_1$ - and  $T_2^*$ -based methods), redox status or melanin content in tissues. Other areas are likely to benefit from the combined EPR/MRI approach, namely cell tracking studies. Finally, the combination of EPR and MRI studies on the same models provides invaluable data regarding tissue oxygenation, hemodynamics and energetics. Our description will be illustrative rather than exhaustive to give to the readers a flavour of 'what EPR can do for MRI'. Copyright © 2014 John Wiley & Sons, Ltd.

**Keywords:** EPR; MRI; contrast agent; nitroxide; oxygen; iron oxide

## 1. INTRODUCTION: NMR VERSUS EPR

Historically, electron paramagnetic resonance (EPR) [or equivalently electron spin resonance (ESR)] and nuclear magnetic resonance (NMR) were discovered at the same period of time. In 1944, Zavoisky first described the EPR phenomenon (1,2). A little later, Bloch and Purcell discovered the NMR phenomenon (3,4). In 1973, Lauterbur published the original idea of applying magnetic field gradients in three dimensions to create NMR images (5). Since that time, MRI has been extensively developed and has revolutionized medical imaging. In other respects, the first *in vivo* EPR experiments were carried out in 1975 (6,7) and the first EPR imaging (EPRI) experiment was reported in 1979 (8,9). In comparison with the flourishing applications of MRI for the diagnosis and treatment of human diseases, the development of *in vivo* EPR and EPR imaging has been rather slow. There are several reasons for this, especially linked to the considerable technical challenges that make *in vivo* EPR difficult (7,10).

There are few fundamental differences between the principles of electron and nuclear magnetic resonance as NMR- and EPR-related techniques share similar basic principles. However, the differences in physical and chemical properties of the resonant species (unpaired electrons vs nuclei with net spin) lead to profound differences in the techniques that are used to record the spectra or to reconstruct an image (7,10–12) (Table 1). There are three major differences between EPR and NMR: (a) the size of the magnetic moments, which fixes the ratio between the frequency and the magnetic field; (b) the very short relaxations times of the electron spins compared with the nuclear spins; and (c) the limited amounts of paramagnetic materials that are present in most biological systems.

The gyromagnetic ratio of an unpaired electron is much larger than that of a proton, so the resonance frequency/magnetic field ratio for the electron is 28 GHz/T vs 42.5 MHz/T for the proton. As

a consequence, standard EPR spectrometers operate at much higher frequencies (microwaves) and lower fields than NMR spectrometers: the most usual commercial EPR spectrometers operate at 9 GHz (X-band) for a magnetic field around 0.34 T. At this high frequency, the nonresonant absorption of the electromagnetic radiation by the liquid water presents serious problems, and limits the sample size to 1 mm. Classical *in vitro* EPR experiments on cells are carried out in thin capillary tubes, and *in vivo* experiments at 9 GHz are limited to pharmacokinetic studies on the tail vein of mice (13,14). Larger aqueous samples or animals can be studied only by reducing the operating frequency, for example by using 1 GHz spectrometers (L-band) (15–17). At 1 GHz, the depth of penetration is of about 1 cm, but with a reduced sensitivity.

A second major difference between NMR and EPR is related to the relaxation times. While longitudinal  $T_1$  and transversal  $T_2$  relaxation times of protons are in the range of 0.1–1 s (18), electronic spins return to the equilibrium after excitation much more rapidly (ns to  $\mu$ s). This feature has two consequences. First, EPR spectra in biological systems are mostly obtained through continuous wave experiments. Instead of applying an energy pulse and recording the transient response as done in NMR and MRI, the sample is continuously irradiated with a low-power microwave radiation while the magnetic field is gradually increased so that electron spins come into resonance. So far, only a few groups have succeeded in addressing the challenges of time-

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## Biographies

**Pierre Danhier** is born in 1986 in Brussels in Belgium. He graduated with a PhD in Pharmaceutical Sciences from the Université catholique de Louvain (Belgium) in 2013. The PhD was completed in the biomedical resonance unit group from Professor Bernard Gallez. Research led during the PhD focused on tracking *in vivo* metastatic cancer cells using magnetic resonance approaches. He is currently occupying a postdoc position under the supervision of Professor Zaver Bhujwalla at the Johns Hopkins University (Baltimore, USA), studying the link between tumor hypoxia and metastasis using optical imaging.



**Bernard Gallez** is born in 1964 in Mons (Belgium). He is pharmacist (1988), industry pharmacist and radiopharmacist (1990), and got his PhD in 1993 with a work focused on the targeting of MRI contrast agents. He made a post-doc in Dartmouth where he learned *in vivo* EPR. In Brussels, he built the Biomedical Magnetic Resonance laboratory with a focus on the development of new MR technologies (MRI and EPR) and their applications to the characterization and manipulation of the tumor microenvironment and the hemodynamic process in tissues (oxygenation, perfusion, oxygen consumption, angiogenesis).



**Table 1.** Comparison between NMR and EPR

|                                | EPR                                                                                                                                                                                                     | NMR                                                                                                                                                                              |
|--------------------------------|---------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| Detectable species             | Compounds with unpaired electron(s)                                                                                                                                                                     | Compounds with nuclear spin $I \neq 0$                                                                                                                                           |
| Frequency/magnetic field ratio | 28 GHz/T                                                                                                                                                                                                | 42.5 MHz/T                                                                                                                                                                       |
| Relaxation time                | ns– $\mu$ s                                                                                                                                                                                             | ms–s                                                                                                                                                                             |
| Need for exogenous probes      | <ul style="list-style-type: none"> <li>No for stable paramagnetic species (Mn, iron oxides, etc.)</li> <li>Yes for labile radicals (spin trapping)</li> <li>Yes for oximetry (spin labeling)</li> </ul> | No ( $H_2O$ concentration in living organisms $>50$ M)                                                                                                                           |
| Sensitivity                    | <ul style="list-style-type: none"> <li>High <i>in vitro</i> or <i>ex vivo</i> (<math>&lt;nm</math>)</li> <li>Limited <i>in vivo</i> owing to RF absorption by water</li> </ul>                          | <ul style="list-style-type: none"> <li>Low (<math>\sim 10^{-4}</math> M)</li> <li>Improved thanks to contrast agents (Gd, iron oxides, etc.) for imaging applications</li> </ul> |
| Clinical applications          | Limited so far: oximetry (spectroscopy) and retrospective dosimetry                                                                                                                                     | Wide range of imaging applications owing to: <ul style="list-style-type: none"> <li>a high anatomical and temporal resolution</li> <li>variety of source of contrasts</li> </ul> |

domain EPR *in vivo* (19–21). The second consequence of short EPR relaxation times is that EPR imaging requires the application of magnetic gradients that are much larger than those used in MRI (11).

The third but not the least difference between both MR techniques relies on the nature of the material that is studied. Compared with the 100 M concentration of endogenous protons available for NMR, there is a lack of sufficient amounts of naturally occurring paramagnetic materials for EPR. Except for melanin that can be easily detected in biological samples (22–26), endogenous paramagnetic species are present in insufficient amounts to be detected by *in vivo* EPR. The short half-life of most free radicals involved in physiology (such as nitric oxide) or in toxicology (such as superoxide or hydroxyl radicals) is another source of difficulty. These reactive species are usually detected using a spin-trapping strategy by the formation of a stable spin adduct after reaction with a nitron (27). Generally, for *in vivo* and *in vitro* EPR biological studies, a stable paramagnetic material has to be introduced in the biological system to be detected. Very interestingly, the paramagnetic reporter may provide

unique information on its environment such as *in vitro* and *in vivo* oxygenation (28–32), redox status (33,34), pH (35–38) or microviscosity (39,40).

For those not familiar with EPR spectroscopy, it should be acknowledged that the main parameters derived from EPR spectra are the *g-values* (analogous to the chemical shift in NMR), the electron–nuclear hyperfine coupling constants (analogous to nuclear spin–spin coupling constants in NMR), linewidths and intensities (41). Because EPR spectra are usually displayed as first derivatives, linewidths are conveniently measured as the distance between points of maximum and minimum slope. Intensities are often measured as the amplitude of each line, but it is preferable to use the double integration of the signal to assess the concentration in a paramagnetic species.

While the development of *in vivo* EPR is a research field *per se*, there is an outstanding potential of EPR-related methods to support MRI developments. The purpose of this manuscript is to describe some of the areas where EPR has provided unique information to MRI developments. The field of application mainly encompasses the EPR characterization of MRI paramagnetic

contrast agents (gadolinium and manganese chelates, nitroxides) and superparamagnetic agents (iron oxide particles). The combined use of MRI and EPR has also been used to qualify or disqualify sources of contrast in MRI. Illustrative examples are presented with attempts to qualify oxygen sensitive contrast (i.e.  $T_1$ - and  $T_2^*$ -based methods), redox status or melanin content in tissues. Other areas are likely to benefit from the combined EPR/MRI approach, namely cell tracking studies. Finally, the combination of EPR and MRI studies on the same models provide invaluable data regarding tissue oxygenation, hemodynamics and energetics. Our description will be illustrative rather than exhaustive to give to the readers a flavour of 'what EPR can do for MRI'. A comprehensive review on this research field was published about 10 years ago (42), so an update is timely.

## 2. CHARACTERIZATION OF MRI CONTRAST AGENTS PROPERTIES WITH EPR

Despite its richness in sources of contrast and its high spatial resolution, MRI lacks sensitivity. It is generally admitted that the use of contrast agents (CAs) may counterbalance this default. Seeking innovative tissue-selective and smart CAs is a major research area for many academic groups and pharmaceutical companies. Virtually all MRI contrast agents contain species with unpaired electrons so that EPR is a method of choice for direct study of their properties. The interaction between the electron magnetic moment and the nuclear spin moment of the water proton will induce an increase in the proton relaxation rates. The relaxivity defines the ability of a CA to affect the relaxation rate of the surrounding protons. The effect of a CA on the MR image will mainly depend on the  $r_2/r_1$  ratio, where  $r_1$  is the longitudinal relaxivity and  $r_2$  is the transversal relaxivity ( $s^{-1} \text{ mM}^{-1}$ ). On the one hand, paramagnetic CAs such as lanthanide derivatives exhibit a low  $r_2/r_1$  ratio (typically, between 1 and 2) and will enhance the signal on  $T_1$ -weighted images when present at low concentration. Such contrast agents are thus called positive CAs. On the other hand, superparamagnetic and ferromagnetic CAs such as iron oxide particles exhibit a high  $r_2/r_1$  ratio (as high as 50) and will decrease the signal on  $T_2$ -weighted images. Such contrast agents are called negative CAs. As described hereafter, EPR may provide invaluable information on both classes (paramagnetic and superparamagnetic) of CAs.

### 2.1. Exogenous Paramagnetic Contrast Agents

#### 2.1.1. Gadolinium

Gadolinium complexes represent the major class of positive CAs used in the clinic. The use of chelating agents is required to prevent toxicity of gadolinium and ensure an effective elimination of the metal. The first generation of gadolinium-based CAs encompasses highly hydrophilic low-molecular-weight complexes with an extracellular biodistribution (based on the DTPA or the DOTA structure). Later on were developed compounds bearing more lipophilic chains in order to modulate the biodistribution with a partial biliary elimination (Gd-EOB-DTPA or gadoxetate, and Gd-BOPTA or gadobenate) and long blood circulation through interactions with albumin (MS-325 or gadofosveset). When developing new Gd-based CAs, it is mandatory to fully characterize the kinetic and thermodynamic stability of the complex (43), the pharmacokinetic profile as well as the relaxivity of the new agent. The relaxivity of a CA can be described by taking

into account the 'inner sphere' and the 'outer sphere' contributions (44). Briefly, the inner sphere contribution relies on the exchange of water molecules remaining in the primary coordination site and the bulk solvent whereas outer sphere relaxation mechanisms involve distant interactions between paramagnetic gadolinium and bulk water molecules. The inner sphere contribution is the most effective and governed by four correlation times: the rotational correlation time of the complex ( $\tau_R$ ), the residence time of a water proton in the inner coordination sphere ( $\tau_m$ ) and its chemical exchange with bulk water, and the electronic longitudinal and transverse relaxation rates ( $1/T_{1e}$  and  $1/T_{2e}$ ) of the metal center. The characterization of the contrast agents is generally obtained thanks to the measurements of relaxivities at different field strengths (NMRD profiles). To elucidate the different contributions to the observed relaxivity of soluble Gd-based CAs, NMRD profiles can advantageously be supplemented by a combination of  $^{17}\text{O}$  studies that measure water exchange rates and EPR spectrometry studies that provide precise measurements of the electronic spin relaxation times (45–51). This simultaneous analysis of experimental data helps to solve the problem of the interdependence of parameters in NMRD analysis and yields to physically meaningful values (52,53). Although less attention has been devoted to the influence of electronic relaxation on the relaxivity, it was worthwhile to investigate how different ligand designs could optimize the electron spin relaxation properties. The overall transverse electronic relaxation rates,  $1/T_{2e}$ , can be obtained from the peak-to-peak EPR line widths of the derivative spectra, and the electron spin relaxation rates for Gd(III) complexes are usually interpreted in terms of a transient zero-field splitting (ZFS) mechanism induced by distortions of the complex (45). A series of more in-depth studies analyzed the contribution of the static ZFS and transient ZFS to the electronic relaxation (53–56). Indeed, the structure and the dynamics of the electronic density of the chelate framework that surrounds the metal determine the ZFS and therefore the electron spin relaxation rates in solution. This was achieved by recording EPR spectra at different temperatures and frequencies on different gadolinium complexes. Overall, these studies performed on many different Gd chelates demonstrate that, for fast rotating molecules, electronic relaxation has little influence on the efficiency of Gd(III)-based MRI contrast agents at high magnetic fields ( $B > 3 \text{ T}$ ), but may have a marked influence at low magnetic field. In other words, the relatively long electronic relaxation time of Gd-based contrast agents (compared with other metals) is not the limiting factor in the relaxivity, at least at the field strengths presently used by the MRI clinical scanners.

The EPR studies were not limited to the study of soluble Gd-based CAs and were also applied for the characterization of nanoparticles. In a study on Gd(III)-doped zeolite NaY nanoparticles (57), EPR spectra suggest that an interaction between neighboring  $\text{Gd}^{3+}$  ions within the same particle produces a significant increase in the transversal electronic relaxation rates in samples with a high  $\text{Gd}^{3+}$  content. A recent study was carried out on gadolinium oxide or gadolinium hydroxide nanoparticles as well as on dysprosium, holmium, terbium and erbium containing particles (58). For these latter compounds, the combination of NMR and EPR studies with theoretical simulations showed that  $T_2$  relaxation is caused by water diffusion in the field inhomogeneities created by the magnetic particle. The different relaxation behaviors were caused by different electron

relaxation times, as estimated by EPR (58). Other EPR studies focused on the interactions of lipophilic Gd-based CAs with biological membranes (59). It was shown that the spin label EPR method using nitroxides can be used to detect the presence and possible location of lipophilic CAs in the model membranes, the changes and distortions in membrane organization, and the changes in the local polarity of the bilayer due to addition of  $\text{Gd}^{3+}$  complexes (59). This work also demonstrated that interaction of  $\text{Gd}^{3+}$  complexes with phospholipid bilayers can be observed directly from changes in their EPR spectra. In addition, the analysis of frequency dependence of the  $g$ -factors provided estimates of the ZFS parameter in physiological conditions and information on how this parameter was affected by interaction with lipids. In a recent study, Lagerstedt *et al.* used a combination of EPR and relaxivity studies to characterize Gd-labeled apoA-lipoprotein A-I as a MRI contrast agent (60). This protein contrast agent was obtained by attaching the thiol-reactive Gd label at Cys residues replaced at four distinct positions in apoA-I. The authors found out that that putative site-directed protein contrast agents can be screened by X-band EPR spectroscopy employing both nitroxide and Gd(III)-DOTA probes. According to the authors (60), the main advantage of using EPR spectroscopy is that the concentration of the protein is significantly lower ( $\sim 10 \mu\text{M}$  spin for nitroxide-labeled and  $\sim 1 \text{ mM}$  for Gd(III)-labeled sample) than is necessary in NMR spectroscopy ( $> 100 \mu\text{M}$  protein). Moreover, the sample volume for EPR experiments ( $\sim 5 \mu\text{l}$ ) is about two orders of magnitude smaller than that used for NMR measurements ( $\sim 500 \mu\text{l}$ ).

Eventually, EPR was also used for the gadolinium quantification in tissues (61). Although the quantification of Gd in tissues is undeniably important when trying to target CAs to different receptors, surprisingly, EPR spectroscopy has been applied for this purpose only exceptionally. An example was the application of EPR to quantify Gd-labeled vesicles targeted to overexpressed glucose receptors in tumors (61). The quantification of the Gd signal intensity is a straightforward method and has the potential to be issued for many other applications. More recently, an alternative indirect method has been suggested by Takeshita *et al.* (62). They used a spin labeling approach by using nitroxide solutions. The presence of paramagnetic Gd induces a decrease in the  $T_{2e}$  and consequently an EPR linebroadening. They found a linear relationship between the concentration of Gd-CAs and the linewidth of the nitroxide probe. Possible applications encompass the Gd quantification in plasma (62).

### 2.1.2. Manganese

Manganese is a paramagnetic transition metal that possesses five free electrons.  $\text{Mn}^{2+}$  has a low  $r_2/r_1$  ratio and serves as a positive CA on  $T_1$ -weighted images (63). In experimental studies, an interesting feature of  $\text{Mn}^{2+}$  is its ability to mimic  $\text{Ca}^{2+}$  *in vivo*. As calcium, manganese ions enter in cells via voltage-gated  $\text{Ca}^{2+}$  channels.  $\text{Mn}^{2+}$  can be taken up by neurons and move along axons to reach synapses (64,65). Hence, manganese-enhanced MRI (MEMRI) was increasingly developed to study the functional connectivity patterns in small animals (66,67). MEMRI is mostly performed using infusions of  $\text{MnCl}_2$ . Unfortunately, as  $\text{Mn}^{2+}$  is neurotoxic (68,69) through the accumulation in the striatum (70,71), it is unlikely that MEMRI applications will eventually be possible in humans. The use of chelates to prevent the toxicity of  $\text{Mn}^{2+}$  was investigated and one compound, Mn-DPDP (Mn-dipyridoxaldiphosphate or mangafodipir), was used in

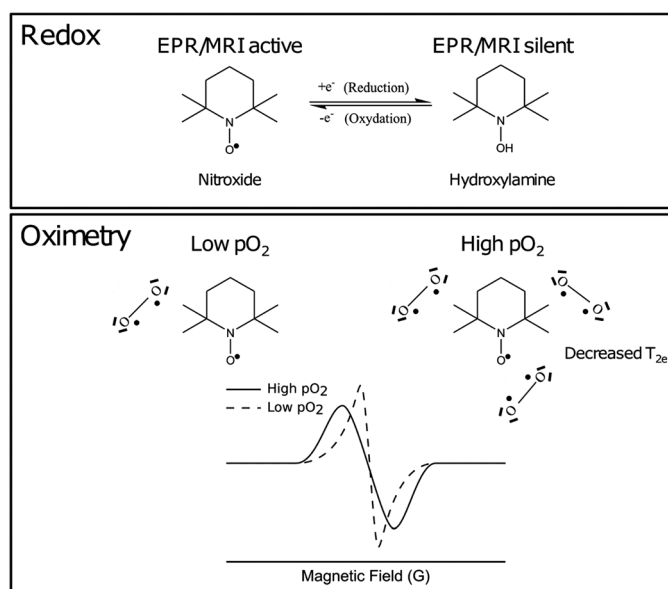
humans for hepatobiliary imaging (72). Rapidly, several reports suggested an *in vivo* dissociation of the Mn-chelate. These assumptions were actually based on the fact that the signal enhancement was persistent after injection in organs where free manganese is known to accumulate (44,73). EPR played a key role in the direct evidence for the dissociation (73). At 9 GHz, free  $\text{Mn}^{2+}$  provides a six-line-EPR spectrum whereas bound manganese is EPR silent (73,74). These features allowed the study of the dissociation of Mn-DPDP in biological tissues (73). Further evidence for this dissociation was provided by radiolabeling studies using different isotopes for the ligand and for the metal (75). Interestingly, these EPR and isotope studies were in agreement with the changes in  $^{31}\text{P}$ -NMR spectra obtained in tissues (73,76). Further EPR studies at different frequencies confirmed the dissociation and suggested the role of  $\text{Zn}^{2+}$  in the transmetallation of Mn-DPDP (74). These EPR data were important to question the relaxation properties of Mn-DPDP. Indeed, it was found that the  $\text{Mn}^{2+}$  released by the complex was binding to proteins, this association being responsible for the large increase in the  $T_1$  relaxivity (73,74). As a consequence of this set of evidence for the dissociation of the Mn complex, it was found that the transport into the hepatic cells was not mediated by the pyridoxine transporter (76,77). It also raised concerns about the possible accumulation of free manganese in the brain and possible associated toxicity (78–80). Although mangafodipir is no longer commercialized, research on Mn-based chelates is still active, and it is likely that EPR may contribute to the characterization of these potential CAs.

### 2.1.3. Nitroxides

Nitroxide compounds are stable organic molecules that possess one unpaired electron (Fig. 1). As gadolinium manganese-based compounds, nitroxides have been proposed as positive CAs for MRI (81–83). The development of nitroxides as MRI CAs was rather challenging because their relaxivity is unfavorable compared with metal-based agents. Contrary to Gd-based compounds, the main contribution is due to a distant interaction between the free electron and the water, and the contribution of the inner-sphere process for hydrophilic nitroxide is considered to be negligible. There were several attempts to increase the relaxivity of nitroxides, namely by using the immobilization of nitroxides directly linked to macromolecules (84–88) or strongly interacting with proteins such as albumin because of their large lipophilicity (89–93). EPR studies demonstrate that the degree of immobilization as evidenced by EPR spectrometry was directly correlated to an increased NMR  $r_1$  relaxivity (86). In addition, it was shown that the number of lipophilic nitroxides adsorbed on albumin (as quantified by EPR spectroscopy) was directly responsible for the increased relaxivity (91).

One challenge that appeared when using nitroxides as MRI CAs is the rapid elimination of the compounds by a metabolic reaction. The paramagnetic nitroxide is rapidly reduced and converted into the corresponding diamagnetic hydroxylamine (Fig. 1), an effect that leads to the fading and disappearance of the contrast induced by the nitroxide on the MR image. Systematic EPR and/or MRI studies were carried out to establish the structure/reactivity relationships of the nitroxides (94–99). It also appeared that this bioreduction was dependent on the oxygenation and the redox status of the tissues (94,100–102). The group of H. M. Swartz understood the value of turning the problem of nitroxide metabolism into virtues, and developed the concept





**Figure 1.** The versatile role of nitroxides (=nitroxyl radicals) for redox and oximetry studies. As shown in the upper panel, paramagnetic nitroxides can be reduced in biological media into electron paramagnetic resonance (EPR) silent hydroxylamines. The rate of decrease in the EPR signal intensity reflects thus the redox state of the surrounding environment. For oximetry studies (lower panel), electronic transversal relaxation times ( $T_{2e}$ ) of nitroxides are affected by the presence of surrounding oxygen molecules. Therefore the linewidth of the EPR spectrum of a nitroxide is proportional to the  $pO_2$  of the sample.

of functional contrast agents that are sensitive to the oxygenation and to the redox status. Historically, this is probably one of the first example of MRI 'smart contrast agents' that are activated or deactivated in the presence of a key factor present inside a tissue. The seminal ideas of using these compounds as redox-sensitive contrast agents are still being actively developed in EPR imaging (103–108) and in MRI (13,34,109–113). In these strategies to qualify these redox-sensitive CAs, we may say that EPR and MRI are complementary because EPR is monitoring directly the concentration of the nitroxide and indirect measurements with MRI provide images with a higher resolution. Moreover, MRI systems are more widespread and accessible to a larger imaging community. However, special care should be taken when linking the signal decay of the nitroxides to the redox status because the kinetics of clearance may also depend on the perfusion (and wash-out from the tissue) and on the viability of the cells (13). Again, direct measurements by EPR may resolve the origin of the decrease in nitroxide concentration (13).

Another possible interesting application is the ability to target nitroxides to specific tissues or receptors. Actually, the nitroxide moiety can be anchored to any specific organic entity that will be responsible for the biodistribution and the accumulation in a tissue of interest. Historically, small hydrophilic nitroxides were first used as positive MRI CAs (82,83,114,115). Later on, larger molecules bearing nitroxides were tested. In addition to the increase in relaxivity linked to the immobilization of the nitroxides, longer blood half-life and slower kidney elimination were obtained (87,116). The first example of targeting of nitroxides was the development of liver-specific CAs. This was achieved with the synthesis and evaluation of nitroxide fatty acid derivatives recognizing the Fatty Acid Binding Protein responsible for the internalization of these derivatives in the hepatocytes (90,91). Another target present at the surface of the hepatocytes was the asialoglycoprotein receptor. Polysaccharides bearing galactose residues, such as arabinogalactans, were functionalized

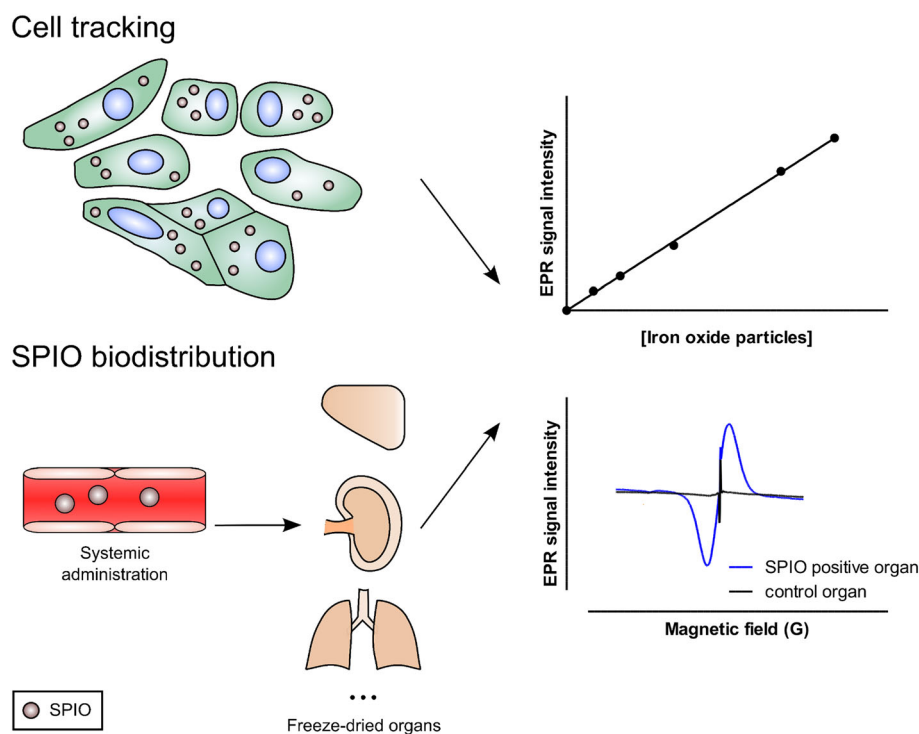
with nitroxides (85). In each case, EPR, relaxometry and *in vivo* MRI studies were used in combination to characterize these MRI CAs. Other compounds were developed in order to gain in liver selectivity thanks to an accumulation in another compartment of the liver, the Kupffer cells, for example using liposomes including lipophilic compounds (117). More recently, lipophilic compounds able to cross the blood–brain barrier were developed and evaluated as EPRI and MRI CAs (108,110,118–123). Finally, in a limited series of studies, EPR spectroscopy was also used to understand how pathological situations affecting liver function and kidney elimination was affecting the kinetics of elimination of nitroxide CAs (124).

## 2.2. Exogenous Superparamagnetic Contrast Agents

Superparamagnetic iron oxide particles (SPIO) CAs exhibit a high  $r_2/r_1$  ratio and induce a signal loss on  $T_2$ - and  $T_2^*$ -weighted images. The SPIOs may vary in size and in the coating surrounding the magnetic iron oxide core, properties that affect their stability and physico-chemical properties. The readers are invited to consult a comprehensive review of Laurent *et al.* to learn more on the chemical synthesis of these particles, the relationships between their physico-chemical design and their magnetic properties and how the constituents of the coating modulate the biological properties (125). The first generation of SPIOs that were developed accumulated preferentially in cells belonging to the reticulo-endothelial system (i.e. Kupffer cells in liver) and was mainly used as liver CAs. Hence, clinically approved ferumoxides were designed for improving the detection of liver tumors (126–128). Ultrasmall iron oxide particles (USPIOs) with a diameter lower than 20 nm are less taken up by the liver and have a longer blood half-life. They may be used as blood pool agents (129,130) or lymphotropic agents (131–133). In the recent years, intense research has been devoted to attempts to functionalize the USPIOs with appropriate targeting molecules

in order to confer on them a high selectivity to tissues or molecular targets overexpressed in specific diseases (125,134–138). The modulation of the biodistribution of iron particles is theoretically more attractive than gadolinium. Indeed, iron oxides can be detected at lower concentration than gadolinium thanks to their higher relaxivities. Moreover, the degradation of the iron core over the time and the physiological iron recycling will preclude the accumulation of a toxic metal contrarily to gadolinium (139,140), although each functionalized iron oxide platform should be carefully checked for potential toxicity (141–145). Because iron oxide particles induce a decrease in the MR signal intensity on  $T_2$ - and  $T_2^*$ -weighted images, the qualification of new CAs should carefully address the origin of the recorded induced hyposignal. Indeed, several physiological events or artifacts linked to the presence of air cavities, blood vessels, bone–tissue interfaces and hemorrhages may also be responsible of signal voids and may be confounding factors on  $T_2$ - and  $T_2^*$ -weighted images (146,147). Generally, the accumulation of the iron oxides in the tissue of interest is confirmed by histochemistry using Prussian Blue staining (148), but this technique is qualitative and not quantitative. The assessment of the concentration of iron oxides particles in tissues of interest represents an important step in the qualification of new CAs (149). To achieve that goal, radioactive  $^{59}\text{Fe}$ -SPIO can be used, but this requires the chemical synthesis of radiolabeled nanoparticles (139) which is cumbersome. Inductively coupled plasma mass spectrometry (ICP-MS) or inductively coupled plasma optical emission spectrometry (ICP-OES) are alternatives that provide a sensitive quantification of elemental iron in tissues (150–152). However, it should be noted that this method is unable to discriminate endogenous iron from exogenous iron oxide nanoparticles. In this respect, EPR spectroscopy is invaluable because the EPR spectrum exhibits different lines for the different chemical forms of

iron. The iron oxide contained in SPIOs exhibit a broad and intense EPR spectrum whereas that can be easily differentiated from the free  $\text{Fe}^{3+}$  ion (153,154) (Fig. 2). A comparison study demonstrated that *ex vivo* EPR spectroscopy was superior to ICP-OES for assessing the SPIO biodistribution after administration (155). Historically, the first application of EPR for quantification of iron oxides in tissues was done on the liver of mice that received SPIOs administration in a model of septic shock (156). As new (U)SPIO particles are currently extensively designed for molecular imaging, theranostic, that is, combining diagnosis and treatment using the same platform, *ex vivo* EPR spectroscopy appears to be a reliable tool to monitor iron oxide nanoparticles after injection. Recent applications in this area demonstrated the usefulness of EPR to determine the concentration of USPIOs targeted to E-selectin in the context of the MRI detection of inflammation (154). By combining EPR and MRI, it was demonstrated that these USPIOs covered with a hexapeptide could be a useful molecular MRI marker for the noninvasive diagnosis and monitoring of inflammation (154). Interestingly, it was even possible to use low-frequency EPR to monitor directly *in vivo* the accumulation of these USPIOs in inflamed tissues. These EPR and MRI data supported the interest for the active targeting by grafting the particles with these peptides compared with ungrafted particles. More recent studies used a similar approach to qualify USPIOs targeting dying cells (157,158). USPIOs covered with hexapeptides targeting the phosphatidylserine were shown to induce a signal loss in apoptotic areas within tumors on  $T_2$ -w images. The same particles were shown to constitute an earlier MR biomarker of tumor cell death (after cytotoxic treatments) compared with diffusion-weighted-MRI and choline measurement by  $^1\text{H}$  MRS. EPR spectroscopy was used in these studies to validate the MR data (157,158). It is likely that this application of EPR may be extended to many iron oxides particles platforms in the search



**Figure 2.** EPR as a sensitive tool to assess the iron oxide content in cells and tissues. Iron oxides (e.g. superparamagnetic iron oxide particles, SPIO) exhibit a typical EPR spectrum whereas endogenous iron in biological samples does not. There is a linear relationship between the iron oxide concentration and the EPR signal intensity. Iron oxides concentrations can be measured using EPR in isolated cells or freeze-dried organs.

of new CAs. Although EPR has mostly been used for iron oxide quantification, EPR was also used in one study in combination with magnetometry and relaxometry to elucidate the mechanism of  $T_2$ -shortening induced by monocrystalline iron oxide particles (159).

### 3. MELANIN AS AN ENDOGENOUS MRI CONTRAST AGENT?

Melanin is a paramagnetic polymeric molecule produced by melanocytes to prevent the toxicity of ultraviolet radiations (160). Eumelanin is also produced by pigmented melanoma, a skin cancer that is responsible for 75% of the deaths linked to skin cancer (161). When the malignant melanoma is small and well localized, removal of the tumor with surgery is often sufficient to cure the patient. If the lesion has spread to distant deep tissues, the 5-year survival drops below 10% (162). This poor outcome for advanced diseases highlights the need for early diagnosis modalities. Visual inspection, dermoscopy and biopsies are the most used methods to stage malignant melanoma (163,164). Owing to the paramagnetic properties of melanin, MRI and EPR imaging may potentially be valuable to visualize human skin melanomas and metastases (22–26,165). Most EPR studies conducted so far have been carried out *ex vivo*, but the first image of this endogenous radical has been achieved at low frequency on mice bearing the B16 melanoma model (22). Melanoma metastases are often found in the brain and have been suggested to have a specific MR signature based on their relaxation times (166). However, this specific contrast pattern is subject to widespread debate and several studies have reported disparate results (167,168). In addition to the melanin content, it should be noticed that the MR contrast of melanoma lesions can also be influenced by the presence of metallic ions and the presence of hemorrhages that are often associated with cancer lesions (169,170). Although a correlation was found between the melanin presence in human biopsies (revealed by histopathology analysis) and  $T_1$  values (166,168), only *in vitro* experiments have shown a correlation between the melanin content and  $T_2$  values (171). As the content in paramagnetic melanin can be accurately quantified by EPR, this method was recently used to try to elucidate the possible role of melanin in the MRI contrast (172). This study compared systematically the content in paramagnetic melanin in the tumors and the  $T_1$ ,  $T_2$  and  $T_2^*$  contrast at high field. This was achieved on different models of melanoma presenting different pigmentation. It was actually found that the melanin alone has a negligible effect on the MRI contrast (172) and that other sources like the binding of melanin to metals are likely to have a key role, hence providing a unique response to this historical debate.

### 4. EPR/MRI APPLICATIONS IN CELL TRACKING

Cell tracking can be defined as the noninvasive monitoring of individual cells after injection *in vivo* using imaging techniques. This approach allows the assessment of the biodistribution of cells and their survival after implantation (173). Cell-based therapies (173) and cancer research (174–176) are the main fields of application of cell tracking. The label can be a reporter gene, for example, the luciferase gene for optical imaging (177), or a contrast agent that is incorporated within the cytoplasm of the cell (147). MRI is a suitable modality to perform cell tracking

owing to its high anatomical resolution. Gadolinium agents can in principle be used for cell tracking, but their low relaxivity compared with iron oxide particles leads to the use of large concentrations of intracellular gadolinium to be detected ( $\sim 10^{-4}$  to  $10^{-5}$  M) (147). Therefore, iron oxide particles such as SPIO or microparticles of iron oxide (MPIO, with a diameter of about 1  $\mu$ m) are largely preferred because these particles induce a strong contrast on  $T_2$ - and  $T_2^*$ -weighted images so that single cell tracking is feasible (176,178–182). Intracellular incorporation of a sufficient amount of CAs is an important preliminary step to perform efficient cell tracking. Quantification of internalized iron oxide particles is thus mandatory to optimize labeling strategies. To reach that goal, the same methods than those described for assessing the content of iron oxides in tissues may be used (149). In this context, EPR was shown to be a sensitive, specific and a reliable method to quantify the iron oxide content in cells (183–186). It is of note that *ex vivo* measurement of iron oxide content after a cell tracking experiment using EPR could also be performed to assess the biodistribution of the labeled cells (Fig. 2) and therefore complement MRI results (187).

Cell death, dilution of the iron oxides particles owing to cell proliferation and uptake of released particles by macrophages are important drawbacks of cell labeling by SPIOs (188,189). Therefore, MRI cell tracking using contrast agents may not be appropriate for rapidly dividing cells (such as cancer cells) or for the long-term monitoring of other labeled cells. In this context, reporter genes, that is, engineered genes allowing specific location and visualization of cells, can be considered as attractive because they are only expressed in living cells and are not subject to the dilution effect. The luciferase gene and the green fluorescent protein are the most common reporter genes in optical imaging. The ferritin, a paramagnetic protein involved in iron storage, has been proposed in several studies as a MRI reporter gene (190–193). This protein carries up to 4500 Fe atoms and increases the  $R_2$  of surrounding protons (129). Unfortunately, the low sensitivity remains an issue for *in vivo* imaging (193). As the ferritin protein displays an EPR spectrum owing to its paramagnetic properties (194), the sensitive EPR spectroscopy could be potentially be used to assess the presence and expression of ferritin genes and complement MRI studies. It has been also suggested that melanin could also serve as a reporter in basic metastasis research. As the tyrosinase protein is the main enzyme involved in the synthesis of the melanin, this enzyme has been proposed as a MR reporter gene (195). An *in vitro* study has shown that breast cancer cells transfected with the tyrosinase gene displayed reduced  $T_1$  values (196). Similar observations were also found in mouse fibroblasts and human embryonic kidney cells (197). As discussed earlier, the *in vivo* MRI contrast depends not only on the melanin content but also on the association of metals to melanin. The natural production of melanin in melanomas constitutes an attractive approach to monitor metastasis, and EPR may be used to determine the melanin content in organs with accumulation of melanoma metastases. For instance, the mouse B16 melanoma model is one of the most studied cell lines in metastasis research. Different cell subtypes have been characterized and possess different metastasis patterns. For instance, B16F10 are highly metastatic to the lungs (198) whereas another variant subline B16-F10-B2 produces brain metastasis (199). As the EPR signal intensity directly correlates with the number of melanoma cells (25), *ex vivo* X-band EPR spectroscopy constitutes a unique tool to monitor melanomas

progression to complement MRI observations (25). This would allow the quantitative assessment of the efficacy of new anti-metastatic treatments.

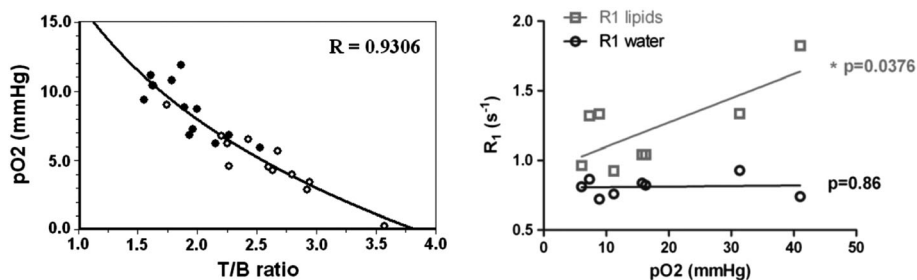
## 5. EPR AS A TOOL TO QUALIFY OR DISQUALIFY OXYGEN-SENSITIVE CONTRAST IN MRI

The oxygen level in tissues is a critical parameter in several pathologies such as stroke (48), peripheral vascular diseases (7), sepsis (49) and cancer (200). For instance, tumor hypoxia has been known for several years to promote aggressiveness, invasion and metastasis (50). In addition, low oxygen tension ( $pO_2 < 5$  mmHg) affects radiotherapy efficacy and restoring an appropriate oxygenation prior to irradiation enhances the tumor radioresponse by a factor 3 (51). Hence, numerous efforts have been accomplished to improve tumor oxygenation using provascular strategies or inhibitors of oxygen consumption (201–206). Direct quantitative measurements of tumor oxygenation can be achieved by using polarographic electrodes and fibre optic probes and phosphorescence quenching based-methods such as the OxyLite system. Unfortunately, these methods are invasive and cannot be repeated on the same tissue over the time. By contrast, EPR oximetry using stable oxygen sensors can be used over long periods of time. The method requires the injection of a small amount paramagnetic probe (few crystals of lithium phthalocyanine or charcoal particles) within the tissue (29,32,207,208) and the  $pO_2$  can be measured noninvasively using low frequency EPR systems. Oxygen, which is paramagnetic, interacts with the paramagnetic centers, inducing a shortening of the  $T_2$  electronic relaxation time and the consequent broadening of the EPR line of the oxygen sensitive probe. More details on EPR oximetry principles and related techniques can be found in several comprehensive reviews (30,32,209). EPR oximetry has been extensively used in oncology to assess the effect of factors or pharmacological agents that modulate the tumor oxygenation (124,201,210–245). Although there are high hopes of translating EPR oximetry into the clinical arena (31,246), the present noninvasive approaches mainly rely on PET/SPECT imaging and MRI (55,56). Hypoxia imaging using PET is obtained through nitroimidazole tracers that preferentially accumulate in hypoxic cells. As this method is indirect, some radiotracers were calibrated with EPR oximetry, which provides absolute measurements of  $pO_2$  (247–249). While these combined PET/EPR studies showed that the accumulation of compounds such as  $^{18}F$ -FMISO,  $^{18}F$ -EF3 and  $^{18}F$ -FAZA in tumors was well correlated to the presence of hypoxic regions with a  $pO_2 < 10$  mmHg, which is radiobiologically relevant (Fig. 3, left), it was found that the accumulation

of  $^{64}Cu$ -ATSM in tumors involved other mechanisms independent of hypoxia (247,249). EPR has also been used to qualify and disqualify sources of MRI contrast that are sensitive to the oxygenation of the tissues. While MRI relaxometry measurements of fluorine compounds are able to provide oxygen maps that actually reflect the tissue oxygenation (250–253), these methods cannot be directly used in patients and most MRI techniques developed so far rely on the exploitation of endogenous contrast. Endogenous sources of contrast include variations of  $T_2^*$  and  $T_1$  values in tissues.  $T_2^*$  is sensitive to the relative deoxyhemoglobin/oxyhemoglobin (Hb/HbO<sub>2</sub>) ratio in vessels (254), while  $T_1$  discloses sensitivity to dissolved paramagnetic oxygen, which acts as a  $T_1$ -shortening paramagnetic contrast agent (255).

$T_2^*$  mapping, also referred to as functional MR imaging or 'BOLD' (blood oxygen level dependent) imaging, is sensitive to variations in oxygenation in the vascular compartment. BOLD imaging has been used in many tissues to monitor changes in oxygenation and perfusion, and has been suggested in oncology as a possible biomarker of tumor hypoxia (256–261). Even if the basal  $T_2^*$  values are unlikely to reflect the actual tumor  $pO_2$  as shown in studies comparing  $T_2^*$  with fiber optic measurements (260), another key question was to establish the capability of BOLD imaging to actually predict changes in tumor oxygenation induced by treatments. Several studies compared the evolution of  $T_2^*$  and of the  $pO_2$  as measured quantitatively by EPR oximetry (202,211,220,262–264). Jordan *et al.* observed that the increase in tumor oxygenation induced by carbogen or by the vasoactive agent, isosorbide dinitrate, correlated well with the evolution of the BOLD signal (211). These results were confirmed in two other tumor models where a positive correlation between  $R_2^*$  values and EPR  $pO_2$  values was observed (262,263). On the contrary, BOLD imaging failed to tackle changes in tumor oxygenation induced by the allosteric hemoglobin modifier RSR13 (220) and by the inhibitors of oxygen consumption, insulin and NS-398 (264). In other words, BOLD seems able to tackle changes in tumor oxygenation if the modulator is increasing at the same time the blood flow and the oxygenation, but cannot predict increase in oxygenation due to other factors like a decrease in oxygen consumption. This limited value of  $R_2^*$  was confirmed in a meta-analysis made on tumor models that compared the respective value of possible predictive markers of response to an association irradiation/co-treatment (202). While EPR oximetry was always predictive of the issue of the anticancer treatment in these models, BOLD-MRI failed to be a predictive biomarker of response to combined treatments (202).

$T_1$ -based techniques are sensitive to dissolved tissue oxygen and changes in tissue oxygen concentrations have been shown to produce changes in relaxation rate  $R_1$  ( $=1/T_1$ ) of water



**Figure 3.** EPR as a tool to qualify oxygen-sensitive imaging approaches. Left: correlation between  $^{18}F$ -FAZA tumor accumulation, as measured by microPET, and  $pO_2$  measurements, as measured by EPR oximetry [adapted from Tran *et al.* (249)]. Right: correlation between the lipid  $R_1$ , as measured by the MOBILE technique, and  $pO_2$  measurements, as measured by EPR oximetry [adapted from Jordan *et al.* (269)].



(255,265–268). In order to counteract the lack of sensitivity of water  $T_1$  measurements (oxygen is poorly soluble in water), Jordan *et al.* exploited the high solubility of oxygen in lipids and developed a method sensitive to  $T_1$  variations in the lipid component of the tissues. This technique, called *MOBILE* (for mapping of oxygen by imaging lipids relaxation enhancement), was applied successfully to monitor tumor oxygenation and some ischemic models including cerebral stroke, peripheral ischemia and liver hypoxia. *MOBILE* was qualified by comparing the lipid  $R_1$  with quantitative oxygen measurements obtained by EPR oximetry (269) (Fig. 3, right). Manipulation of the tumor oxygenation was obtained by carbogen challenge. While no correlation was obtained between the water  $R_1$  and the  $pO_2$  values, a significant correlation was obtained between the lipid  $R_1$  and the  $pO_2$  values as measured by EPR (269).

## 6. COMBINED EPR / MRI STUDIES OF TISSUE HEMODYNAMICS AND TISSUE ENERGETICS

In addition to the qualification of new MRI protocols, complementary information may be obtained by combining EPR and MRI. In the first examples of combination of both techniques, MRI was actually simply used to localize without ambiguity the site of the EPR oxygen sensors (210,270–272). More interestingly, EPR and MRI have been used together in a multimodality approach in order to gain complementary information on the physiological status of a tissue. We have already mentioned in the description of the contrast agents the interest for this combination to map the redox status of tissues using nitroxides as CAs. One example of successful combination involved EPR oximetry with the application of gradient echo sequences in the kidneys using models of sepsis (273). Thanks to the  $pO_2$  EPR readings, the source of contrast in the kidneys was linked to the oxygenation status. The combination of EPR oximetry with perfusion sensitive MRI methods has been applied mainly to characterize the tumor microenvironment (124,214,221–226,236,239,243,244), helping to identify the different components to the evolution of the tumor hemodynamics by dissecting its components, the blood flow and the oxygen consumption. In addition to these components, a study carefully looked at the metabolic status of tumor areas by adding  $^1H$ -MRS in order to determine the lactate concentration (274). In another study, EPR oximetry and  $^{31}P$  NMR were used together to elucidate the complex link between the brain oxygenation and the cellular energetics (275).

The best example for combining MRI and EPR systems is by exploiting the dynamic nuclear polarization (DNP), also called the Overhauser effect, to detect free radicals. This process allows the signal enhancement of the signal of nuclear spins located in the closed vicinity of paramagnetic agents (276–278). In other words, information on free radicals can be obtained with a high anatomical resolution and a good sensitivity using NMR scanners (279). Overhauser magnetic resonance imaging (OMRI) or proton electron double resonance imaging (PEDRI) has been developed to map the spatial distribution of free radicals in living organisms (280,281). To achieve a DNP, the samples must be irradiated with an EPR irradiation pulse. To counter this possible heating by the irradiation at the resonance frequency of the free radicals, field-cycled PEDRI was developed (277). The magnetic field strength is decreased during the EPR pulse so that the irradiation frequency is kept relatively low (282), then the magnetic field is rapidly increased to gain in sensitivity for recording the NMR

signal. For this type of application, paramagnetic with narrow EPR linewidth (in other words, with a relatively long  $T_2e$ ) is preferred. Typically, most studies have focused on the detection of nitroxides (283) and triaryl methyl or trityl radicals (284). The method is sensitive to changes in EPR linewidths and to the interaction with proteins as shown for the nitroxides (283). Trityl radicals are less sensitive to the immobilization and their EPR linewidth vary linearly as a function of the oxygenation (284). A good example developed by the group of Krishna at the NCI was the application of OMRI to monitor tumor oxygenation coregistered with MRI anatomy (285). Another study led by Matsumoto *et al.* showed that the OMRI technology allowed the simultaneous assessment of tissue oxygenation and microvascular permeability, parameters that were calculated through the analysis of the OMRI signal enhancement profiles. The obtained information with these techniques is promising to elucidate the link between the tumor oxygenation and the vascular permeability (286). Focusing on redox imaging, it has been demonstrated that the biodistribution, the reduction and the clearance of nitroxides could be imaged in small animals using PEDRI (34,279,287,288). OMRI techniques also allow simultaneous registration of several probes that are specific for different molecular events. For instance, Utsumi *et al.* reported the simultaneous imaging of  $^{14}N$ - and  $^{15}N$ -labeled nitroxides in mice. OMRI simultaneously obtained dual images of two individual chemical processes. Oxidation and reduction were monitored in a rate-dependent manner at nanometer scale by labeling membrane-permeable and -impermeable nitroxyl radicals with  $^{14}N$  and  $^{15}N$  nuclei. OMRI offers thus great promises for *in vivo* molecular imaging of oxidative and reductive events (289).

## 7. CONCLUSION

Unlike MRI, EPR remains not well known among biomedical scientists. Possible reasons for this may be the complex EPR physics, the cost of the EPR equipments and the lack of clinical applications. Maybe more importantly, the absence of accessible educational handbooks for biomedical scientists (most books start to describe EPR with the spin Hamiltonian and very long equations) makes EPR repelling and unpopular in the medical community. Nevertheless, EPR is the most straightforward technique to assess the properties and the concentrations of paramagnetic species in biological media, in cells or in tissues. We briefly summarized in this review how EPR is important for MRI experiments. This can be explained by the fact that MRI contrast is often indirect (e.g. contrast of melanin, confounding factors associated with SPIO-induced contrast) and needs to be confirmed by alternative methods. Therefore EPR, which is a sensitive technique that assess the nature and the concentration of paramagnetic molecules, gives unambiguous information to support MRI data. Combining EPR and MRI research may have important applications in the development of new MR platforms for theranostic applications and for characterizing tissues properties, as demonstrated in the characterization of the tumor microenvironment (oxygenation, hemodynamics, redox status, pH, etc.).

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